



PhD Thesis

Amanda-Louise Fenger Carlander
Department of Otorhinolaryngology, Head and Neck Surgery & Audiology, Rigshospitalet,
Copenhagen University Hospital, Denmark

In the Era of HPV-Positive Oropharyngeal Squamous Cell Carcinoma: Patient Demographics, Prognosis, and Mesenchymal Stem Cell Therapy for Post-Treatment Side Effects

This thesis has been submitted to the Graduate School of Health and Medical Sciences, University of Copenhagen the 25th of September 2024

Academic advisors

Principal supervisor: Christian von Buchwald, MD, DMSc, Professor
Department of Otorhinolaryngology, Head and Neck Surgery & Audiology,
Rigshospitalet, Copenhagen University Hospital, Denmark

Co-supervisors: Christian Grønhøj, MD, PhD, Dr. Med., Associate professor
Department of Otorhinolaryngology, Head and Neck Surgery & Audiology,
Rigshospitalet, Copenhagen University Hospital, Denmark

Jens Kastrup, MD, PhD, DMSc, Professor Emeritus
Department of Cardiology Stem Cell Centre, The Heart Centre,
Rigshospitalet, Copenhagen University Hospital, Copenhagen

Chair: Ulrik Lassen, MD, PhD, Professor
Department of Oncology, Copenhagen, Rigshospitalet, Denmark

Assessment Committee: Antti Makitie, MD, Professor,
Department of Otorhinolaryngology - Head and Neck Surgery, University of
Helsinki and Helsinki University Hospital, Helsinki, Finland

Katarina Le Blanc, MD, PhD, Professor
Department of Laboratory Medicine, Karolinska Institutet, Stockholm, Sweden

Table of contents

List of scientific papers in the PhD	4
Acknowledgements	5
Disclosures	6
Abbreviations	7
English summary	9
Danish summary (dansk resume)	12
Introduction	15
Oropharyngeal cancer	15
Human papillomavirus and OPSCC.....	16
The HPV+ OPSCC patient	19
Diagnosis of OPSCC.....	20
Treatment and follow-up of OPSCC.....	21
Treatment-related toxicities: xerostomia and salivary gland damage.....	23
Prognostication and treatment de-escalation strategies.....	24
Screening and vaccine	25
Mesenchymal stem cells and radiation-induced xerostomia.....	26
The salivary proteome and mesenchymal stem cells.....	28
Aims and hypotheses	31
Materials and Methods	33
Summary of the key results	40
Discussion	44
Conclusion and perspectives	51
First and co-authorships in HPV, head and neck cancer, and mesenchymal stem cells	54
References	58
Appendix I – 8 th edition of the TNM AJCC/UICC staging manual for OPSCC ⁵³	71
Appendix II – Xerogenic medicine ¹⁶⁴	72
Appendix III – EORCT QLQ H&N35	74
Appendix IV – Xerostomia Questionnaire.....	76
Appendix V – Scientific papers I-IV	77

List of scientific papers in the PhD

The thesis is comprised of the following papers:

Paper I:

Carlander AF, Bendtsen SK, Rasmussen JH, Jakobsen KK, Garset-Zamani M, Grønhøj C, Friborg J, Hutcheson K, Johnson FM, Fuller CD, Moreno AC, Babarinde T, Gross ND, Myers JN, von Buchwald C. "Clinical and prognostic differences in oropharyngeal squamous cell carcinoma in USA and Denmark, two HPV high-prevalence areas." *Eur J Cancer*. 2024 May;202:113983. doi: 10.1016/j.ejca.2024.113983. Epub 2024 Mar 2. PMID: 38452723; PMCID: PMC11357839.

Paper II:

Carlander AF, Gundestrup AK, Jansson PM, Follin B, Hoeg C, Kousholt BS, Larsen RT, Jakobsen KK, Rimborg S, Fischer-Nielsen A, Grønhøj C, Buchwald CV, Lynggaard CD. "Mesenchymal Stromal/Stem Cell Therapy Improves Salivary Flow Rate in Radiation-Induced Salivary Gland Hypofunction in Preclinical in vivo Models: A Systematic Review and Meta-Analysis." *Stem Cell Rev Rep*. 2024 May;20(4):1078-1092. doi: 10.1007/s12015-024-10700-y. Epub 2024 Mar 2. PMID: 38430363; PMCID: PMC11087340.

Paper III:

Carlander AF, Jakobsen KK, Todsén T, Paaske N, Madsen AKØ, Bendtsen S, Kastrup J, Friborg J, Lynggaard CD, Hauge AW, Christensen R, Grønhøj C, von Buchwald C. "Long-term effectiveness and safety of mesenchymal stromal cell therapy for radiation-induced hyposalivation in head and neck cancer survivors: A randomised, phase-2, trial." *In review*.

Paper IV:

Carlander AF, Jakobsen KK, Jersie-Christensen R, Nielsen MK, Kastrup J, Belstrøm D, Lynggaard CD, Grønhøj C and von Buchwald C. "No changes in the salivary proteome composition detected after mesenchymal stem cell therapy for radiation-induced hyposalivation in head and neck cancer patients: A randomized, phase 2 trial." *Draft*.

Acknowledgements

I want to express my deepest gratitude to my family—Andreas, Petra, August, and Carla. Their support and encouragement have been essential throughout this journey. Without their steadfast backing, this thesis could not have been accomplished.

A special thanks to the many patients who participated in the MESRIX-III trial, without whom the study would not have been possible. I wish to thank the generous funding of the MESRIX-III trial and this thesis by the non-profit organization, Candys Foundation.

This work is a collaborative achievement, made possible by the invaluable knowledge and contributions of many dedicated colleagues. A sincere thanks to my supervisors for their guidance and encouragement. Their expertise has been essential in shaping the direction of this research. I am particularly grateful to my principal supervisor Christian von Buchwald, whose support and mentorship have been especially significant and important. His experience has greatly influenced the quality of this work and his insight and belief in me has profoundly impacted my growth as a researcher.

I wish to thank my co-supervisor Christian Grønhøj for continuous guidance throughout my research work, not only that which is included in this thesis. The MESRIX-III trial is only one part of a long journey, and I am grateful to stand on the shoulders of the previous work done by him, Charlotte Duch Lynggaard, and Kathrine Kronberg Jakobsen. I wish to thank my fellow PhD colleague Kathrine Kronberg Jakobsen, with whom I collaborated to manage the MESRIX-III trial, and who never took a break, even while I was on maternity leave.

Thank you to all the dedicated colleagues who have been invaluable for the work included in this thesis: Jacob Melchior, Tobias Todsén, Anne Østergaard Madsen, Natasja Paaske all from the Department of Otorhinolaryngology, Head and Neck Surgery & Audiology, Rigshospitalet; Jens Kastrup, Annette Ekblond, and Mandana Haack-Sørensen from the Cardiology Stem Cell Center, Rigshospitalet and Anne Werner Hauge, Department of Clinical Immunology, Rigshospitalet; Jeppe Friberg, Department of Oncology, Rigshospitalet. Their contributions to the MESRIX-III trial and Paper III were essential. A special thank you to Rosa Jersie-Christensen for your committed work on the saliva proteomic analysis and Paper IV.

Thank you to Jeffrey Myers and our colleagues at the University of Texas MD Anderson Cancer Center for a fun and educational collaboration, and not least your patience with the Danish research legislation which

almost jeopardized the foundation for our Paper I. A huge thanks to Jeppe Friborg and Jacob Rasmussen for your invaluable help with Paper I.

A heartfelt thank you to all my colleagues at the PhD office for the scientific discussions, huge amounts of coffee and moral support, especially when it was impossible to see the finish line. Finally, I wish to thank my parents, brothers, and friends for their endless support and for always being by my side.

Disclosures

Rigshospitalet, Copenhagen University Hospital and University of Copenhagen owns a patent for “Stem cell therapy for patients with salivary gland dysfunction”, PCT/EP2020/053878 for which Charlotte Lynggaard, Christian Grønhøj, and Christian von Buchwald are co-inventors. Jens Kastrup, Annette Ekblond, and Mandana Haack-Sørensen hold a patent for “Stem cell therapy based on adipose-derived stem cells”, WO2017068140.

Abbreviations

ASC	adipose-derived mesenchymal stem cell
AJCC	American Joint Committee on Cancer
ARRIVE	Animal Research: Reporting of In Vivo Experiments Guidelines
ASCO	American Society of Oncology
BSC	bone marrow-derived mesenchymal stem cell
CDK	cyclin D-cyclin dependent kinase
CRT	chemoradiotherapy
CT	computed tomography
ctHPV DNA	circulating tumor HPV DNA
DAHANCA	the Danish Head and Neck Cancer Group
DNA	deoxyribonucleic acid
DSA	donor specific antibodies
DSMO	dimethyl sulfoxide
EORCT QLQ-H&N35	European Organisation for Research and Treatment of Cancer Quality of Life Questionnaire - Head and Neck 35
EORCT HNDR	EORCT domains for dry mouth
EORCT HNSS	EORCT domains for sticky saliva
EOCRT HNSW	EORCT domains for swallowing
FEES	fiberoptic endoscopic evaluation of swallowing
G1 phase	growth 1 phase
GCP	Good Clinical Practice
Gy	gray
HLA	human leukocyte antigen
HPV	human papillomavirus
HPV+	human papillomavirus-positive
HPV-	human papillomavirus-negative
HR	hazard ratio
IHC	immunohistochemistry
IMRT	intensity modulated radiotherapy
ISH	in situ hybridization
MALT	mucosa-associated lymphoid tissue

MBS	modified barium swallowing
MeSH	medical subject headings
MDT	multidisciplinary team
MHC	major histocompatibility complex
MSC	mesenchymal stem cell
MRI	magnetic resonance imaging;
mRNA	messenger ribonucleic acid
OPSCC	oropharyngeal squamous cell carcinoma
OS	overall survival
p16	p16 ^{Ink4a} -tumor suppressor protein
p16+	p16-positive
p16-	p16-negative
PCR	polymerase chain reaction
PET-CT	positron emission tomography-computed tomography
PROMs	patient-reported outcomes measurements
QoL	quality of life
RFI	recurrence-free interval
RT	radiotherapy
SAE	serious adverse events
SFR	salivary flow rate
SMD	standardized mean difference
S phase	synthesis phase
SYRCLE	Systematic Review Center for Laboratory Animal Experimentation
SWS	stimulated salivary flow rate
The U.S.	the United States of America
TORS	transoral robotic surgery
UTMDACC	University of Texas MD Anderson Cancer Center
UWS	unstimulated salivary flow rate
XQ	xerostomia questionnaire

English summary

The incidence of oropharyngeal squamous cell carcinoma (OPSCC) has been increasing in Denmark and worldwide for the past decades, driven by infection with human papillomavirus (HPV). HPV-positive (HPV+) OPSCC is a distinct clinical and biological entity from OPSCC driven by the traditional risk factors tobacco smoking and alcohol consumption, and, since the early 2000s, a new patient group has emerged. Patients with HPV+ OPSCC are usually younger, have higher socioeconomic status, have less comorbidities, are diagnosed at lower cancer stages and, most importantly, have a better prognosis and survival than patients with HPV-negative (HPV-) OPSCC.

Most head and neck cancer patients, including OPSCC, receive radiotherapy (RT) which is related to substantial acute and late toxicities. As HPV+ OPSCC is associated with a high overall survival, there has been a pursuit to identify a subgroup of patients eligible for de-escalated therapy to reduce treatment-related side effects without compromising survival outcomes. So far, it has not been possible to reliably identify such a subgroup since still 15% of patients with HPV+ OPSCC experience a worse prognosis, with 6% developing distant metastases, most frequently in the lungs. If future de-escalation strategies are to be successful worldwide, across diverse OPSCC populations with distinct healthcare systems, we need to know to what extent different OPSCC cohorts are comparable. Although most patients with OPSCC receive RT worldwide, differences in treatment modalities exist. In Denmark, trans-oral robotic surgery (TORS) has been available since 2013 for low-stage disease (T1-T2/N0-N1) to minimize the side effects from radiation. A broader understanding of the differences in practice patterns and preferred treatment regimens is essential for establishing more uniform guidelines and, hopefully, future implementation of de-escalation strategies.

RT is related to severe long-term toxicities, and the most common side effects are salivary gland damage, hyposalivation, and xerostomia (dry mouth syndrome). Radiation-induced xerostomia and hyposalivation affect oral health; impact chewing, swallowing, and speech; and severely impact the quality of life. Currently, no disease-modifying therapies exist, and only symptomatic treatment is available. Mesenchymal stem cells (MCSs) have immunomodulatory and regenerative abilities and have been investigated as a potential therapeutic agent for radiation-induced salivary gland hypofunction and xerostomia, but the long-term effect in a larger, randomized trial has not been evaluated. The mode of action is not fully understood, but it has been indicated that the salivary proteome composition is significantly altered after MCS therapy, although not restored back to normal. However, this has not yet been validated in randomized trials.

The aim of this thesis was to investigate the interplay of patient specific factors, oncological outcomes, and rehabilitation in head and neck cancer patients in the era of HPV.

Paper I examined the clinical, treatment, and prognostic differences between two high HPV-prevalence OPSCC cohorts from the University of Texas MD Anderson Cancer Center, USA (UTMDACC) and Eastern Denmark. These cohorts represent distinct populations and healthcare systems. In Denmark, we diagnose and treat unselected cancer patients within a universal healthcare system and a focused budget, while UTMDACC, as the highest-ranked cancer center in the USA, operates within an insurance-based system, mostly available for highly-selected, resourceful patients. The study found significant demographic, clinical, and treatment differences, despite both cohorts having a high HPV prevalence. Moreover, the study revealed notable differences in the prognosis, with a higher risk of recurrence in the overall Copenhagen cohort and among advanced stages (III-IV). This highlights the necessity of understanding cohort comparability for optimizing treatment stratification in clinical trials across diverse geographical areas.

Papers II and III examined the safety and effect of transplantation with MSCs for radiation-induced salivary gland hypofunction and xerostomia. **Paper II** was a systematic review and meta-analysis investigating the effect of MSCs in animal models. The study included MSCs from all origins (adipose tissue, bone marrow, and salivary gland tissue) and all administration routes (intraglandular and systemic transplantation). MSCs was associated with a significant increase in salivary flow rate. Remodeling and regenerative effects were observed, highlighting the potential for MSCs as a therapeutic agent for hyposalivation. **Paper III** was a randomized, placebo-controlled clinical trial in humans, investigating the long-term effect of intraglandular transplantation with adipose-derived MSCs (ASCs) for radiation-induced salivary gland hypofunction in head and neck cancer survivors. The study included 120 patients, who were randomized 1:1 to receive either ASCs or placebo in both submandibular glands and were followed for 12 months. The results from the four-month follow-up have previously been published. The primary endpoint was effect of ASCs on unstimulated salivary flow rate (UWS) compared to placebo, while secondary endpoints were effect on the patient-reported outcome measurements of sticky saliva, dry mouth, swallowing, and xerostomia at 12 months. The study revealed no significant objective differences in UWS in patients receiving ASCs compared to placebo. However, there was a significant improvement in the subjective feeling of dry mouth following ASCs compared to placebo at 12 months. Both ASCs and placebo were associated with an increase in UWS, indicating a continuous natural restoration of the salivary glands. Transplantation with ASCs was safe and did not result in serious adverse events, and it was associated with a transient immune response, as most patients who developed donor-specific antibodies at four months exhibited a reduced or resolved response by 12 months. In **Paper IV** we investigated the mode of action following intraglandular

ASC therapy by analyzing the salivary proteome composition at four months in the group receiving ASC therapy compared to placebo. The study did not detect any significant differences in the salivary proteome composition following ASC therapy compared to placebo. However, a non-significant upregulation of salivary proteins upregulated in healthy salivary was seen following ASCs, suggesting a partial repair.

In conclusion, Paper I contributes to a broader understanding of the diversity between a high HPV prevalence OPSCC cohort from UTMDACC with a highly selected OPSCC population from an insurance-based healthcare system and from Eastern Denmark with an unselected OPSCC population from a universal healthcare system, which differences are important to consider for the reproducibility and validation of future de-escalation trials worldwide. Papers II-IV provide novel and translational findings on the potential of ASCs as a treatment for radiation-induced salivary gland hypofunction and xerostomia in head and neck cancer survivors. While Paper II underlines the preclinical effect of MSC therapy, Papers III-IV shows the long-term effect and mode of action through salivary proteomic profiling of intraglandular ASC therapy in a randomized, phase 2 clinical trial.

Danish summary (dansk resume)

Forekomsten af oropharynx cancer (OPSCC) er steget i Danmark og den vestlige verden over de seneste årtier, på grund af infektion med human papillomavirus (HPV). HPV-positiv (HPV+) OPSCC adskiller sig fra OPSCC forårsaget af de traditionelle risikofaktorer tobaksrygning og alkoholforbrug, og siden begyndelsen af 2000 er en ny patientgruppe derfor opstået. Patienter med HPV+ OPSCC er typisk yngre, har højere socioøkonomisk status, færre komorbiditeter, diagnosticeres i tidligere stadier af kræft og vigtigst af alt, har de en bedre overlevelse sammenlignet med patienter med HPV-negativ (HPV-) OPSCC.

De fleste patienter med hovedhalskræft, herunder OPSCC, modtager strålebehandling, som er forbundet med betydelige akutte og sene bivirkninger. Da patienter med HPV+ OPSCC har en høj overlevelse, har man forsøgt at identificere en undergruppe af patienter, der kan få deeskaleret behandling for at reducere den behandlingsrelaterede toksicitet uden at gå på kompromis med overlevelsen. Indtil videre har det ikke været muligt at identificere en sådan undergruppe, hvilket bl.a. også skyldes at 15% af patienterne med HPV+ OPSCC har en dårlig prognose og 7% får fjernmetastaser, oftest til lungerne. For at fremtidige de-eskaleringsstrategier skal være succesfulde globalt, på tværs af forskellige OPSCC-kohorter og med forskellige sundhedssystemer, er det nødvendigt at forstå i hvilken udstrækning forskellige OPSCC-populationer er sammenlignelige. Selvom de fleste patienter med hovedhalskræft får strålebehandling, er der forskelle i de anvendte behandlingsregimer. I Danmark har transoral robotkirurgi (TORS) været tilgængelig siden 2013 for patienter med lav sygdomsbyrde (T1-T2/N0-N1) for at minimere bivirkninger til strålebehandling. En bredere forståelse af forskellene i praksismønstre og foretrukne behandlingsregimer er afgørende for at etablere mere ensartede retningslinjer og forhåbentlig fremtidig implementering af de-eskaleringsstrategier.

Strålebehandling er forbundet med alvorlige og langsigtede bivirkninger, og den mest almindelige bivirkning er stråle-induceret nedsat spytkirtelfunktion, hyposalivation og tør mund, xerostomi. Stråleinduceret xerostomi og hyposalivation påvirker den orale sundhed, komplicerer tygning, synkning og tale og har en alvorlig indvirkning på patienternes livskvalitet. Aktuelt findes der ingen sygdomsmodificerende behandlingsstrategier, og de eneste tilgængelige behandlinger er symptomatiske. Mesenkymale stamceller (MSCs) har immunmodulerende og regenerative evner og er blevet undersøgt som mulig terapeutisk behandling mod stråleinduceret hyposalivation og xerostomi, men langtidseffekten er endnu ikke blevet undersøgt i større, randomiserede studier. Hvordan de virker, er endnu ikke fuldt belyst, men mindre studier tyder på at sammensætningen af spytproteomet ændrer sig markant efter

behandling med MSCs, dog ikke tilbage til et normalt og rask spytproteom. Dette er dog ikke blevet valideret i større, randomiserede studier.

Formålet med denne afhandling var at undersøge samspillet mellem patientspecifikke faktorer, prognose og rehabilitering hos patienter med hovedhalskræft i HPV-æraen.

Paper I undersøgte kliniske, behandlingsmæssige og prognostiske forskelle mellem to OPSCC-kohorter med høj HPV-prævalens fra The University of Texas MD Anderson Cancer Center, USA (UTMDACC) og Østdanmark. Disse kohorter repræsenterer forskellige populationer og sundhedssystemer: I Danmark diagnosticeres og behandles uselekerede kræftpatienter i et offentligt tilgængeligt sygehusvæsen indenfor visse budgetrammer. Modsat UTMDACC, der som det højest rangerede kræft-hospital i USA opererer i et forsikringsbaseret sundhedssystem, der sædvanligvis kun er tilgængeligt for højt selekterede, ressourcestærke patienter. Studiet fandt betydelige forskelle i demografiske, kliniske og behandlingsmæssige faktorer, på trods af at begge kohorter havde en høj HPV-prævalens. Derudover viste studiet væsentlige forskelle i prognosen, med en højere risiko for recidiv i den østdanske kohorte som helhed og for patienter med avanceret sygdom (stadie III-IV). Resultaterne fremhæver nødvendigheden af at forstå forskellige OPSCC-populationers sammenlignelighed for at optimere behandlingsstratificering i kliniske forsøg på tværs af forskellige geografiske områder.

Paper II og III undersøgte sikkerheden og effekten af transplantation med MSCs som behandling af stråleinduceret hyposalivation og xerostomi. **Paper II** var et systematisk review og meta-analyse, der undersøgte effekten af MSCs i dyremodeller. Studiet omfattede MSCs uanset oprindelse (adipøst væv, knoglemarv og spytkirtelvæv) og alle administrationsveje (intraglandulær og systemisk transplantation). Studiet viste at MSCs var associeret med en signifikant stigning i spytflowraten. Studiet viste også at behandling med MSCs var associeret med strukturelle og regenerative effekter i spytkirtlerne, hvilket understreger deres potentiale som terapeutisk behandling mod hyposalivation. **Paper III** var et humant, randomiseret, placebokontrolleret klinisk studie, der undersøgte den langsigtede effekt af intraglandulær transplantation med fedtderiverede MSCs (ASCs) mod stråleinduceret nedsat spytkirtelfunktion hos patienter med tidligere hovedhalskræft. Studiet inkluderede 120 patienter, der blev randomiseret 1:1 til at modtage enten ASCs eller placebo i begge gll. submandibularis og blev fulgt i 12 måneder. Resultaterne fra fire måneders opfølgningen er allerede udgivet. Det primære endepunkt var effekten af ASCs på ustimuleret spytflowrate (UWS) sammenlignet med placebo, mens sekundære endepunkter var effekt på patientrapporterede oplysninger vedrørende klistret spyt, tør mund, synkefunktion og xerostomi efter 12 måneder.

Studiet viste ingen signifikante objektive forskelle i UWS hos patienter, der modtog ASCs sammenlignet med placebo, men der var en signifikant forbedring af den subjektive følelse af tør mund efter ASCs sammenlignet med placebo. Behandling med både ASCs og placebo var associeret med en stigning i UWS, hvilket indikerer en kontinuerlig naturlig heling af spytkirtlerne. Behandling med ASCs var sikker og var ikke forbundet med alvorlige bivirkninger. Behandlingen var forbundet med et forbigående immunrespons, da de fleste patienter, der udviklede donor-specifikke antistoffer efter fire måneder, havde intet eller et reduceret respons efter 12 måneder. I **Paper IV** undersøgte vi mode of action for ASC-behandling sammenlignet med placebo, ved at undersøge sammensætningen af spytproteomet efter fire måneder. Studiet viste ikke nogen forskelle i sammensætningen af spytproteomet hos dem der fik ASCs sammenlignet med dem der fik placebo. Vi kunne dog vise at flere vigtige spytproteiner, der er opreguleret i rask spyt, også var opreguleret os dem, der fik ASCs sammenlignet med placebo, selvom det ikke var signifikant.

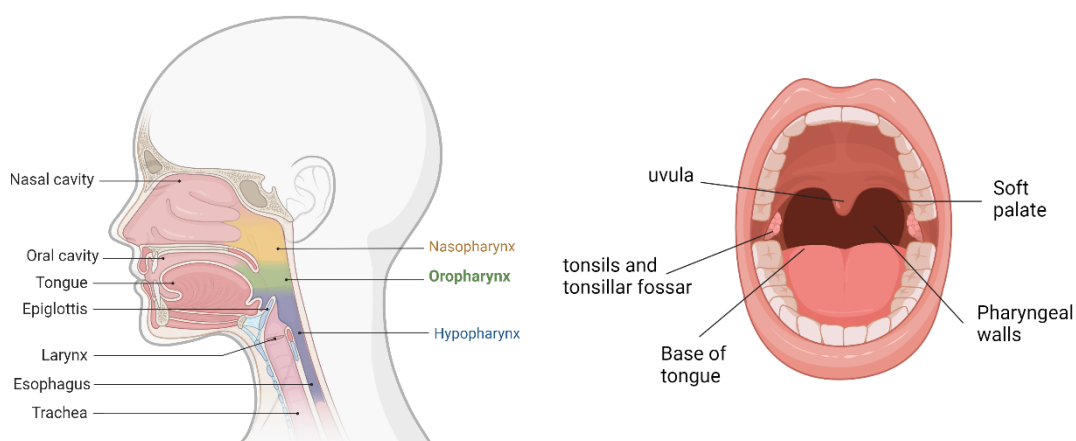
Afslutningsvis, Paper I bidrager til en bredere forståelse af forskellene mellem OPSCC-populationer med høj HPV-prævalens fra UTMDACC med en højt selekteret OPSCC-population i et forsikringsbaseret sundhedssystem og fra Østdanmark med en uselekteret OPSCC-population i et offentligt tilgængeligt sundhedssystem, som er vigtige at overveje for reproducerbarheden og validering af fremtidige deeskaleringsforsøg på tværs af geografiske områder. Paper II-IV giver nye og klinisknære resultater om potentialet for ASCs mod stråleinduceret hyposalivation og xerostomi hos patienter med tidligere hovedhalskræft. Mens Paper II understreger den prækliniske effekt af MSC-behandling, viser Paper III-IV langtidseffekten samt virkemåde ved undersøgelse af sammensætningen af spytproteomet af intraglandulær ASC-behandling i et randomiseret, fase 2 studie.

Introduction

Oropharyngeal cancer

The oropharynx is the middle part of the upper aerodigestive tract and is located posterior the mouth; see **Figure 1**. The anatomy of the oropharynx is complex, and consists of the palatine tonsils and tonsillar fossa, the base of the tongue, soft palate comprising the uvula, and the superior pharyngeal walls¹. Most often, oropharyngeal cancer arises from the mucosal tissue and the squamous cells as oropharyngeal squamous cell carcinoma (OPSCC)^{2,3}. The lingual tonsils are located in the base of the tongue with no midline raphe. The palatine and lingual tonsils consist of specialized lymphoid tissue and are part of the Waldeyer's ring. The palatine tonsils consist of 10-30 invaginated, branched crypts, while the lingual tonsils consist of only one⁴. The crypts are lined with a single layer of stratified squamous, non-keratinized epithelium with mucosa-associated lymphoid tissue (MALT) beneath⁴. The epithelium is directly exposed to the oral cavity, and lymphocytes from the underlying MALT can infiltrate the epithelial layer, creating a close interaction with oral pathogens such as human papillomavirus (HPV)⁴.

Figure 1. Anatomy of the oropharynx. The oropharynx is part of the upper aerodigestive tract bounded superiorly by the nasopharynx and inferiorly by the hypopharynx. It comprises the palatine tonsils and tonsillar fossa, the base of the tongue with the lingual tonsils, the superior pharyngeal walls, and the soft palate.



*Created with BioRender.com.

Historically, head and neck cancers, including OPSCC, have been linked to lifestyle risk factors such as tobacco smoking and alcohol consumption, but older age and being male are also associated with head and neck cancer^{5,6}.

Human papillomavirus and OPSCC

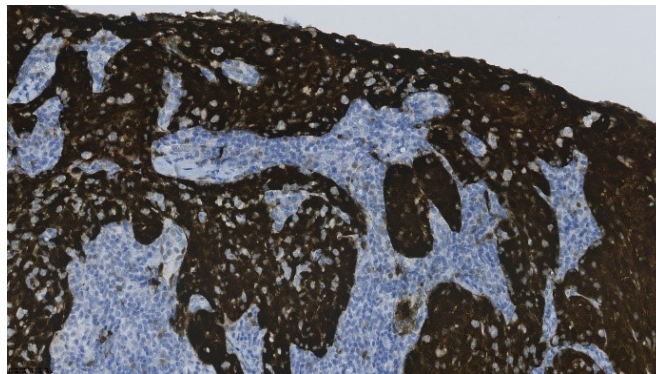
Twenty-five years ago, it became evident that HPV was associated with a distinct entity of head and neck cancers⁷⁻¹⁰. Since 2007, infection with HPV has been a well-established risk factor for OPSCC¹¹ and the incidence of HPV-positive (HPV+) OPSCC has been increasing worldwide since the early 2000s¹²⁻²⁰. A rise in the age-adjusted incidence rate pr. 100,00 from 0.9 in the year 2000 to 3.5 in 2020 has been observed in Eastern Denmark²⁰. In the same period, the female:male ratio was 1:2.6 for all OPSCC compared to 1:3.1 for HPV+ OPSCC. The burden of OPSCC among men has already surpassed the burden of cervical cancer in the U.S and UK²¹. The prevalence of HPV+ OPSCC varies across different geographical areas²², and the highest rates are observed in Northern European countries and in the U.S, but Lebanon and South Korea also have a high HPV prevalence^{16,18,19,22}. In Eastern Denmark, the HPV prevalence has increased from 46.1% from 2000-2002 to 64.3% from 2018-2020²⁰, while this is even higher in Sweden¹⁸ and the U.S.¹⁹. To prevent carcinogenic HPV infections, Denmark implemented a national vaccination program for girls in 2009, aimed at reducing HPV-driven cervical cancer. In 2019, the program was expanded to include boys to increase herd immunity, and with the expectation that it will reduce the incidence of HPV+ OPSCC in the next 30 to 40 years²³.

HPV is the most common sexually transmitted disease, with more than 200 both low-risk and high-risk genotypes²⁴. The latter are carcinogenic and responsible for HPV-driven cancers and the most common genotypes are HPV16 and HPV18^{25,26}. HPV+ OPSCC is most often caused by chronic infection with HPV16 accounting for over 80% of HPV+ OPSCCs but the high-risk genotypes HPV18, 31, 33, 35, 45, 51, and 58 are observed in relation to HPV+ OPSCC as well^{12,16,27}. Transient oral HPV infection is common in the broad population and is most often cleared within one to two years, but some will develop into a persistent infection and a few of these will develop into OPSCC. The natural history of OPSCC development spans over 30-40 years and without precancerous lesions²¹.

The gold standard HPV test method is detection of transcriptionally active messenger ribonucleic acid (mRNA) E6/E7, but this is technically demanding and expensive²⁸. Therefore, other more cost-effective and accessible techniques are used for evaluating HPV-positivity. Previously, the presence of the protein p16^{INK4a} (p16) assessed by immunohistochemistry (IHC) staining has been used as a surrogate marker for

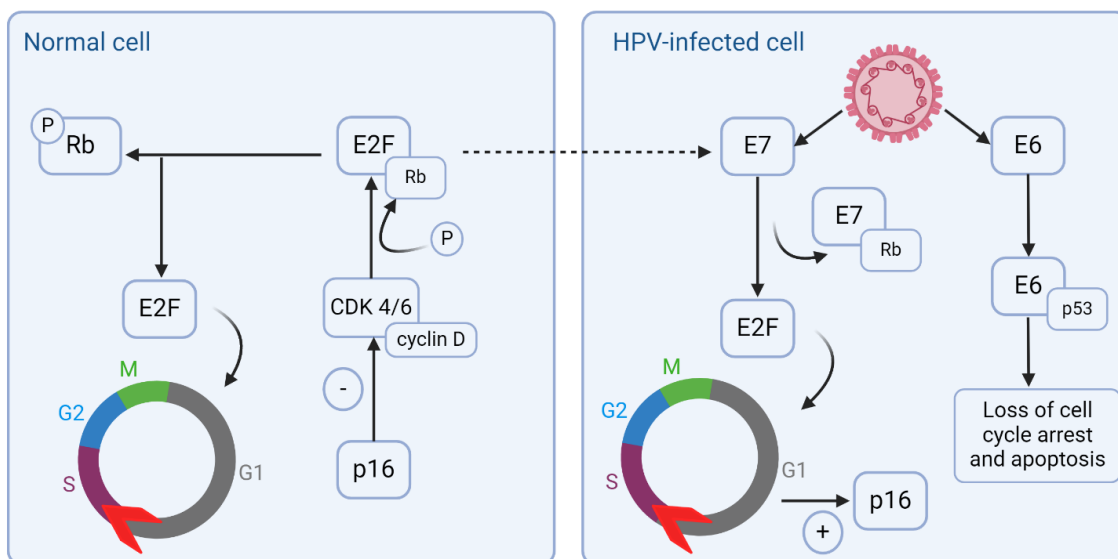
HPV positivity²⁸⁻³⁰. A cut-off of 70% nuclear and cytoplasmic staining is recommended by the American Society of Oncology (ASCO) for classifying p16-positivity; see **Figure 2**²⁸.

Figure 2. Immunohistochemistry showing p16-positivity. >70% positive nuclear and cytoplasmic immunostaining for p16 is considered positive. Picture kindly provided by the Department of Pathology, Rigshospitalet, Copenhagen.



The tumor suppressor gene p16 prevents excessive cell growth and division³¹. In normal cells, p16 inhibits cyclin D-cyclin dependent kinase (CDK) 4/6, preventing the phosphorylation of the tumor suppressor protein retinoblastoma (pRb), which in turn hinders the release of the transcription factor E2F into its active state. Consequently, the expression of downstream gene products necessary for the cell to transition from the growth 1 (G1) to the synthesis (S) phase is suppressed. Infection with HPV can lead to upregulation of p16, which is why p16 may be used as a surrogate marker for HPV infection. Integration of HPV deoxyribonucleic acid (DNA) in the host genome leads to the expression of the oncoproteins E6 and E7 which play a crucial role in HPV-induced carcinogenesis^{32,33}. E7 displays the E2F from pRb in the E2F-pRB complex, and subsequently E2F becomes active, allowing the cell to enter the S phase. Then p16 is continuously upregulated to inhibit further cell proliferation through feedback mechanisms. Thereafter, E6 binds to the tumor suppressor protein p53, which loses its regulatory function, leading to abnormal cell cycle progression, and inhibits apoptosis. See **Figure 3**.

Figure 3. Regulation pathway of p16 in normal and HPV-infected cells. HPV infection leads to the expression of the viral oncogenes E6 and E7, which inactivate the tumor suppressor proteins p53 and Rb, respectively. This inactivation results in the upregulation of p16, a surrogate biomarker often used to indicate HPV-driven carcinogenesis in OPSCC.



Abbreviations: CDK, cyclin D-cyclin dependent kinase; G, growth; HPV, human papillomavirus; M, mitosis; OPSCC, oropharyngeal squamous cell carcinoma; P, phosphorylation; p16, p16^{INK4a}, Rb, retinoblastoma; S, synthesis.

*Created with BioRender.com

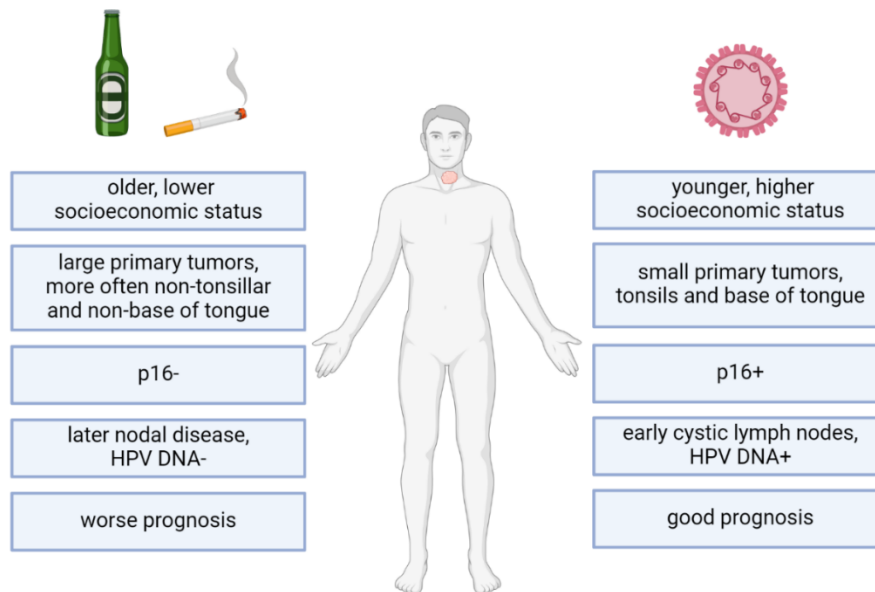
**Figure inspired by Wai et al., *Cells*, 2020³⁴.

However, the use of p16 as a stand-alone surrogate marker is insufficient, as not all HPV+ tumors show p16 overexpression, and p16 overexpression can occur in tumors not driven by HPV, which impacts the prognosis^{12,35}. Other available HPV test methods comprise detection of HPV DNA by polymerase chain reaction (PCR) or in situ hybridization (ISH) and can be used alone or in combination with p16 IHC²⁸. These tests enable detection of the presence of HPV DNA in a tumor sample, but they do not confirm whether the HPV DNA is transcriptionally active within the tumor cells. All detection methods have shown overall high sensitivity and specificity³⁶, but a combination of diagnostic test to assign HPV-status is the most attractive strategy, as double HPV/p16 status is prognostic^{12,35,37,38}.

The HPV+ OPSCC patient

HPV+ OPSCC exhibit a unique demographic, clinical, genetic, and histopathological profile compared to HPV-negative (HPV-) OPSCC caused by smoking and alcohol consumption. Patients are more often men, with limited history of smoking or alcohol consumption, higher social status, and fewer comorbidities^{12,16–18,39–44}. Patients with HPV+ OPSCC are slightly younger than patients with HPV- OPSCC (median age 59 vs. 60)¹², but a rising incidence among elderly and among women has been observed^{45,46}. HPV+ OPSCC is most often diagnosed specifically in the palatine tonsils or base of tongue, with patients frequently presenting with a cystic neck mass and small primary tumors^{12,47–49}. Importantly, patients with HPV+ OPSCC have a more favorable prognosis^{12,42}. The risk factors for acquiring an oral HPV infection are closely related to sexual behavior (e.g., lifetime number of sexual partners and age at sex debut)⁵⁰. See **Figure 4**.

Figure 4. Demographic, clinical, and prognostic differences between the HPV+ and HPV- OPSCC patient. HPV+ OPSCC patients are typically younger, of higher socioeconomic status, and present with small, p16+ tumors specifically located in the palatine tonsils or base of the tongue, often accompanied by cystic nodal metastases, and generally have a favorable overall survival.



Abbreviations: DNA, deoxyribonucleic acid; HPV, human papillomavirus; HPV-, human papilloma virus-negative; HPV+, human papilloma virus-positive; OPSCC, oropharyngeal squamous cell carcinoma; P, phosphorylation; p16, p16^{INK4a},

*Created with BioRender.com

In Denmark, patients with HPV+ OPCC have a high five-year overall survival (OS) of 79% compared to 35% in HPV- OPSCC²⁰. The long-term prognosis is favorable with 10-year OS rates of 65% for HPV+ OPSCC versus 23% for HPV- OPSCC and a low risk of late recurrences^{51,52}. To account for the distinct biology of HPV+OPSCC, which includes nodal metastases but a favorable prognosis, the latest American Joint Committee on Cancer (AJCC) and the Union for International Cancer Control (UICC) staging system (TNM 8th edition) incorporated p16-positivity to distinguish HPV+ OPSCC from HPV- OPSCC, enhancing prognostication⁵³. Currently, HPV DNA testing to confirm transcriptionally active HPV is of debate, but up to 9% of OPSCC patients have discordant p16/HPV status, with p16+/HPV+ patients having a more favorable prognosis compared to p16+/HPV- depending on geographical area and anatomical subsite¹². To further improve prognostication, combined p16 and HPV testing is important¹².

Diagnosis of OPSCC

Denmark has a universal, publicly-available healthcare system. Cancer diagnosis, treatment, and follow-up regimens are performed within cancer care packages to secure faster and standardized diagnosis and treatment initiation at public university hospitals; see **Figure 5**.^{54,55}. Patients with a suspicion of head and neck cancer should be seen within six days, diagnosed within 15 days, and treatment should be initiated within 7-11 days from diagnosis (depending on the treatment regimen given). The total process time is 28 days for surgically-treated patients and 32 days for radio- and/or chemotherapy-treated patients⁵⁴.

Figure 5. Map of Denmark. The map shows the distribution of the Danish department involved in diagnosis and treatment of head and neck cancer. Red dots represent otorhinolaryngology departments while green dots represent oncological departments.



Red flag symptoms indicative for head and neck cancer including OPSCC, and which should be referred to evaluation within the cancer fast-track program, encompass non-healing wounds or ulcers, visible or palpable tumors, and visualized neck mass but also sore throat, globus sensation, otalgia, persistent pain, or hoarseness > 2 weeks as these could indicate cancer^{54,56,57}. The evaluation is conducted at an otorhinolaryngology department in accordance with the Danish Head and Neck Cancer Group (DAHANCA) clinical guidelines⁵⁵. The evaluation comprises a thorough clinical examination and includes both fiberoptic and ultrasound examination. Depending on the patient and tumor-specific characteristics, patients will undergo imaging and a tumor biopsy. Imaging is used to evaluate the extension of the primary tumor (magnetic resonance imaging; MRI), bone involvement (computed tomography; CT) and distant metastasis (positron emission tomography-CT; PET-CT). As HPV+ OPSCC patients often present with a neck mass, fine-needle aspiration can be used to detect HPV DNA within the mass to differentiate metastases from benign disease, guiding the following diagnostic work-up⁵⁸.

A definitive diagnosis, including TNM staging, and treatment decision are determined based on a combined assessment of the biopsy, imaging, and clinical examination at a multidisciplinary team (MDT) conference with the patient, an otorhinolaryngologist, and an oncologist⁵⁴.

Treatment and follow-up of OPSCC

Three primary treatment modalities for OPSCC are used either alone or in combination: 1) surgery 2) radiotherapy (RT), and 3) chemotherapy, with chemoradiotherapy (CRT) as the standard treatment regimen⁵⁵. The clinical guidelines have not been updated since the introduction of trans-oral robotic surgery (TORS) in Denmark, resulting in the absence of TORS from the current guidelines. The guidelines are currently under revision, which are expected to be implemented in 2026. The choice of treatment depends on several factors such as tumor stage, anatomical location, performance status and comorbidities, functional considerations, and the preferences of both patients and clinicians⁵⁵. Still, most patients undergo curatively-intended, radiation-based primary treatment⁵⁹, which in Denmark consists of moderately accelerated RT administered in 33-34 fractions for six days a week with or without concurrent weekly Cisplatin (40 mg/m²) (CRT) in locoregionally advanced tumors and if tolerated by the patient⁵⁵. The standard total radiation dose equals 66-68 gray (Gy) to the target tissue, while a lower dose of 60 Gy is given to the surrounding tissue and 50 Gy to elective targets⁵⁵, minimizing radiation to the healthy, adjacent tissues. Additionally, all patients are offered nimorazole, a hypoxic radiosensitizer^{55,60}.

Historically, surgery for OPSCC involved extensive, open surgery, but since 2013 TORS has been a treatment option at Rigshospitalet. In May 2024, our department at Rigshospitalet received a new, Da Vinci single port for TORS. TORS is compared with RT as an option for OPSCC patients with low-stage (T1-T2, N0-N1) disease in the randomized clinical trial, DAHANCA34 Trial/QoLATI, (clinicaltrials.gov identification number: NCT04124198) or if primary RT cannot be performed⁶¹. The primary endpoints in the QoLATI Trial are quality of life (QoL) and swallowing function evaluated by both modified barium swallowing (MBS) and fiberoptic endoscopic evaluation of swallowing (FEES) after TORS, compared to RT, and is nearing its completion with only seven patients awaiting inclusion (+ one year follow-up). TORS is minimally invasive compared to previous open surgical techniques, and it is expected that TORS will avoid RT-related toxicities without compromising the prognosis⁶²⁻⁶⁴. Primary TORS could pave the way for salvage RT in the event of recurrent disease. However, TORS is still associated with sequelae as reported by the ORATOR trial including early-stage HPV+ OPSCC, though it should be noted that the TORS group underwent neck dissection at the time of surgery or within two weeks^{65,66}. Adverse prognostic features such as positive margins or extranodal extension following TORS require adjuvant therapy⁶⁷, and to accommodate this, the approach in Denmark is to perform a neck dissection prior to initiation of RT or TORS to minimize the need for multimodality treatment. The question of which treatment modality has a superior sequelae profile is still not fully elucidated and is under investigation in the beforementioned QoLATI trial. When the results from the QoLATI trial are available, it is expected that TORS will find its role in the new treatment guidelines addressing whether TORS is superior to RT in the treatment of T1-T2/N0-N1 OPSCCs. Even though early-stage tumors might be treated with single-modality treatment, some patients with OPSCC require multimodality treatment with increased treatment-related side effects¹⁶. Neoadjuvant chemotherapy aiming at tumor reduction may enable more patients to receive subsequent TORS⁶⁸, and a feasibility study to explore this approach will be initiated at Rigshospitalet in early 2025, comprising patients with advanced T-site (T2-T3/N0-N1) that are not immediately eligible for TORS.

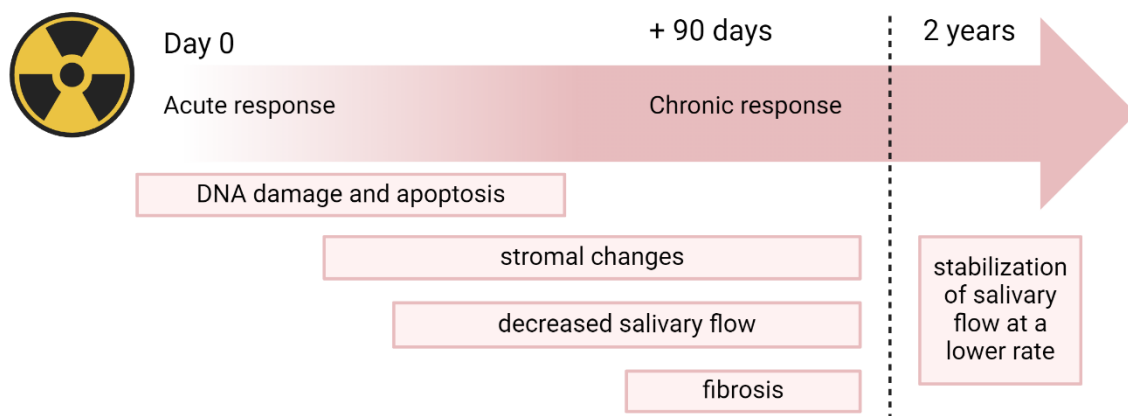
Post-treatment, the patients in Denmark are followed for five years to monitor treatment-failure, recurrence, treatment-related side effects, and rehabilitation in a standardized follow-up program⁶⁹. In Denmark, 23% of the OPSCC patients experience a recurrence, with the majority being locoregional (16%)⁷⁰. Salvage therapy (e.g., TORS following previous RT) in case of recurrence is challenging, with limited possibilities to offer curatively-intended treatment, and the survival following recurrence is very poor regardless of HPV-status⁷⁰. Early detection of recurrences is the key to improving the chances of curative treatment, but despite regular clinical examinations, the ability to detect recurrent disease is limited. Circulating tumor HPV DNA (ctHPV DNA) measurements in plasma is a promising new biomarker as it has demonstrated high sensitivity in detecting recurrence and, importantly, at Rigshospitalet it is possible to

detect the recurrence three to four months earlier than when using the otherwise standardized options^{71,72}. Consequently, a prospective, randomized trial has just been initiated at Rigshospitalet to investigate the use of ctHPV DNA as part of the follow-up regimen (Regional Scientific Ethical Committee H-23071576).

Treatment-related toxicities: xerostomia and salivary gland damage

CRT is the traditional treatment regimen for OPSCC in Denmark, and most patients in Denmark and worldwide receive radiation-based treatment⁵⁹, which severely impacts QoL and causes long-term side effects⁷³⁻⁷⁵. The salivary glands are especially radiation-sensitive and are often damaged by radiation, which promotes both acute and chronic responses characterized by inflammation, interstitial fibrosis, glandular shrinkage, loss of acinar cells, salivary gland stem cells and blood vessels leading to salivary gland hypofunction and hyposalivation, and xerostomia or “dry mouth syndrome”⁷⁶⁻⁷⁸. See **Figure 6**.

Figure 6. Radiation damage to the salivary glands over time. Radiation causes both acute and chronic damage to the salivary glands. Chronic changes include fibrosis, glandular atrophy, and persistent hyposalivation and xerostomia. These changes significantly impact the overall oral health and QoL.



Abbreviations: DNA, deoxyribonucleic acid, QoL, quality of life.

*Created with BioRender.com

**Figure inspired by Jasmer et al., *J Clin Med*, 2020⁷⁸.

Even though intensity-modulated radiotherapy (IMRT) can reduce irradiation to the parotid glands⁷⁹, xerostomia is the most frequent side effect following RT and it has been estimated to affect more than 80% of patients⁷⁷. The risk of salivary gland damage is associated with the level of delivered RT and is influenced by several treatment and patient-related factors such as the target tissue/anatomical area in question, the total radiation dose, and dose per fraction^{80,81}. A cumulative dose of 24-26 Gy to the parotid glands and 35 Gy to the submandibular glands have been recommended to allow the salivary glands to recover and thereby diminish the side effects^{81,82}.

Salivary gland function and salivation are crucial for overall oral health: they aid in food digestion and swallowing, protect the teeth and oral mucosa, and moisten the palate for articulation⁸³. Radiation-induced salivary gland hypofunction is characterized by reduced saliva production and impaired saliva composition^{84,85} and has a significant impact on the daily functioning and overall QoL among cancer survivors⁷³. Salivary gland hypofunction and hyposalivation increase the risk of oral infections and dental decay, impair chewing and swallowing with risk of malnutrition, and affect speech and sleep quality^{64,76,85}. Consequently, many patients are restricted in their daily activities and social life⁸⁶. Currently, treatment options for patients with salivary gland hypofunction are limited to symptomatic management. Consequently, there is a significant and unmet need for novel, disease-modifying therapeutic approaches for hyposalivation and xerostomi⁸⁷.

Prognostication and treatment de-escalation strategies

Standard treatment regimens are associated with a high burden of side effects and reduced QoL⁷³, and since most patients with HPV+ OPSCC have an excellent prognosis, there has been a growing interest in personalized and de-escalated treatment strategies to reduce the treatment-related toxicities without compromising the prognosis for selected patients in clinical trials^{88,89}. However, this is complicated by the fact that, despite a high survival for HPV+ OPSCC in general, a subset of patients (15%) still has a worse prognosis with both locoregional recurrences and distant metastases⁷⁰.

De-escalation strategies in clinical trials vary but encompass largely dose-reducing or replacing CRT, TORS, and induction chemotherapy. To reduce toxicity, substitution of Cisplatin with Cetuximab has been investigated as a de-escalation strategy in low-risk HPV+ OPSCC but has reduced tumor control^{90,91}. RT deintensification strategies to reduce RT-related toxicities include, among others, parotid sparing RT⁷⁹. The use of TORS as a de-escalation strategy to preserve swallowing function is of debate, as the ORATOR Trial did not demonstrate its superiority based on MD Anderson Dysphagia Inventory (MDADI) scores⁶⁶.

However, as previously noted, the TORS group underwent neck dissection at the time of surgery or within two weeks, and as a consequence, patients with adverse prognostic features received multimodality treatment. However, the results of the ongoing QoLATI study are highly anticipated as swallowing function is assessed by both MBS and FEES.

No international consensus on how to stratify HPV+ OPSCC patients in clinical trials exists, and even the definition of HPV-status differs between de-escalation studies^{88,89}. Since p16+/HPV+ patients have the best prognosis, using p16 as a stand-alone in stratification might incorrectly classify 10% of p16+ patients as HPV+^{12,92}. Other risk factors that could be considered are smoking⁹³ and oropharyngeal subsite⁵¹. Precise risk stratification is crucial if de-escalation strategies are to succeed and is important in a broader perspective if these strategies are to be implemented in diverse geographical areas with differences in demographics such as smoking prevalence and differences in p16/HPV discordance¹².

Screening and vaccine

A way to achieve widespread prevention of HPV-driven cancer, including HPV+ OPSCC, is through national vaccination programs. The quadrivalent HPV vaccination Gardasil®, covering the genotypes HPV16, -18, -6 and HPV11 was implemented in the Danish Children Vaccination Programme in 2009 to girls to prevent HPV-driven cervical cancer²³. From 2017 it was replaced with the ninevalent vaccine covering the additional genotypes HPV31, -33, -45, -52, and HPV58⁹⁴ which covers more than 95% of HPV+ OPSCCs^{16,27} and in 2019 boys were included²³. Including boys in the vaccination programs is important to mitigate the risk of HPV+ cancer, since this approach will not only protect the vaccinated boys themselves but aid vaccine coverage to reach herd immunity levels as well⁹⁵.

Due to the natural history of OPSCC spanning over 30-40 years, the effect of HPV vaccination on the risk of HPV+ OPSCC is still immature, but studies indicate that the vaccine reduces the presence of oral HPV^{96,97}. This is encouraging and supports the belief that HPV vaccination is efficient in preventing HPV+ OPSCC as well, but it will most likely not be evident in the incidence rates for the next decades⁹⁸. This underlines that the disease burden of HPV+ OPSCC will continue within the coming years at the cost of both the patients and society. It is immensely crucial to continue expanding the knowledge on the burden of HPV+ OPSCC among the public, healthcare professionals, and authorities to ensure continuous awareness, medical recognition of HPV+ OPCC, and continued political attention⁹⁹⁻¹⁰². Still, only girls are offered catch-up vaccination, while this is not available for boys¹⁰³. In addition, the incidence might be affected by vaccination pressure which may cause the virus to adapt to new vaccine-resistant strains¹⁰⁴.

Another method to prevent HPV-driven cancer is through screening. The purpose of cancer screening is to identify patients with either precancerous lesions or early-stage disease, allowing single-modality and more amenable treatment. On the other hand, late diagnosis of head and neck cancer and OPSCC are associated with higher tumor volume and more advanced stage at diagnosis^{105,106}, which is related to poorer survival and increases the necessity of multimodality treatment with increased morbidity⁴². Detecting HPV+ OPSCC as early as possible could lead to diagnosing the cancer at earlier stages with lower tumor burden, potentially improving the prognosis and reducing treatment side effects.

Cervical cytology screening has been very successful in reducing the cervical cancer incidence rates^{107,108}. However, several differences exist between cervical cancer and OPSCC driven by HPV. The development of HPV+ OPSCC is associated with oral infection with HPV, but the natural history from transient oral HPV infection to persistent infection and the development of OPSCC is poorly understood compared to cervical HPV infection and cancer²¹. Understanding this multi-step progression is further complicated by the absence of identifiable pre-cancerous lesions, and the fact that OPSCC development typically spans 30 to 40 years from initial exposure²¹. In addition, oral HPV infection tends to be more transient than cervical-genital HPV infection and are often cleared within 1-2 years¹⁰⁹, yet studies have shown that persistent infection as indicated by the presence of oral HPV antibodies or DNA is associated with OPSCC^{110,111}.

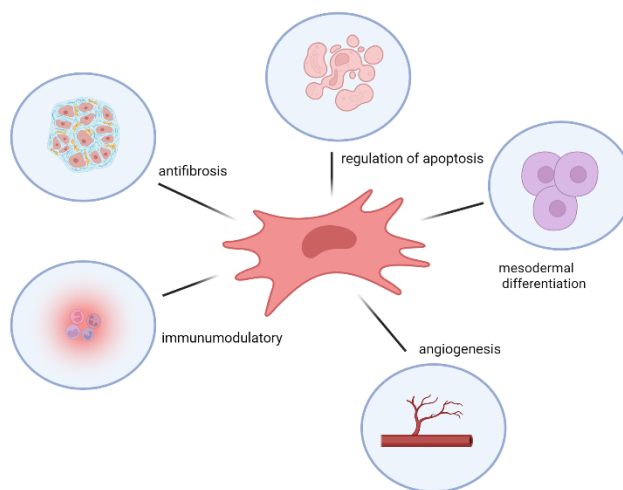
These circumstances complicate the possibilities for screening for HPV+ OPSCC. As with cervical screening, cytological screening has been investigated, but even in selected high-risk populations, cytological testing was not able to reliably screen for OPSCC¹¹². Another strategy is detection of oral HPV infection, but this has shown both low sensitivity and specificity, suggesting it would imply both many false negatives and false positives^{109,113}. The presence of sera positive HPV16 E6 antibodies is associated with the development of later HPV+ OPSCC, but they can be detected decades before the development of cancer and the far majority will not develop OPSCC^{114,115}. In conclusion, appropriate screening on a population level remains challenging, but might be more feasible in high-risk populations.

Mesenchymal stem cells and radiation-induced xerostomia

Mesenchymal stem cells (MSCs) are fibroblast-like, multipotent, adult stem cells, and were first described in the bone marrow, bone marrow-derived MSCs (BSCs), over 40 years ago by Friedenstein et al., but can reside in most connective tissue, including adipose tissue, and adipose-derived mesenchymal stem cells (ASCs)^{116,117}. ASCs have several advantages compared to BSCs: they are easily harvested by a simple liposuction in local anesthesia, contains a high concentration of MSCs, and they grow faster during ex vivo

expansion¹¹⁸. The definition of MSCs is their ability to adhere to plastic surfaces, presence (CD73, CD90, CD105) and absence (CD14, CD34, CD45) of specific surface molecules, and potential to differentiate into various cells, such as osteoblasts, chondrocytes, and adipocytes, from the mesodermal germ layer, highlighting their potential for regenerative medicine^{119,120}. Rather than playing a crucial role in engraftment¹²¹, studies have demonstrated that MSCs exhibit various microenvironmental reorganizational properties through both paracrine and trophic mechanisms; see **Figure 7**. These properties include anti-inflammatory, regulatory apoptotic, immunomodulatory, and angiogenic effects, although the exact mechanisms of action are not yet fully understood^{117,122}.

Figure 7. Mechanisms of MSCs. MSCs exert their therapeutic effect primarily through paracrine signaling and differentiation (engraftment). MSCs can give rise to mesodermal cells. Through paracrine signaling, MSCs can reduce fibrosis, promote angiogenesis, regulate apoptosis, and modulate the immune system. As a result, transplanted MSCs induce tissue regeneration and repair.



Abbreviations: MSCs, mesenchymal stem cells.

*Created with BioRender.com

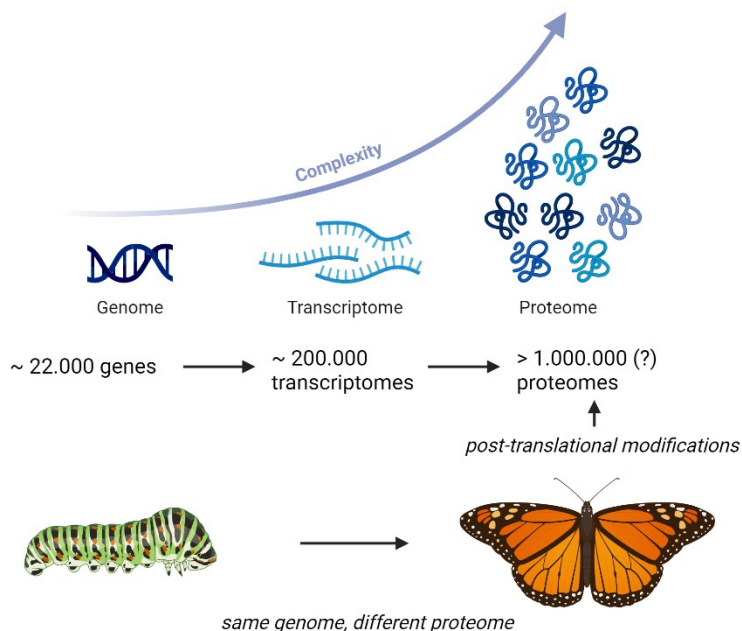
Due to their regenerative potential, MSCs have been investigated in numerous clinical trials for various disorders, such as wound healing, Crohn's disease, and fistulas, with promising results and only minimal side effects^{123–125}. MSCs are believed to be immune evasive since they express low levels of major histocompatibility complex (MHC) class I and lack expression of MHC class II^{126,127}. The use of allogeneic, immune-evasive MSCs allows a standardized, ready-to-use product, i.e., “off-the-shelf” treatment.

Our research group has completed several encouraging and original studies investigating the safety and efficacy of intraglandular ASCs treatment for radiation-induced salivary gland hypofunction after head and neck cancer to regenerate the salivary gland function and enhance the saliva production¹²⁸⁻¹³⁰. Our initial pilot study was a randomized, placebo-controlled trial with 30 patients, which assessed the safety and efficacy of autologous ASCs¹³⁰. The study showed that intraglandular treatment with ASCs was safe without serious adverse events (SAEs) and was associated with an increase in the saliva flow rate (SFR) of 33-50%¹³⁰. This was reflected in the patient-reported outcome measurements (PROMs), where patients who received ASCs experienced diminished trouble in eating both at short-term follow-up after four months and after long-term follow-up^{130,131}. But, we experienced considerable individual variability in the manufacturing of autologous ASCs, particularly in terms of cell yield, cell quality, and expansion time. To address these challenges and to avoid the need for each patient to undergo liposuction, we shifted to allogeneic ASCs. The safety of allogeneic ASCs from healthy donors as a standardized treatment was investigated in a *first-in-man* feasibility study¹²⁹. This study proved allogeneic ASCs to be safe without SAEs and with promising efficacy results with 46% increase in the saliva production and improved PROMs regarding dry mouth, sticky saliva, and swallowing¹²⁹. Moving forward, a single-center, blinded, randomized, placebo-controlled, phase II study to investigate the efficacy of treatment with ASCs for radiation-induced salivary gland hypofunction and xerostomia in previously irradiated head and neck cancer patients was initiated^{128,132}. The study included 120 patients who were randomized 1:1 to receive either ASCs or placebo in the submandibular glands. ASCs therapy significantly increased the saliva production by 38%, although the treatment was not superior to placebo after four months¹²⁸. Intraglandular BSC therapy has shown an encouraging effect on radiation-induced xerostomia as well¹³³. However, the long-term effect of intraglandular MSC therapy has not been investigated in larger, randomized, clinical trials.

The salivary proteome and mesenchymal stem cells

Proteomics are the large-scale study of proteins, including both their structure and functions¹³⁴. It encompasses the identification, quantification, and analysis of the entire protein complement in a unit: for example, in a cell, tissue, or organism¹³⁴. While genomics is relatively static, proteomics provides extensive and complex overview of the dynamic, post-translational protein landscape within a biological context; see **Figure 8**. Thus, researchers can gain insights into how proteins work together or how they change in response to various conditions or diseases. In contrast, single protein biomarkers (e.g., C-reactive protein) serve as specific indicators and provide more targeted information.

Figure 8. Proteomics and the relation to genomics. Genomics is the study of the entire DNA sequence that is transcribed into RNA. Transcriptomics measure the gene expression at the RNA level. Proteomics capture the complexity of protein expression, including post-translational modifications, offering a more functional insight.



*Created with BioRender.com

Proteomics can be investigated in different ways providing distinct information: measuring the abundance of different proteins, comparing proteomes between different samples or cohorts (conditions), structural analysis, changes in proteins' expression under a given condition, protein interaction, and functional analysis. The most used method to analyze proteomics is mass spectrometry. Briefly, this technique includes ionization and enzymatic digestion of proteins into smaller peptides, which are then separated by their mass-to-charge ratio followed by protein identification¹³⁵.

Saliva contains a diverse array of proteins reflective of the overall oral health¹³⁶. As a result, investigating the saliva proteome holds promises for e.g., early detection, monitoring disease progression, and tailoring personalized treatment strategies. The whole salivary proteome encompasses secreted proteins from all the salivary glands (parotid, submandibular, sublingual, and minor salivary glands) and therefore does not reflect proteins' alterations in specific salivary gland tissue. Studies have shown that radiation alters specific proteins in the saliva^{137–140}. These changes include alterations in the levels of various proteins such as amylase and mucins, as well as inflammatory markers associated with periodontal status and the feeling of oral dryness^{137–140}.

Treatment with ASCs may induce significant changes in the saliva proteome as a possible mechanism of action, even though the proteome after treatment with ASCs is not restored to normal¹³⁸. The changes observed following ASC include upregulated proteins involved in tissue regeneration, immune system, and cell growth¹³⁸. These findings have not yet been validated in larger, clinical randomized trials.

Aims and hypotheses

Paper I: Clinical and Prognostic Differences in Oropharyngeal Squamous Cell Carcinoma in USA and Denmark, Two HPV High-Prevalence Areas

The aim of this study is to investigate if differences exist among OPSCC patients between two high HPV-prevalence areas but with distinct populations and healthcare systems – in terms of demographics, clinical characteristics, treatment modalities, and prognosis at the UTMDACC and in Eastern Denmark. In Denmark, patients with OPSCC are diagnosed and treated within a universal healthcare system with a focused budget, representing an unselected and broad population. In contrast, the UTMDACC operates within an insurance-based system, serving a highly selected, resourceful patient population. Additionally, UTMDACC is the top-ranked cancer center in the U.S.¹⁴¹, with a more flexible budget, attracting patients from across the country. Differences in treatment guidelines exist; for example, neoadjuvant chemotherapy is available at UTMDACC but is not currently an option in Denmark.

The hypotheses are:

- 1) Differences in demographics, clinical characteristics, and treatment modalities exist between the U.S. and Eastern Denmark, despite both being high HPV-prevalence areas, associated with the disparities in the healthcare systems and treatment practice patterns.
- 2) The prognosis for OPSCC patients varies between the U.S and Eastern Denmark, despite both regions having a high HPV prevalence.

Paper II: Mesenchymal Stromal/Stem Cell Therapy Improves Salivary Flow Rate in Radiation-Induced Salivary Gland Hypofunction in Preclinical In Vivo Models: A Systematic Review and Meta-Analysis

The aim of this study is to review the current literature on the safety and effectiveness of MSCs as a therapy for radiation-induced salivary gland damage and hypofunction in animal models.

The hypotheses are:

- 1) MSC therapy for radiation-induced salivary gland damage in animal models is safe.
- 2) MSC therapy can restore the salivary gland function following radiation-damage in animal models.

Paper III: Long-term Effectiveness and Safety of Mesenchymal Stromal Cell Therapy for Radiation-induced Hyposalivation in Head and Neck Cancer Survivors: A Randomized, Placebo-controlled, Phase-2 Trial

The aim of this study is to investigate the long-term effectiveness and safety of intraglandular treatment with allogeneic ASCs in the submandibular glands to restore the salivary gland function in patients with radiation-induced salivary gland damage and hypofunction in previously irradiated head and neck cancer patient in a randomized, placebo-controlled trial. The study represents the long-term results of the previously published, primary four-month endpoint (MESRIX-III)¹²⁸.

The hypotheses are:

- 1) Intraglandular allogeneic ASC therapy can effectively restore the salivary gland function following radiation-induced salivary gland damage in head and neck cancer patients compared to placebo.
- 2) Intraglandular allogeneic ASC therapy to restore the salivary gland function following radiation-induced salivary gland damage in head and neck cancer patients is safe.

Paper IV: No Changes in the Salivary Proteome Composition Detected After Intraglandular Mesenchymal Stem Cell Therapy for Radiation-Induced Xerostomia in Previous Head and Neck Cancer Patients: A Randomized Phase 2 Trial

The aim of this study is to investigate the mode of action of intraglandular treatment with allogeneic ASCs in the submandibular glands to restore the salivary gland function in patients with radiation-induced salivary gland damage and hypofunction in previously irradiated head and neck cancer patients in a randomized, placebo-controlled trial evaluated by changes in the salivary proteome.

The hypotheses are:

- 1) Intraglandular ASC therapy induces changes in the composition of the salivary proteome in head and neck cancer patients with radiation-induced salivary gland damage and hypofunction compared to placebo.
- 2) The composition of the salivary proteome is altered in previously irradiated head and neck cancer patients compared to healthy controls.

Materials and Methods

Paper I: Clinical and Prognostic Differences in Oropharyngeal Squamous Cell Carcinoma in USA and Denmark, Two HPV High-Prevalence Areas

Study design and population

The study was a retrospective, cohort study including two distinct cohorts of patients with OPSCC from 2015-2020 in two high-HPV-prevalence areas: one population-based cohort from Eastern Denmark, and one selected cohort from the University of Texas MD Anderson Cancer Center, USA (UTMDACC).

The Copenhagen cohort was approved by The Regional Scientific Ethical Committee (H-20072877). The Houston cohort was approved by the Institutional Review Board of the UTMDACC (protocol PA 14-0947).

Outcomes

The primary endpoints of the study were to compare the demographic and clinical characteristics and treatment modalities given between the two centers. The secondary endpoints were to compare the prognosis evaluated as the three-year OS and three-year recurrence-free interval (RFI) and subgroup analyses of low-risk OPSCC patients (T1-T2/N0/M0) and high-risk patients (stage III-IV).

Statistics

Statistical analyses were performed in R Studio (version 4.1.3). Associations between variables were evaluated with either Pearson's chi-squared, Fischer's exact test, or *t*-test depending on the type of data and sample size. A *p*-value < 0.05 was considered statistically significant. OS and RFI were evaluated with Kaplan-Meier curves, the log-rank method, and by uni- and multivariable Cox regression analyses.

A comprehensive description of the material and methods can be found in Paper I.

Paper II: Mesenchymal Stromal/Stem Cell Therapy Improves Salivary Flow Rate in Radiation-Induced Salivary Gland Hypofunction in Preclinical In Vivo Models: A Systematic Review and Meta-Analysis

Study design and outcomes

This study was a systematic review and meta-analysis including preclinical in vivo models that assessed the effect of MSC therapy to restore the salivary gland function following radiation-induced salivary gland

damage. The primary endpoint was effect on SFR while secondary endpoints were effect on salivary gland morphology and histology, saliva composition, paracrine effects, and safety evaluated as adverse events. The study protocol was published and registered at PROSPERO (www.crd.ac.uk/prospero, CRD42021227336) prior initiation of this study¹⁴².

Systematic search and data extraction

The search string was created using medical subject headings (MeSH), Emtree, and text words connected to MSCs, radiation-induced salivary gland hypofunction, damage or dysfunction, and xerostomia.

Two authors (CH and ALFC) independently searched PubMed and Embase and screened studies for eligibility. Details of the preclinical in vivo model (species, sex, sample size, age), the study design (controlled, uncontrolled, randomized, blinding), irradiation details (dose, days from irradiation to MSC therapy), MSC therapy (type, concentration, administration route, follow-up time), statistical analysis, and outcomes (salivary flow rate, molecular) were extracted from each study.

Quality assessment and risk of bias

Quality assessment was performed using the Animal Research: Reporting of In Vivo Experiments (ARRIVE) guidelines¹⁴³, while risk of bias was assessed using the Systematic Review Center for Laboratory Animal Experimentation (SYRCLE) guidelines¹⁴⁴.

Data analysis

The random effect meta-analysis adjusted to Hedge's *g* was used to evaluate the efficacy of MSC therapy on salivary flow rate. A standardized mean difference (SMD) was used to assess the effect on SFR. We investigated the following subgroups: species, strain, sex, administration route, age, radiation duration, frequency of treatment, radiation dose, and time from radiation to MSC therapy.

A comprehensive description of the material and methods can be found in Paper II.

Paper III: Long-Term Effectiveness and Safety of Mesenchymal Stromal Cell Therapy for Radiation-Induced Hyposalivation in Head and Neck Cancer Survivors: A Randomized, Placebo-Controlled, Phase-2 Trial

Study design

The study was the long-term follow-up of the investigator-initiated, single-center, randomized, placebo-controlled trial investigating the efficacy and safety of allogeneic intraglandular ASCs for radiation-induced salivary gland damage and hypofunction¹²⁸. The study was initiated in January 2020 and conducted collaboratively by my fellow PhD student, Kathrine K. Jakobsen, and myself. The primary endpoint was the effect after four months and has been previously published¹²⁸. The study was approved by the National Ethics Committee (Protocol number: 1802872), The Danish Medical Agency (EudraCT: 2018-000348-24), and the Danish Data Protection Agency (Protocol number P-2020-1164). The trial was monitoring by an external Good Clinical Practice unit (GCP). The study protocol has been published¹³².

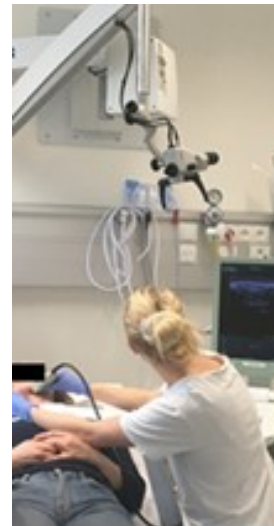
Study population

The study included 120 previously irradiated, head and neck cancer patients with radiation-induced salivary gland damage and hypofunction¹²⁸. The inclusion criteria were clinically reduced salivary flow rate, a minimum of two years since radiation therapy, age from 18-75 years, without recurrence, and informed consent. Patients were excluded if they had any cancer in the previous four years (excluding the head and neck cancer), received xerogenic medicine, had penicillin or streptomycin allergy, had prior salivary gland disease or surgery, were pregnant or breastfeeding, were currently smoking, or were abusing alcohol¹²⁸.

Interventions and study flow

Patients were randomized 1:1 to receive either placebo (CryoStor10, BiolifeSolutions, containing 10% Dimethyl sulfoxide [DSMO]) or 25 million allogeneic ASCs in each submandibular gland¹²⁸. The interventions were performed freehand guided by ultrasound, and without anesthesia; see **Figure 9**.

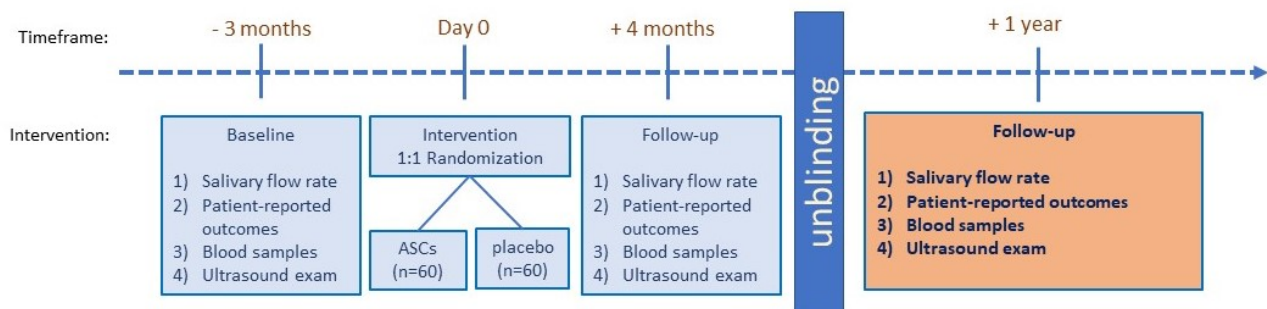
Figure 9. Intervention (with permission from the patient). The intervention was performed as an out-patient visit and done freehand guided by ultrasound, and without anesthesia. The yellow circle shows the submandibular gland while the blue line shows the needle during the injection of ASCs.



Abbreviations: ASCs, adipose-derived mesenchymal stem cells.

At baseline, the unstimulated whole salivary flow rate (UWS) and the stimulated whole salivary flow rate (SWS) were evaluated by sialometry. PROMs were evaluated with the European Organisation for Research and Treatment of Cancer Quality of Life Questionnaire - Head and Neck 35 (EORTC QLQ-H&N35), and xerostomia questionnaire (XQ). An ultrasound of their submandibular glands was made, and presence of human leukocyte antigen (HLA) antibodies evaluated in a blood sample. Effect of ASC therapy at four months was the study's primary endpoint and is previously published¹²⁸. At four months, both patients and study personnel were unblinded to the treatment, and patients were subsequently followed for a total of 12 months post-intervention. The study flow is illustrated in **Figure 10**.

Figure 10. Study flow. The study was initiated in January 2020 and conducted collaboratively by my fellow PhD student, Kathrine K. Jakobsen, and myself. The primary endpoint was the effect after four months and has been previously published (shown in blue)¹²⁸. After four months, the study became unblinded and the long-term effect was evaluated 12 months after intervention (shown in orange).



Abbreviations: ASCs, adipose-derived mesenchymal stem cells.

Outcomes

The primary endpoint of this study was the long-term effectiveness of intraglandular ASC therapy evaluated as the change in mL/min in UWS. Secondary endpoints were the long-term change in SWS and change in the PROMs XQ and EORTC QLQ-H&N35, domains HNSS (sticky saliva), HNDR (dry mouth), and HNSW (swallowing). Safety was evaluated as the development of de novo donor-specific HLA antibodies (DSA) and treatment-related SAEs.

Statistics

A statistical analysis plan was made prior to data analysis. Data was analyzed with a multilevel repeated measurements mixed effects model, with the participants as a random effect factor and the outcome variable as a dependent variable. Based on a restricted maximum likelihood model, the time was set as a fixed factor (month 0, 4, and 12). Considered significant were p-values > 0.05. A sensitivity analysis was performed for missing values using a simplistic non-responder technique with baseline values carried forward.

Paper IV: No Changes in the Salivary Proteome Composition Detected After Intraglandular Mesenchymal Stem Cell Therapy for Radiation-Induced Xerostomia in Previous Head and Neck Cancer Patients: A Randomized, Phase 2 Trial

Study design

The study was an add-on to the randomized, placebo-controlled trial investigating the efficacy and safety of allogeneic intraglandular ASCs for radiation-induced salivary gland damage and hypofunction¹²⁸. The study was approved by the National Ethics Committee (Protocol number: 1802872), The Danish Medical Agency (EudraCT: 2018-000348-24), and the Danish Data Protection Agency (Protocol number P-2020-1164). The trial was monitored by an external Good Clinical Practice unit (GCP). A study protocol to this additional study has been submitted¹⁴⁵.

Study population

As described previously, the study included 120 previously irradiated head and neck cancer patients with radiation-induced hyposalivation¹²⁸. Salivary proteomic output from 10 healthy controls was obtained from our previous study¹³⁸.

Interventions and study flow

As described previously, patients were randomized 1:1 to receive either placebo (CryoStor10, BiolifeSolutions, containing 10% DMSO) or 25 million allogeneic ASCs in each submandibular gland¹²⁸. At baseline and four months after intervention, UWS samples were collected and immediately put on dry ice and stored at -80°C.

Outcomes

The primary endpoint of this study was change in the salivary proteome composition following ASC therapy compared to placebo four months following ASC therapy. Secondary endpoints included 1) change in the salivary proteome composition following ASC therapy at baseline compared to four months, 2) evaluation of the salivary proteome composition in sub-groups associated with enhanced clinical effect (smoking, mean radiation dose to the four large salivary glands, development of donor specific antibodies and clinical

effect of 30% or more), and 3) descriptive analysis of the salivary proteome composition in irradiated patients compared to healthy controls.

Mass spectrometry analysis and protein identification

The saliva samples were analyzed using liquid chromatography-tandem mass spectrometry (LC-MS/MS). Significant differentially expressed proteins were reviewed in the Uniprot database. The salivary proteome composition in the group receiving ASC therapy was compared to the salivary proteome composition in the group receiving placebo group at four months after intervention. Results were visualized with PCA plots and volcano plots, and significance levels between the groups were evaluated with double-sided t-test. Functional annotation was evaluated in the Database of Annotation, Visualization, and Integrated Discovery (DAVID) encompassing Gene Ontology Biological Process (GO:BP) using cluster analysis.

Subgroup analyses of ASC-treated patients were chosen based on the clinical effect on increase in UWS reported at four months, with effect on UWS being associated with smoking, mean radiation dose, and development of DSAs¹²⁸. We hypothesized that a change in the salivary proteome composition would be more evident in patients with a substantial clinical effect. Hence, subgroup analysis within the ASC group included never smokers vs. ever smokers, patients who did not develop DSAs vs. patients who did not, patients who received a mean radiation dose <40 Gy vs. >40 Gy and those with a substantial clinical effect on UWS of 30% or more vs. clinical effect on UWS <30% at four months. Lastly, a comparison of the irradiated salivary proteome at baseline compared to the healthy salivary proteome composition was made, with an emphasis on Cystatin-D, -S, -SN and -SA, Glutaredoxin-1, Histatin-1, Lipocalin-1, and statherin which were significantly upregulated in healthy controls compared to irradiated patients in our previous study¹³⁸.

Summary of the key results

Paper I: Clinical and Prognostic Differences in Oropharyngeal Squamous Cell Carcinoma in USA and Denmark, Two HPV High-Prevalence Areas

The study included 2,484 patients: a large cohort of 1,216 patients with OPSCC from Eastern Denmark and a corresponding large cohort of 1,268 patients with OPSCC from UTMDACC. The Copenhagen cohort demonstrated:

- lower HPV prevalence (63% vs. 88%)
- more females (26% vs. 11%)
- older age (median age 62 years vs. 60 years).
- more ever smokers (76% vs. 47%).
- more tumors located in other OPSCC subsites than palatine tonsil and base of tongue (18% vs. 4%)
- more tumors at higher T-stages (T3-T4: 34% vs. 25%).
- more tumors with advanced nodal disease (N2-N3: 26% vs. 21%).
- more tumors at higher overall stage (III-IV: 29% vs. 16%).
- more single modality therapy (RT alone 30% vs. 15%; surgery alone 16% vs. 9%).
- worse three-year OS (85% vs 95%, log-rank $p < 0.001$), but not in the multivariable analysis, (hazard ratio [HR] 1.21, $p = 0.23$).
- worse three-year recurrence-free interval (83% vs. 91%, log-rank $p < 0.001$), and in the multivariable analysis (HR of 1.74, $p = 0.003$)

We found significant differences in clinical characteristics and treatment modalities given in subgroup analyses of low-risk (T1/T2N0M0) and high-risk (stage III-IV) patients. Significantly worse OS and RFI were found for high-risk patients, but not for low-risk patients. In low-risk patients, significantly more patients in the Copenhagen cohort received single-modality treatment.

Main findings

Despite both cohorts having a high HPV prevalence, they were significantly different regarding their demographic, clinical characteristics, and treatment modalities given, which was reflected in significant differences in the three-year prognosis.

Paper II: Mesenchymal Stromal/Stem Cell Therapy Improves Salivary Flow Rate in Radiation-Induced Salivary Gland Hypofunction in Preclinical In Vivo Models: A Systematic Review and Meta-Analysis

The systematic review included 16 studies of which 13 were included in the meta-analysis. Four studies used BSCs, 10 studies used ASCs, and two studies used salivary gland tissue-derived MSCs. Treatment with MSCs was associated with:

- no SAEs.
- increased SFR with an overall effect of 6.99 SMD, (95% CI: 2.55 to 11.42).
- improvements in acinar tissue, vascular areas, and paracrine factors.
- Reduction in apoptotic cells.
- Intraglandular injection were superior to intravenous injection.

Main findings

Treatment with MSCs in preclinical in vivo models was safe and significantly improved the salivary gland function and SFR. Additionally, MSCs induced structural and potentially regenerative changes in salivary gland tissue following radiotherapy.

Paper III: Long-term Effectiveness and Safety of Mesenchymal Stromal Cell Therapy for Radiation-Induced Hyposalivation in Head and Neck Cancer Survivors: A Randomized, Placebo-controlled, Phase-2 Trial

The baseline characteristics of the enrolled patients are previously published¹²⁸. Briefly, a total of 120 patients were included; most were males (73%) with a median duration since RT of 4.3 years (IQR 2.8 to 6.8) and had an HPV+/p16+ OPSCC (78%)¹²⁸. Of these, 117/120 (97.5%) were assessed at 12 months after intervention. The long-term effect of treatment with ASCs was associated with:

- no difference in change in UWS compared to placebo (0.007 mL/min; 95% CI [-0.02 to 0.0]3).
- a significant and clinically relevant reduction in the symptom burden for dry mouth compared to placebo (difference in change was -5.93 units; 95% CI [-10.62 to -1.22]).
- no differences in change were observed in the PROMs regarding sticky saliva, swallowing, or xerostomia compared to placebo.
- no increased risk of treatment-related SAEs compared to placebo.
- Transient immune response for those who developed donor specific antibodies.

Main findings

Treatment with intraglandular ASCs alleviated the subjective feeling of dry mouth compared to placebo, but we did not find that ASCs were superior to placebo to restore salivary gland function as evaluated by objective measurements. Treatment with ASCs was safe, without long-term SAEs.

Paper IV: No Changes in the Salivary Proteome Composition Detected After Intraglandular Mesenchymal Stem Cell Therapy for Radiation-Induced Xerostomia in Previous Head and Neck Cancer Patients: A Randomized Phase 2 Trial

All 120 enrolled patients were assessed at four months following intervention¹²⁸. Of these, 89 (74%) were included in the proteomic analyses while 30 were excluded due to inconsistent sample preparation and one patient did not produce any saliva at four months. Treatment with ASCs was associated with the following.

- No differentially expressed proteins were observed in the group receiving ASC therapy compared to placebo at four months.
- Several proteins (Cystatin-S, Cystatin-D, Cystatin-SA, Cystatin-SN, Glutaredoxin-1, and Lipocalin-1) that are upregulated in the healthy salivary proteome compared to irradiated patients were insignificantly upregulated in the ASC group compared to placebo at four months, although the levels did not reach that observed in healthy saliva.
- No changes in the salivary proteome were observed following ASC therapy at four months compared to baseline.
- Changes in the salivary proteome in the ASC group were not associated with smoking, mean radiation dose to the four large salivary glands, or clinical effect (>30% increase UWS).
- An insignificant enrichment of protein also upregulated in healthy saliva was detected in ASC-treated who did not develop DSAs compared to those who did experience an immunological response with development of DSAs.
- An increased metabolic rate and oxidative stress were observed in irradiated patients compared to healthy controls.
- In irradiated patients, important salivary proteins (cystatins and lipocalin) were upregulated in patients who had received a mean radiation dose to the four salivary glands below 40 Gy compared to more than 40 Gy, but the salivary proteome was not affected by smoking status.

Main findings

The study did not demonstrate changes in the salivary proteome composition following treatment with intraglandular ASCs compared to placebo, but treatment with ASCs did tend to induce upregulation of important salivary proteins upregulated in healthy controls. No association of smoking, mean radiation dose, or clinical effect was established on the mode of action of ASCs, but immunological tolerance tended to be associated with upregulation of salivary proteins upregulated in healthy saliva. Important functional differences were observed between irradiated patients and healthy controls, and patients who had received a dose below 40 Gy had a significant upregulation of proteins related to the oral immune system and taste perception.

Discussion

Paper I: Clinical and Prognostic Differences in Oropharyngeal Squamous Cell Carcinoma in USA and Denmark, Two HPV High-Prevalence Areas

Paper I was the largest, retrospective, cohort study investigating if clinical, treatment, and prognostic differences exist between two high HPV-prevalence OPSCC cohorts: one cohort from the highest-ranking cancer center in the U.S.¹⁴¹, UTMDACC, and one cohort from Eastern Denmark, representing not only distinct population, but distinct healthcare systems. The diverse healthcare systems might introduce a selection bias, with an unselected patient population in Eastern Denmark and a highly-selected patient population in the UTMDACC cohort. Studies have shown that insurance status is predictive of tumor stage and overall survival, and that patients without private insurance present at higher tumor stages, select treatment sites with poorer survival outcomes, and have a worse overall survival^{146,147}.

Given the current emphasis on optimizing treatment stratification for either de-escalation or escalation in clinical trials^{88,89}, accurate patient stratification is essential. If results from clinical studies are to be broadly applicable across different OPSCC cohorts, it is crucial to understand the extent to which these cohorts are comparable. Our results showed significant differences in demographic and clinical characteristics despite both cohorts having a high HPV-prevalence, both in the overall OPCC cohorts and in low- and high-risk patients potentially eligible for de-escalated or escalated treatment, respectively. This is in line with other studies¹⁴⁸, and it might be important to include multiple factors for stratification to ensure reproducibility across diverse geographical areas.

The study revealed significant differences in treatment modalities given, with single-modality RT or TORS being more prevalent in the Copenhagen cohort. This may indicate differences that influence the treatment selection not included in this study, such as comorbidities. Differences in practice patterns might play a role and have recently been reported between a Danish and a Toronto cohort¹⁴⁸. It is noteworthy that significantly more patients with low-risk OPSCC (T1-T2/N0/M0) received single-modality treatment in the Copenhagen cohort, yet no increased HR was found for both overall survival and recurrence-free interval.

The study revealed significant differences in prognosis. Although overall survival was comparable after adjusting for the demographic and clinical differences between the cohorts, the risk of recurrence remained higher in the Copenhagen cohort. This disparity may be multifaceted, potentially influenced by variations in follow-up regimens, differences in the definition and registration of recurrence, and possibly distinct biological factors, although the latter remains speculative. In the Danish universal healthcare system, where each patient is assigned a unique personal identification number, tracking and documenting disease recurrence is feasible even across different treatment sites.

The strengths of the study are that it is the largest cohort study comparing European and American high HPV-prevalence OPSCC populations from diverse healthcare systems. The study has limitations. Importantly, we were not able to use double p16/HPV-positivity in the UTMDACC cohort, which has proven to be superior in prognostication¹². Utilizing both p16 and HPV-status is recommended, especially in areas with high discordance^{12,149}. Second, we did not include socioeconomic status, performance score, or comorbidities, which are prognostic and potentially impact treatment selection. It would be interesting to investigate these factors in future studies, since we believe these vary between the two cohorts, in part driven by the differences in healthcare systems. A slightly longer follow-up time was observed in the Copenhagen cohort (2.3 versus 2.1 years, $p < 0.001$), which might impact the prognostic analysis. Although interesting differences were indicated in both low- and high-risk OPSCC patients, the sample size of especially the low-risk group was too small with too few events to make strong conclusions but were rather hypothesis generating.

Paper II: Mesenchymal Stromal/Stem Cell Therapy Improves Salivary Flow Rate in Radiation-Induced Salivary Gland Hypofunction in Preclinical In Vivo Models: A Systematic Review and Meta-Analysis

Paper II was a systematic review and meta-analysis investigating the safety and effect of MSC therapy for radiation-induced salivary gland damage and hypofunction in animal models. The study showed a significant increase in SFR following MSC treatment, which was the primary endpoint. Another interesting finding was that the effect on the SFR was associated with administration route, with intraglandular administration being more favorable than systemic transplantation. To elucidate the mode of action, most studies reported aspects of salivary gland remodeling following MSC treatment, including increased density of acinar cells, improved acinar structure, and reduced fibrosis. Upregulation of structure-related genes, epithelial markers and genes involved in cell migration, and cell survival and differentiation were also reported. In line with the reorganizational and regenerative properties of MSCs¹¹⁷, this study reported increased vascular areas and increased paracrine functioning through several growth factors contributing to cellular proliferation, angiogenesis, and neural regeneration. These findings suggest a complex mode of action underlying MSC therapy for radiation-induced salivary gland damage and hypofunction.

The strength of this study is that it comprises a substantial sample size and included a meta-analysis of the primary endpoint, SFR. Second, a protocol article was published prior to the initiation of the study¹⁴², for transparency and to reduce data-driven analysis. Lastly, the studies were quality evaluated according to both the ARRIVE guidelines¹⁴³ and the SYRCLE risk of bias tool¹⁴⁴.

The study has several limitations. Firstly, the included studies exhibited a high degree of heterogeneity with diverse methodologies. These differences encompassed variations in study design, animal models used, origin of MSCs, administration routes, radiation protocols, and follow-up regimen. Second, the RT regimens used were standardized and do not reflect the RT regimens used in head and neck cancer treatment; for example, RT was delivered as a single dose in all studies. This is important, because the results might not be translational to clinical practice. Other factors that might influence the effect of MSCs include the duration of radiation and the timing from radiotherapy to MSC therapy, both of which are challenging to translate from animal models to a clinical setting.

Paper II showed a significant therapeutic potential for MSC therapy in treating radiation-induced salivary gland damage and hypofunction, but long-term, randomized, clinical trials in humans are needed to evaluate the effect in a clinical setting.

Paper III: Long-term Effectiveness and Safety of Mesenchymal Stromal Cell Therapy for Radiation-Induced Hyposalivation in Head and Neck Cancer Survivors: A Randomized, Placebo-Controlled, Phase-2 Trial

This study represents the largest randomized trial evaluating the long-term results of the MESRIX-III clinical trial, investigating intraglandular ASCs as a treatment for radiation-induced salivary gland damage and hypofunction in head and neck cancer survivors. The primary endpoint was effect on salivary gland function, measured as UWS. Secondary aims included SWS, safety (SAEs), PROMs, and immune response (development of de novo DSAs).

The study revealed no significant difference in change in UWS between the group receiving ASCs and the group receiving placebo, with a 25% increase in the ASC group and a 27% increase in the placebo group at 12 months following intervention. This is consistent with the four-month results of the trial¹²⁸. A significant decrease in the symptom burden related to the subjective feeling of dry mouth (EORTH-H&N35 HNDR) in the ASC group compared to placebo was observed, which was not reported at four months¹²⁸. However, this is in line with the results from the MESRIX-I study, which reported significant long-term effect of ASCs in PROMs¹³¹. A decrease in the symptom burden for sticky saliva, swallowing, and xerostomia was found for ASC therapy, but with no significant differences from patients receiving placebo.

It is noteworthy that 38% (n=23) of ASC patients developed DSA, although MSCs in general are thought to be immune evasive¹²⁶. Our findings suggest that the immune response is transient, and 70% of those who developed an immune response at four months experienced a resolved or reduced response at 12 months. A temporary immune response¹⁵⁰ has also been seen in platelet transfusions¹⁵⁰. The development of DSA was

associated with a reduced effect of ASCs; however, this should be interpreted cautiously due to the small sample size of the group. DSAs pose a considerable challenge in solid organ transplantation and could be relevant for a subset of patients undergoing ASC therapy to achieve therapeutic effect¹⁵¹.

The study revealed an effect of placebo in all the included endpoints, both subjective and objective measurements. This could indicate a continuous natural restoration of the salivary glands beyond two years from RT, which is supported by others¹⁵². However, the placebo solution consisting of Cryostor10 (BiolifeSolutions) with 10% DMSO may exhibit anti-inflammatory properties acting as a therapeutic agent, as shown in other diseases¹⁵³⁻¹⁵⁵. We used ASCs from three healthy donors but observed variability in donor efficacy, with some donors producing a greater increase in UWS compared to others. Donor-variation have been reported by others¹⁵⁶, and while this remains speculative, it could be an important factor to consider in future trials.

The strengths of the study include the large sample size, as the study is the largest randomized trial investigating the long-term effect of ASCs for radiation-induced salivary gland damage and hypofunction. Still, this study was not conducted across multiple centers, which would have strengthened our findings. Further, the sample size was insufficient to thoroughly investigate subgroups, and larger, multicenter studies are needed to validate these findings comprising patient-specific factors such as development of DSAs, smoking, mean radiation dose, and time since radiation treatment. These studies are necessary to identify patients who may have a greater effect from ASCs, which this study was not powered to examine.

The study benefits from a randomized design, but it became unblinded after four months for both study personnel and participants, resulting in unblinded long-term follow-up. This is important to keep in mind when interpreting the results as this might overestimate the effect of ASCs and underestimate the effect of placebo, and might especially impact the PROMs.

Another strength is that the sialometry was performed as standardized as possible¹²⁸: 1) it was performed at the same time of the day throughout the study period to minimize daily fluctuation in SFR, 2) patients were instructed to drink a minimum of two liters the day before to minimize the risk of dehydration, and 3) patients were instructed not to eat, drink, or do oral hygiene one hour before. Conversely, the sialometry evaluations were conducted only once per follow-up period, which may not fully capture the fluctuating nature of hyposalivation, which can be influenced by other factors such as stress and can vary with the seasons and room temperature^{157,158}. Even though only validated questionnaires were used for evaluating PROMs, they might not encapsulate the entire picture, when collected at a single time point. Another limitation lies in the inherent nature of the PROMs. For example, the XQ covers both sleep disturbances, and difficulties eating and chewing. However, not all patients are affected by all three, but this will not be

evident in the sum scores. In addition, ASC therapy might have a larger impact on one domain than the others, but improvements in one domain might be invisible in the sum scores. No questions in the questionnaires are related to subjects like social eating, taste, or avoidance of certain foods or substances. Although these domains were frequently mentioned by the patients, we did not evaluate them. Only one question in the XQ was related to sleep disturbance but it was often reported by the patients. Lastly, the subjective importance of certain symptoms may vary among patients, as one domain might be more significant to one patient than to another; however, this variation may not be adequately captured by the questionnaires. It would be interesting to evaluate the QoL outcomes in a qualitative study, and maybe as patient-*defined* outcomes. A future approach could include a goal attainment scale, which is an individual scoring system to evaluate the extent of progress within a patient's individual goal as defined before the intervention. This method would account for variability in patients' symptoms, while emphasizing the aspects that are most important to each patient as well.

Paper IV: No Changes in the Salivary Proteome Composition Detected After Intraglandular Mesenchymal Stem Cell Therapy for Radiation-Induced Xerostomia in Previous Head and Neck Cancer Patients: A Randomized Phase 2 Trial

This study represents the largest randomized, placebo-controlled trial evaluating the mode of action of intraglandular ASCs as a treatment for radiation-induced salivary gland damage and hypofunction in head and neck cancer survivors. The primary endpoint was change in the salivary proteome composition following ASC therapy at four months compared to placebo, as evaluated by mass spectrometry analysis.

The study detected no changes in the salivary proteome following ASC therapy compared to placebo at four months. This is in line with the clinical results from the study, which did not establish superiority of ASC therapy over placebo in effect on UWS. However, we expected to see some changes as our pilot study, including 10 patients with early stage OPSCC (Stage I-II), revealed significant alterations following ASC therapy at four months compared to baseline, including upregulation of important salivary proteins involved in regeneration and immune defense, yet not restored to normal¹³⁸. The pilot study identified significant upregulation of salivary proteins in healthy controls compared to irradiated patient. We did observe a non-significant upregulation of most of these proteins following ASCs compared to placebo (e.g., cystatins and Lipocalin-1) indicating a trend towards a healthier saliva proteome composition following ASCs and partial repair. These proteins are, among others, important for the overall oral health and taste perception¹⁵⁹.

The increase in the clinical outcome UWS was associated with smoking, higher mean radiation dose to the four large salivary glands and development of DSAs at four months¹²⁸. Therefore, we hypothesized that an impact on the salivary proteome would be more prominent in the subgroups where we expected the highest effect of ASCs; that is among never smokers, patients who received a lower mean radiation dose (40 Gy), patients who did not develop DSAs, and those who experienced a clinical effect (>30% increase in UWS). No changes in the salivary proteome were detected among these subgroups, but a tendency towards upregulation of salivary proteins upregulated in healthy saliva was observed in those who did not develop DSAs. This indicates that immunological tolerance might be important to restore the saliva composition.

We compared the irradiated salivary proteome with healthy controls. Enriched biological terms in the irradiated proteome comprised an increased metabolic rate, reflecting the metabolic processes necessary in saliva homeostasis, but also possibly due to a higher need to repair chronic radiation damage. RT leads to oxidative stress¹⁶⁰, which was enriched in irradiated patients. Increased oxidative stress and inflammation in saliva from irradiated patients have been reported by others^{139,140}. Interestingly, the study revealed that the salivary proteome in patients who received a lower mean radiation (<40 Gy) was associated with significant upregulation of Cystatin-S, -SN and -SA and Liocalin-1, which are upregulated in healthy saliva^{138,161}. This points out the importance of minimizing radiation dose to protect the salivary glands to improve both the salivary flow rates and the salivary quality.

The study benefits from a randomized design and the large sample size. The sialometry was conducted as standardized as possible as described for Paper III, but we only collected the saliva samples once per follow-up visit, which is a limitation of our study. Other limitations include a high proportion of excluded samples, with significantly more smokers in the excluded population compared to the included population. Smoking negatively impacted the clinical effect on UWS¹²⁸, and could influence the effect on the salivary proteome composition. However, we did not observe any differences in never smokers in the ASC group at four months compared to baseline or in the irradiated patients with a history of smoking compared to never smokers.

Our study population was heterogenous, encompassing various subsites, differing radiation doses, and varying time intervals since radiation exposure. Additionally, there was significant interpersonal variability in salivary proteins levels. This reduces the likelihood of identifying small changes caused by ASC therapy. In line with this, the study was powered to identify an effect in the clinical outcomes UWS, and not changes in the salivary proteome. Lastly, we compared unstimulated salivary samples from irradiated patients with stimulated salivary samples in healthy controls, which have been shown to differ^{162,163}. For example, the

metabolic rate is higher in stimulated saliva, but our results indicated a higher metabolic rate in unstimulated saliva from irradiated patients, and this difference may be even more prominent.

Conclusion and perspectives

To advance treatment stratification and rehabilitation of head and neck cancer patients in the era of HPV, understanding the nuanced interplay of patient specific factors, oncological outcomes, and recovery is essential. This thesis seeks to address distinct but interconnected aspects of these domains, focusing on the clinical, treatment, and prognostic differences in diverse HPV+ OPSCC populations (**Paper I**), the potential of ASC therapy in regenerative medicine for radiation-induced salivary gland damage and hypofunction in animal models (**Paper II**), and the long-term effect and mode of action of intraglandular ASC therapy in alleviating radiation-induced salivary gland damage and hypofunction, and dry mouth in head and neck cancer survivors (**papers III and IV**).

In **Paper I** we investigated to what extent HPV+ OPSCC populations differ in the largest retrospective cohort study comparing European and American high HPV-prevalence OPSCC populations from diverse healthcare systems. The study comprised patients from the highest-ranked cancer center in the U.S, UTMDACC, and from Eastern Denmark, representing an insurance-based and a universal, public healthcare system, respectively. We found significant differences in demographics, clinical characteristics, and treatment regimen used. While the three-year overall survival was comparable, the Copenhagen cohort exhibited a higher risk of recurrence. These findings highlight critical clinical, therapeutic, and prognostic differences between a Northern European and a North American OPSCC population. Such variations must be taken into account when comparing outcomes and stratifying patients in clinical trials to escalated or de-escalated therapy. To advance treatment stratification in clinical trials and future clinical practice, it is essential to explore these inter-cohort differences. This will help ensure that results from clinical studies are globally applicable across diverse OPSCC populations and healthcare systems. Multicenter studies are needed to thoroughly understand these transnational differences, and between OPSCC cohorts from healthcare systems that are more alike. To investigate this further, our research group has just initiated a Nordic collaboration with centers in Sweden (Karolinska, Uppsala and Lund University Hospitals), Finland (Helsinki University Hospital), Norway (Bergen University Hospital), and Iceland (Landspítali University Hospital) all operating within universal, public healthcare systems and with unique personal identification numbers. Important strengths of such a collaboration are the long follow-up available within such a healthcare setup and the availability of double HPV/p16 status. Multiple studies are in the pipeline but comprise, among others, a similar comparison of clinical, treatment, and prognostic differences, examination of the long-term prognosis with focus on the risk of late recurrences, and the use of HPV/p16 status in other oropharyngeal subsites.

Given that the incidence of HPV+ OPSCC continues to increase²⁰ and is expected to remain high for many years to come, despite effective HPV vaccination, due to the long incubation period of HPV infections, ongoing research to optimize treatment strategies is crucial. It is essential to identify HPV+ OPSCC patients at higher risk of severe disease progression and gain a deeper understanding of the mutational profile in those who develop distant metastases. A distinct mutational profile has been found in HPV+ OPSCC with recurrent disease compared to HPV+ OPSCC without recurrent disease¹⁶⁴. Although this has not yet been validated in larger studies, mutational profiling might be useful in identifying the subgroup with aggressive, metastasizing HPV+ OPSCCs that would not benefit from de-escalated treatment. Our research group has started a consecutive, population-based study focused on the years from 2000-2020 with the aim of evaluating the mutational profile (Regional Scientific Ethical Committee H-230476672) in HPV+ OPSCCs with lung metastasis. Additionally, early detection of recurrence is important to increase the chances of curative treatment. To improve the ability to detect recurrent disease, a prospective, randomized trial investigating the use of ctHPV DNA, as part of the follow-up regimen, has been initiated (Regional Scientific Ethical Committee H-23071576), based on the promising results from a pilot study conducted at Rigshospitalet⁷². Furthermore, to enable more patients to receive TORS as the primary treatment modality, a feasibility study investigating the use of neoadjuvant chemotherapy aimed at tumor reduction prior to TORS, will be initiated in early 2025. It is expected that the study will include patients with larger primary HPV+ OPSCC tumors, T2-T3 (N0-N1), that would not otherwise be eligible for TORS. In the long run, this might pave the way for a randomized trial in a Danish setting.

In **Paper II**, we investigated the potential of MSC therapy to restore the salivary gland function following RT in animal models. The study revealed a significant effect of MSC therapy in increasing the SFR and remodeling salivary gland tissue. These findings suggest that MSC therapy has therapeutic potential for radiation-induced salivary gland damage and hypofunction. However, to validate these results and assess the efficacy in clinical practice, human randomized clinical trials are needed.

In **Paper III**, we investigated the long-term effect and safety of intraglandular ASC treatment for radiation-induced salivary gland damage and hyposalivation. The study showed that intraglandular ASC therapy significantly reduced the subjective feeling of dry mouth compared to placebo. Although both the ASC and placebo groups demonstrated increases in UWS, there was no significant objective improvement in salivary gland function with ASC therapy compared to placebo. ASC therapy may only benefit a subset of patients since effect on UWS was associated with mean radiation dose, smoking, and development of DSAs. This suggests that although intraglandular ASC therapy may alleviate the sensation of dry mouth, its impact on objectively restoring salivary gland function remains inconclusive and warrants further investigation in a

multicenter setting and powered to investigate the effect in subgroups. A more qualitative approach to evaluation of PROMs might reveal improvements in domains not fully elucidated in the questionnaires included in this study, such as sleep disturbances, while also considering the patients' unique challenges.

The previous MESRIX-I study utilized MRIs and salivary biopsies to assess the mode of action, but these methods were not incorporated in this study due to practical and ethical considerations¹³⁰. As a mode of action, The MESRIX-II study examined the salivary proteome and identified upregulation of proteins associated with regeneration¹³⁸, and this approach was incorporated in the MESRIX-III trial, **Paper IV**. The study did not detect changes in the salivary proteome composition following ASC therapy compared to placebo, although several important salivary proteins upregulated in healthy saliva were non-significantly upregulated the ASC group, indicating an incomplete restoration. Future studies on the salivary composition and salivary proteome are valuable, as ASCs may not only enhance saliva production but improve saliva quality as well.

In multiple sclerosis and osteoarthritis, repeated administrations with MSCs have shown beneficial results^{165,166}. This has not yet been investigated in radiation-induced hyposalivation and may be favorable as well. A randomized study addressing repeated treatment of patients who previously participated and received ASCs in our MESRIX-I, -II, and -III studies is expected to begin primo 2025. Additionally, ASC treatment may be beneficial in xerostomia and hyposalivation caused by inflammatory disease, such as Sjögrens Syndrome¹⁶⁷, which we will investigate in a randomized, phase 2 study scheduled to begin early 2025. Finally, intraglandular ASC therapy has not yet been validated in a large, multicenter setting, which is necessary to identify subgroups that may have an improved effect. We expect to conduct such a multicenter study in collaboration with the Karolinska University Hospital in the near future.

First and co-authorships in HPV, head and neck cancer, and mesenchymal stem cells

Papers 1-3 are included in the thesis.

1. Carlander AF, Bendtsen SK, Rasmussen JH, Jakobsen KK, Garset-Zamani M, Grønhøj C, Friberg J, Hutcheson K, Johnson FM, Fuller CD, Moreno AC, Babarinde T, Gross ND, Myers JN, von Buchwald C. Clinical and prognostic differences in oropharyngeal squamous cell carcinoma in USA and Denmark, two HPV high-prevalence areas. *Eur J Cancer*. 2024 May;202:113983. doi: 10.1016/j.ejca.2024.113983. Epub 2024 Mar 2. PMID: 38452723.
2. Carlander AF, Gundestrup AK, Jansson PM, Follin B, Hoeeg C, Kousholt BS, Larsen RT, Jakobsen KK, Rimborg S, Fischer-Nielsen A, Grønhøj C, Buchwald CV, Lynggaard CD. Mesenchymal Stromal/Stem Cell Therapy Improves Salivary Flow Rate in Radiation-Induced Salivary Gland Hypofunction in Preclinical in vivo Models: A Systematic Review and Meta-Analysis. *Stem Cell Rev Rep*. 2024 May;20(4):1078-1092. doi: 10.1007/s12015-024-10700-y. Epub 2024 Mar 2. PMID: 38430363; PMCID: PMC11087340.
3. AF Carlander, Jakobsen KK, Todsén T, Paaske N, Madsen AKØ, Bendtsen S, Kastrup J, Friberg J, Lynggaard CD, Hauge AW, Christensen R, Grønhøj C, von Buchwald C. Long-term effectiveness and safety of mesenchymal stromal cell therapy for radiation-induced hyposalivation in head and neck cancer survivors: A randomised, phase-2, trial. August 2024. *In review*.
4. Carlander AF, Jakobsen KK, Jersie-Christensen R, Hansen J, Bendtsen SK, Kastrup J, Belstrøm D, Lynggaard CD, Grønhøj C, von Buchwald C. Exploring the salivary proteome following intraglandular mesenchymal stromal cell therapy for radiation-induced hyposalivation in previous head and neck cancer patients: a secondary study protocol for the MESRIX-III, randomized, controlled trial. 2024 June. *In review*.
5. Carlander AF, Jakobsen KK, Bendtsen SK, Garset-Zamani M, Lynggaard CD, Jensen JS, Grønhøj C, Buchwald CV. A Contemporary Systematic Review on Repartition of HPV-Positivity in Oropharyngeal Cancer Worldwide. *Viruses*. 2021 Jul 9;13(7):1326. Doi: 10.3390/v13071326. PMID: 34372532; PMCID: PMC8310083.
6. Carlander AE, Grønhøj C, Hebbelstrup D, et al. Continuing rise in oropharyngeal cancer in a high HPV prevalence area: A Danish population-based study from 2011 to 2014. *Eur J Cancer*. 2017;70:75-82. Doi:10.1016/j.ejca.2016.10.015.

7. von Buchwald C, Lauritzen BB, [Carlander AF](#), Jakobsen KK, Garset-Zamani M, Bendtsen SK, Grønhøj C. Response to: Comments on “Epidemiological trends and survival of oropharyngeal cancer in a high HPV-prevalent area: A Danish population-based study from 2000 to 2020”. *Int J Cancer*. 2024 Sep 6. Doi: 10.1002/ijc.35177. Epub ahead of print. PMID: 39239857.
8. Simonsen MG, [Carlander AF](#), Jakobsen KK, Grønhøj C, von Buchwald C. The impact of the COVID-19 pandemic on time to treatment in head and neck cancer management: a systematic review. August 2024. *Submitted*.
9. Von Buchwald C, Jacobsen KK, [Carlander AF](#), Tous S, Grønhøj C, Rasmussen JH, Brooks J, Taberna M, Mena M, Morey F, Bruni L, Batis N, Brakenhoff RH, Leemans RC, Baatenburg de Jong RJ, Klussmann JP, Wuerdemann N, Wagner S, Dalianis T, Marklund L, Mirghani H, Schache A, James JA, Huang SH, O’Sullivan B, Nankivell P, Broglie MA, Hoffmann M, Quabis ES, Anderson LA, Craig SG, Alemany L, Mehanna H (HNCIG-EPIC group) . TNM 8 staging system beyond p16: double HPV/p16 status is superior to p16 alone in predicting outcome in oropharyngeal squamous cell carcinoma. *European Journal of Cancer*. September 2024. *Accepted for publication*.
10. Hastrup SV, [Carlander AF](#), Jakobsen KK, Christian Grønhøj C, von Buchwald C. The impact of COVID-19 on oropharyngeal squamous cell carcinoma patient demographics, management, and oncologic outcomes: A population based, comparative study. 2024 July. *Submitted*.
11. Lauritzen BB, Grønlund MW, Jakobsen KK, Justesen MM, Garset-Zamani M, [Carlander AF](#), Rasmussen JH, Bendtsen SK, Kiss K, Andersen G, Rosenørn MR, Friborg J, Bentzen JKD, Grønhøj C, von Buchwald C. Epidemiological trends and survival of oropharyngeal cancer in a high HPV-prevalent area: A Danish population-based study from 2000 to 2020. *Int J Cancer*. 2024 Jul 17. Doi: 10.1002/ijc.35099. Epub ahead of print. PMID: 39016028.
12. Kibsgaard CJ, Ida Salskov I, [Carlander AF](#), Kvist AS, Jakobsen KK, Grønhøj C, von Buchwald C. Development of depression in patients with oropharyngeal cancer: A systematic review. 2024 June. *Submitted*.
13. Jakobsen KK, Lynggaard CD, Paaske N, [Carlander AF](#), Kastrup J, Hauge AW, Christensen R, Grønhøj C, Buchwald CV. Long-Term Outcome Following Treatment With Allogeneic Mesenchymal Stem/Stromal Cells for Radiation-Induced Hyposalivation and Xerostomia. *Stem Cells Transl Med*. 2024 Jun 14;13(6):515-521. Doi: 10.1093/stcltm/szae017. PMID: 38578768; PMCID: PMC11165157.
14. Jakobsen KK, [Carlander AF](#), Todsén T, et al. Mesenchymal Stem/Stromal Cell Therapy for Radiation-Induced Xerostomia in Previous Head and Neck Cancer Patients: A Phase II Randomized, Placebo-Controlled Trial. *Clin Cancer Res*. 2024 May 15;30(10):2078-2084. Doi: 10.1158/1078-0432.CCR-23-3675. PMID: 38441659; PMCID:

PMC11094414.

15. Channir HI, Bendtsen SK, Melchior LC, Sandholm PR, Mordhorst C, Carlander AF, von Buchwald C, Kiss K. Validation of the VisionArray® Chip Assay for HPV DNA Testing in Histology Specimens of Oropharyngeal Squamous Cell Carcinoma. *Head Neck Pathol.* 2024 Mar 27;18(1):27. Doi: 10.1007/s12105-024-01628-3. PMID: 38536624; PMCID: PMC10973319.
16. Jakobsen KK, Carlander AF, Grønhøj C, Todsén T, Melchior J, Paaske N, Madsen AKØ, Kastrup J, Ekblond A, Haack-Sørensen M, Farhadi M, Maare C, Friberg J, Lynggaard CD, von Buchwald C. Effectiveness and safety of mesenchymal stem/stromal cell for radiation-induced hyposalivation and xerostomia in previous head and neck cancer patients (MESRIX-III): a study protocol for a single-centre, double-blinded, 56-randomized, placebo-controlled, phase II study. *Trials.* 2023 Sep 1;24(1):567. Doi: 10.1186/s13063-023-07594-5. PMID: 37658468; PMCID: PMC10474624.
17. Justesen MM, Jakobsen KK, Bendtsen SK, Garset-Zamani M, Mordhorst C, Carlander AF, Gothelf AB, Grønhøj C, von Buchwald C. Pretreatment Neutrophil-to-Lymphocyte Ratio as a Prognostic Marker for the Outcome of HPV-Positive and HPV-Negative Oropharyngeal Squamous Cell Carcinoma. *Viruses.* 2023 Jan 10;15(1):198. Doi: 10.3390/v15010198. PMID: 36680237; PMCID: PMC9863220.
18. Andersen L, Jakobsen KK, Carlander AF, Garset-Zamani M, Friberg J, Kiss K, Marvig RL, Olsen C, Nielsen FC, Andersen E, Grønhøj C, Buchwald CV. The Incidence, Survival, and HPV Impact of Second Primary Cancer following Primary Oropharyngeal Squamous Cell Carcinoma: A 20-Year Retrospective and Population-Based Study. *Viruses.* 2022 Dec 22;15(1):34. Doi: 10.3390/v15010034. PMID: 36680074; PMCID: PMC9867066.
19. Garset-Zamani M, Carlander AF, Jakobsen KK, Friberg J, Kiss K, Marvig RL, Olsen C, Nielsen FC, Andersen E, Grønhøj C, von Buchwald C. Impact of specific high-risk human papillomavirus genotypes on survival in oropharyngeal cancer. *Int J Cancer.* 2022 Apr 1;150(7):1174-1183. Doi: 10.1002/ijc.33893. Epub 2021 Dec 22. PMID: 34894151.
20. la Cour CD, Dehlendorff C, von Buchwald C, Garset-Zamani M, Grønhøj C, Carlander AF, Friis S, Kjaer SK. Non-aspirin NSAIDs and head and neck cancer mortality in a Danish nationwide cohort study. *Cancer Epidemiol.* 2022 Apr;77:102121. Doi: 10.1016/j.canep.2022.102121. Epub 2022 Feb 17. PMID: 35183905
21. Jansson PM, Lynggaard CD, Carlander AF, Jensen SB, Follin B, Hoeg C, Kousholt BS, Larsen RT, Grønhøj C, Jakobsen KK, Rimborg S, Fischer-Nielsen A, Menon JML, von Buchwald C. Mesenchymal stromal/stem cell therapy for radiation-induced salivary gland hypofunction in animal models: a protocol for a

systematic review and meta-analysis. *Syst Rev*. 2022 Apr 18;11(1):72. Doi: 10.1186/s13643-022-01943-2. PMID: 35436971; PMCID: PMC9016929

22. de la Cour CD, von Buchwald C, Dehlendorff C, Garset-Zamani M, Grønhøj C, Carlander AF, Friis S, Kjaer SK. Low-dose aspirin use and mortality risk in patients with head and neck cancer: A nationwide cohort study of 10 770 patients. *Int J Cancer*. 2021 Sep 18. Doi: 10.1002/ijc.33814. Epub ahead of print. PMID: 3453629
23. Jakobsen KK, Carlander AF, Bendtsen SK, Garset-Zamani M, Lynggaard CD, Grønhøj C, von Buchwald C. Diagnostic Accuracy of HPV Detection in Patients with Oropharyngeal Squamous Cell Carcinomas: A Systematic Review and Meta-Analysis. *Viruses*. 2021 Aug 26;13(9):1692. Doi: 10.3390/v13091692. PMID: 34578274; PMCID: PMC8473001.
24. Bendtsen SK, Jakobsen KK, Carlander AF, Grønhøj C, von Buchwald C. Focal Epithelial Hyperplasia. *Viruses*. 2021 Aug 2;13(8):1529. Doi: 10.3390/v13081529. PMID: 34452393; PMCID: PMC8402694.
25. Skovvang A, Jensen JS, Garset-Zamani M, Carlander A, Grønhøj C, von Buchwald C. The impact of HPV genotypes on survival in HPV-positive oropharyngeal squamous cell carcinomas: a systematic review. *Acta Otolaryngol*. 2021 Jul;141(7):724-728. Doi: 10.1080/00016489.2021.1927173. Epub 2021 Jun 8. PMID: 34101529.
26. Zamani M, Grønhøj C, Jensen DH, Carlander AF, Agander T, Kiss K, Olsen C, Baandrup L, Nielsen FC, Andersen E, Friberg J, von Buchwald C. The current epidemic of HPV-associated oropharyngeal cancer: An 18-year Danish population-based study with 2,169 patients. *Eur J Cancer*. 2020 Jul;134:52-59. Doi: 10.1016/j.ejca.2020.04.027. Epub 2020 May 24. PMID: 324601
27. Nami Saber C, Grønhøj C, Jensen DH, Nørregaard C, Carlander A, Garnæs E, Kiss K, Specht L, von Buchwald C. Synchronous, bilateral tonsillar carcinomas: Patients characteristics and human papillomavirus genotypes. *Oral Oncol*. 2017 Nov;74:105-110. Doi:10.1016/j.oraloncology.2017.09. Epub 2017 Oct 3.
28. Larsen C, Jensen D, Carlander AF, et al. Novel nomograms for survival and progression in HPV+ and HPV- oropharyngeal cancer: a population-based study of 1,542 consecutive patients. *Oncotarget*. 2016;7(33). Doi:10.18632/oncotarget.12335

References

1. Duvvuri U, Myers JN. Contemporary management of oropharyngeal cancer: anatomy and physiology of the oropharynx. *Curr Probl Surg*. 2009;46(2):119-184. doi:10.1067/j.cpsurg.2008.10.003
2. Kathrine Kronberg Jakobsen, Grønhøj C, Jensen DH, et al. Increasing Incidence and Survival of Head and Neck Cancers in Denmark: A nation-wide study from 1980-2014. Published online 2017.
3. Lambert R, Sauvaget C, de Camargo Cancela M, Sankaranarayanan R. Epidemiology of cancer from the oral cavity and oropharynx. *Eur J Gastroenterol Hepatol*. 2011;23(8):633-641. doi:10.1097/MEG.0b013e3283484795
4. Nave H, Gebert A, Pabst R. Morphology and immunology of the human palatine tonsil. *Anat Embryol (Berl)*. 2001;204(5):367-373. doi:10.1007/s004290100210
5. Hashibe M, Brennan P, Benhamou S, et al. Alcohol drinking in never users of tobacco, cigarette smoking in never drinkers, and the risk of head and neck cancer: pooled analysis in the International Head and Neck Cancer Epidemiology Consortium. *J Natl Cancer Inst*. 2007;99(10):777-789. doi:10.1093/jnci/djk179
6. Blot WJ, McLaughlin JK, Winn DM, et al. Smoking and drinking in relation to oral and pharyngeal cancer. *Cancer Res*. 1988;48(11):3282-3287.
7. Gillison ML, Shah K V. Human papillomavirus-associated head and neck squamous cell carcinoma: mounting evidence for an etiologic role for human papillomavirus in a subset of head and neck cancers. *Curr Opin Oncol*. 2001;13(3):183-188. doi:10.1097/00001622-200105000-00009
8. Gillison ML, Koch WM CR. Evidence for a causal association between human papillomavirus and a subset of head and neck cancers. *J Natl Cancer Inst*. 2000;92(9):709-720.
9. Gillison ML, Koch WM, Shah K V. Human papillomavirus in head and neck squamous cell carcinoma: are some head and neck cancers a sexually transmitted disease? *Curr Opin Oncol*. 1999;11(3):191-199. doi:10.1097/00001622-199905000-00010
10. Mellin H, Friesland S, Lewensohn R, Dalianis T, Munck-Wikland E. Human papillomavirus (HPV) DNA in tonsillar cancer: clinical correlates, risk of relapse, and survival. *International journal of cancer Journal international du cancer*. 2000;89(3):300-304. doi:10.1002/1097-0215(20000520)89:3<300::AID-IJC14>3.0.CO;2-G [pii]
11. IARC. *Monographs on the Evaluation of Carcinogenic Risk to Human*. 100C. Lyon: International Agency for Research on Cancer.; 2007.
12. Mehanna H, Taberna M, von Buchwald C, et al. Prognostic implications of p16 and HPV discordance in oropharyngeal cancer (HNCIG-EPIC-OPC): a multicentre, multinational, individual patient data analysis. *Lancet Oncol*. Published online 2023:239-251. doi:10.1016/S1470-2045(23)00013-X
13. Gillison ML, Chaturvedi AK, Anderson WF, Fakhry C. Epidemiology of human papillomavirus-positive head and neck squamous cell carcinoma. *Journal of Clinical Oncology*. Published online 2015. doi:10.1200/JCO.2015.61.6995

14. Chaturvedi AK, Anderson WF, Lortet-Tieulent J, et al. Worldwide Trends in Incidence Rates for Oral Cavity and Oropharyngeal Cancers. *Journal of Clinical Oncology*. 2013;31(36):4550-4559. doi:10.1200/JCO.2013.50.3870
15. Garnaes E, Kiss K, Andersen L, et al. A high and increasing HPV prevalence in tonsillar cancers in Eastern Denmark, 2000-2010: The largest registry-based study to date. *International journal of cancer Journal internationale du cancer*. 2014;00(July):1-8. doi:10.1002/ijc.29254
16. Zamani M, Grønhøj C, Jensen DH, et al. The current epidemic of HPV-associated oropharyngeal cancer: An 18-year Danish population-based study with 2,169 patients. *Eur J Cancer*. 2020;134:52-59. doi:10.1016/j.ejca.2020.04.027
17. Carlander A Louise F, Grønhøj C, Hebbelstrup D, et al. Continuing rise in oropharyngeal cancer in a high HPV prevalence area : A Danish population-based study from 2011 to 2014. *Eur J Cancer*. 2017;70:75-82. doi:10.1016/j.ejca.2016.10.015
18. Haeggeblom L, Attoff T, Yu J, et al. Changes in incidence and prevalence of human papillomavirus in tonsillar and base of tongue cancer during 2000-2016 in the Stockholm region and Sweden. *Head Neck*. 2019;41(6):1583-1590. doi:10.1002/hed.25585
19. Chaturvedi AK, Engels EA, Pfeiffer RM, et al. Human papillomavirus and rising oropharyngeal cancer incidence in the United States. *J Clin Oncol*. 2011;29(32):4294-4301. doi:10.1200/JCO.2011.36.4596
20. Lauritzen BB, Grønlund MW, Jakobsen KK, et al. Epidemiological trends and survival of oropharyngeal cancer in a high HPV-prevalent area: A Danish population-based study from 2000 to 2020. *Int J Cancer*. 2024;(August):1-2. doi:10.1002/ijc.35099
21. Lechner M, Liu J, Masterson L, Fenton TR. HPV-associated oropharyngeal cancer: epidemiology, molecular biology and clinical management. *Nat Rev Clin Oncol*. 2022;19(5):306-327. doi:10.1038/s41571-022-00603-7
22. Carlander AF, Jakobsen KK, Bendtsen SK, et al. A Contemporary Systematic Review on Repartition of HPV-Positivity in Oropharyngeal Cancer Worldwide. *Viruses*. 2021;13(7). doi:10.3390/v13071326
23. <https://en.ssi.dk/vaccination/the-danish-childhood-vaccination-programme>.
24. De Villiers EM, Fauquet C, Broker TR, Bernard HU, Zur Hausen H. Classification of papillomaviruses. *Virology*. 2004;324(1):17-27. doi:10.1016/j.virol.2004.03.033
25. de Martel C, Plummer M, Vignat J, Franceschi S. Worldwide burden of cancer attributable to HPV by site, country and HPV type. *Int J Cancer*. 2017;141(4):664-670. doi:10.1002/ijc.30716
26. Tommasino M. The human papillomavirus family and its role in carcinogenesis. *Semin Cancer Biol*. 2014;26(2014):13-21. doi:10.1016/j.semcancer.2013.11.002
27. Garset-Zamani M, Carlander AF, Jakobsen KK, et al. Impact of specific high-risk human papillomavirus genotypes on survival in oropharyngeal cancer. *Int J Cancer*. 2022;150(7):1174-1183. doi:10.1002/ijc.33893
28. Lewis JS, Beadle B, Bishop JA, et al. Human papillomavirus testing in head and neck carcinomas guideline from the college of American pathologists. *Arch Pathol Lab Med*. 2018;142(5):559-597. doi:10.5858/arpa.2017-0286-CP

29. Lewis JSJ. p16 Immunohistochemistry as a standalone test for risk stratification in oropharyngeal squamous cell carcinoma. *Head Neck Pathol.* 2012;6 Suppl 1(Suppl 1):S75-82. doi:10.1007/s12105-012-0369-0
30. Machiels JP, René Leemans C, Golusinski W, Grau C, Licitra L, Gregoire V. Squamous cell carcinoma of the oral cavity, larynx, oropharynx and hypopharynx: EHNS-ESMO-ESTRO Clinical Practice Guidelines for diagnosis, treatment and follow-up. *Ann Oncol.* 2020;31(11):1462-1475. doi:10.1016/j.annonc.2020.07.011
31. Liggett WHJ, Sidransky D. Role of the p16 tumor suppressor gene in cancer. *J Clin Oncol.* 1998;16(3):1197-1206. doi:10.1200/JCO.1998.16.3.1197
32. Moody CA, Laimins LA. Human papillomavirus oncoproteins: pathways to transformation. *Nat Rev Cancer.* 2010;10(8):550-560. doi:10.1038/nrc2886
33. Comprehensive genomic characterization of head and neck squamous cell carcinomas. *Nature.* 2015;517(7536):576-582. doi:doi:10.1038/nature14129
34. Wai K, Strohl M, Zante A, Ha P. Molecular Diagnostics in Human Papillomavirus-Related Head and Neck Squamous Cell Carcinoma. *Cells.* 2020;9:500. doi:10.3390/cells9020500
35. von Buchwald C, Jakobsen KK, Carlander ALF, et al. TNM 8 staging system beyond p16: Double HPV/p16 status is superior to p16 alone in predicting outcome in oropharyngeal squamous cell carcinoma. *Eur J Cancer.* 2024;Accepted.
36. Jakobsen KK, Carlander ALF, Bendtsen SK, et al. Diagnostic Accuracy of HPV Detection in Patients with Oropharyngeal Squamous Cell Carcinomas: A Systematic Review and Meta-Analysis. *Viruses.* 2021;13(9). doi:10.3390/v13091692
37. Prigge ES, Arbyn M, von Knebel Doeberitz M, Reuschenbach M. Diagnostic accuracy of p16(INK4a) immunohistochemistry in oropharyngeal squamous cell carcinomas: A systematic review and meta-analysis. *Int J Cancer.* 2017;140(5):1186-1198. doi:10.1002/ijc.30516
38. Craig SG, Anderson LA, Schache AG, et al. Recommendations for determining HPV status in patients with oropharyngeal cancers under TNM8 guidelines: a two-tier approach. *Br J Cancer.* 2019;120(8):827-833. doi:10.1038/s41416-019-0414-9
39. Hayes DN, Van Waes C, Seiwert TY. Genetic Landscape of Human Papillomavirus-Associated Head and Neck Cancer and Comparison to Tobacco-Related Tumors. *J Clin Oncol.* 2015;33(29):3227-3234. doi:10.1200/JCO.2015.62.1086
40. Lewis JS, Khan RA, Masand RP, et al. Recognition of nonkeratinizing morphology in oropharyngeal squamous cell carcinoma - a prospective cohort and interobserver variability study. *Histopathology.* 2012;60(3):427-436. doi:10.1111/j.1365-2559.2011.04092.x
41. Reder H, Wagner S, Wuerdemann N, et al. Mutation patterns in recurrent and/or metastatic oropharyngeal squamous cell carcinomas in relation to human papillomavirus status. *Cancer Med.* 2021;10(4):1347-1356. doi:10.1002/cam4.3741
42. Ang KK, Harris J, Wheeler R, et al. Human papillomavirus and survival of patients with oropharyngeal cancer. *N Engl J Med.* 2010;363(1):24-35. doi:10.1056/NEJMoa0912217

43. Gillison ML, D'Souza G, Westra W, et al. Distinct risk factor profiles for human papillomavirus type 16-positive and human papillomavirus type 16-negative head and neck cancers. *J Natl Cancer Inst.* 2008;100(6):407-420. doi:10.1093/jnci/djn025
44. Eytan DF, Blackford AL, Eisele DW, Fakhry C. Prevalence of comorbidities and effect on survival in survivors of human papillomavirus-related and human papillomavirus-unrelated head and neck cancer in the United States. *Cancer.* 2019;125(2):249-260. doi:10.1002/cncr.31800
45. D'Souza G, Westra WH, Wang SJ, et al. Differences in the Prevalence of Human Papillomavirus (HPV) in Head and Neck Squamous Cell Cancers by Sex, Race, Anatomic Tumor Site, and HPV Detection Method. *JAMA Oncol.* 2017;3(2):169-177. doi:10.1001/jamaoncol.2016.3067
46. Windon MJ, D'Souza G, Rettig EM, et al. Increasing prevalence of human papillomavirus-positive oropharyngeal cancers among older adults. *Cancer.* 2018;124(14):2993-2999. doi:10.1002/cncr.31385
47. Khalid MB, Ting P, Pai A, et al. Initial presentation of human papillomavirus-related head and neck cancer: A retrospective review. *Laryngoscope.* 2019;129(4):877-882. doi:10.1002/lary.27296
48. Goldenberg D, Begum S, Westra W, et al. Cystic lymph node metastasis in patients with head and neck cancer: an HPV-associated phenomenon. *Head Neck.* 2008;30(7):898-903. doi:10.1002/hed
49. Truong Lam M, O'Sullivan B, Gullane P, Huang SH. Challenges in establishing the diagnosis of human papillomavirus-related oropharyngeal carcinoma. *Laryngoscope.* 2016;126(10):2270-2275. doi:10.1002/lary.25985
50. D'Souza G, Agrawal Y, Halpern J, Bodison S, Gillison ML. Oral sexual behaviors associated with prevalent oral human papillomavirus infection. *J Infect Dis.* 2009;199(9):1263-1269. doi:10.1086/597755
51. Wendt M, Hammarstedt-Nordenvall L, Zupancic M, et al. Long-Term Survival and Recurrence in Oropharyngeal Squamous Cell Carcinoma in Relation to Subsites, HPV, and p16-Status. *Cancers (Basel).* 2021;13(11). doi:10.3390/cancers13112553
52. Autio TJ, Atula T, Jouhi L, et al. Timing and frequency of oropharyngeal squamous cell carcinoma recurrences after treatment with curative intent. *Acta Otolaryngol.* 2023;143(4):328-333. doi:10.1080/00016489.2023.2188892
53. O'Sullivan B, Huang SH, Su J, et al. Development and validation of a staging system for HPV-related oropharyngeal cancer by the International Collaboration on Oropharyngeal cancer Network for Staging (ICON-S): a multicentre cohort study. *Lancet Oncol.* 2016;17(4):440-451. doi:10.1016/S1470-2045(15)00560-4
54. Sundhedsstyrelsen. *Pakkeforløb for Hoved-Og Halskræft For Fagfolk.*; 2020. www.sst.dk
55. Dansk Selskab for Hoved- og Hals Onkologi (DSHHO) DDHHC (DAHANCA). Nationale Retningslinjer for Udredning, Behandling, Rehabilitering og Kontrolforløb for Patienter med Pharynx- og Larynx-cancer i Danmark. 2011;(april):1-69.
56. McIlwain WR, Sood AJ, Nguyen SA, Day TA. Initial symptoms in patients with HPV-positive and HPV-negative oropharyngeal cancer. *JAMA Otolaryngol Head Neck Surg.* 2014;140(5):441-447. doi:10.1001/jamaoto.2014.141

57. Carpén T, Sjöblom A, Lundberg M, et al. Presenting symptoms and clinical findings in HPV-positive and HPV-negative oropharyngeal cancer patients. *Acta Otolaryngol.* 2018;138(5):513-518. doi:10.1080/00016489.2017.1405279
58. Channir HI, Grønhøj Larsen C, Ahlborn LB, et al. Validation study of HPV DNA detection from stained FNA smears by polymerase chain reaction: Improving the diagnostic workup of patients with a tumor on the neck. *Cancer Cytopathol.* 2016;124(11):820-827. doi:10.1002/cncy.21753
59. Borrás JM, Barton M, Grau C, et al. The impact of cancer incidence and stage on optimal utilization of radiotherapy: Methodology of a population based analysis by the ESTRO-HERO project. *Radiotherapy and Oncology.* 2015;116(1):45-50. doi:10.1016/j.radonc.2015.04.021
60. Overgaard J, Hansen HS, Overgaard M, et al. A randomized double-blind phase III study of nimorazole as a hypoxic radiosensitizer of primary radiotherapy in supraglottic larynx and pharynx carcinoma. Results of the Danish Head and Neck Cancer Study (DAHANCA) Protocol 5-85. *Radiother Oncol.* 1998;46(2):135-146. doi:10.1016/s0167-8140(97)00220-x
61. Rubek N, Channir HI, Charabi BW, et al. Primary transoral robotic surgery with concurrent neck dissection for early stage oropharyngeal squamous cell carcinoma implemented at a Danish head and neck cancer center: a phase II trial on feasibility and tumour margin status. *Eur Arch Otorhinolaryngol.* 2017;274(5):2229-2237. doi:10.1007/s00405-016-4433-3
62. Meldgaard Justesen M, Kronberg Jakobsen K, Fenger Carlander AL, et al. Outcomes of transoral robotic surgery for early-stage oropharyngeal squamous cell carcinoma with low rates of adjuvant therapy: A consecutive single-institution study from 2013 to 2020. *Oral Oncol.* 2024;152:106783. doi:10.1016/j.oraloncology.2024.106783
63. Ling DC, Chapman B V, Kim J, et al. Oncologic outcomes and patient-reported quality of life in patients with oropharyngeal squamous cell carcinoma treated with definitive transoral robotic surgery versus definitive chemoradiation. *Oral Oncol.* 2016;61:41-46. doi:10.1016/j.oraloncology.2016.08.004
64. Scott SI, Kathrine Ø Madsen A, Rubek N, et al. Long-term quality of life & functional outcomes after treatment of oropharyngeal cancer. *Cancer Med.* 2021;10(2):483-495. doi:10.1002/cam4.3599
65. Nichols AC, Theurer J, Prisman E, et al. Randomized Trial of Radiotherapy Versus Transoral Robotic Surgery for Oropharyngeal Squamous Cell Carcinoma: Long-Term Results of the ORATOR Trial. *J Clin Oncol.* 2022;40(8):866-875. doi:10.1200/JCO.21.01961
66. Palma DA, Prisman E, Berthelet E, et al. Assessment of Toxic Effects and Survival in Treatment Deescalation With Radiotherapy vs Transoral Surgery for HPV-Associated Oropharyngeal Squamous Cell Carcinoma: The ORATOR2 Phase 2 Randomized Clinical Trial. *JAMA Oncol.* 2022;8(6):1-7. doi:10.1001/jamaoncol.2022.0615
67. Sher DJ, Adelstein DJ, Bajaj GK, et al. Radiation therapy for oropharyngeal squamous cell carcinoma: Executive summary of an ASTRO Evidence-Based Clinical Practice Guideline. *Pract Radiat Oncol.* 2017;7(4):246-253. doi:10.1016/j.prro.2017.02.002
68. Costantino A, Sampieri C, De Virgilio A, Kim SH. Neo-adjuvant chemotherapy and transoral robotic surgery in locoregionally advanced oropharyngeal cancer. *Eur J Surg Oncol.* 2023;49(12):107121. doi:10.1016/j.ejso.2023.107121

69. Sundhedsstyrelsen. *Opfølgingsprogram for Hoved- Og Hals- Kræft (Follow-up Programme for Head and Neck Cancer)*.; 2015.
70. Grønhøj C, Jakobsen KK, Jensen DH, et al. Pattern of and survival following loco-regional and distant recurrence in patients with HPV+ and HPV- oropharyngeal squamous cell carcinoma: A population-based study. *Oral Oncol*. 2018;83(March 2018):127-133. doi:10.1016/j.oraloncology.2018.06.012
71. Chera BS, Kumar S, Shen C, et al. Plasma Circulating Tumor HPV DNA for the Surveillance of Cancer Recurrence in HPV-Associated Oropharyngeal Cancer. Published online 2020. doi:10.1200/JCO.19
72. Jakobsen KK, Bendtsen SK, Pallisgaard N, et al. Liquid Biopsies with Circulating Plasma HPV-DNA Measurements-A Clinically Applicable Surveillance Tool for Patients with HPV-Positive Oropharyngeal Cancer. *Clin Cancer Res*. 2023;29(19):3914-3923. doi:10.1158/1078-0432.CCR-23-1064
73. Høxbroe Michaelsen S, Grønhøj C, Høxbroe Michaelsen J, Friberg J, von Buchwald C. Quality of life in survivors of oropharyngeal cancer: A systematic review and meta-analysis of 1366 patients. *Eur J Cancer*. 2017;78:91-102. doi:10.1016/j.ejca.2017.03.006
74. Buchberger AMS, Strzelczyk EA, Wollenberg B, Combs SE, Pickhard A, Pigorsch SU. Report on Late Toxicity in Head-and-Neck Tumor Patients with Long Term Survival after Radiochemotherapy. *Cancers (Basel)*. 2021;13(17). doi:10.3390/cancers13174292
75. Langendijk JA, Doornaert P, Verdonck-de Leeuw IM, Leemans CR, Aaronson NK, Slotman BJ. Impact of late treatment-related toxicity on quality of life among patients with head and neck cancer treated with radiotherapy. *J Clin Oncol*. 2008;26(22):3770-3776. doi:10.1200/JCO.2007.14.6647
76. Vissink A, Jansma J, Spijkervet FKL, Burlage FR, Coppes RP. Oral sequelae of head and neck radiotherapy. *Crit Rev Oral Biol Med*. 2003;14(3):199-212.
77. Jensen SB, Pedersen AML, Vissink A, et al. A systematic review of salivary gland hypofunction and xerostomia induced by cancer therapies: Management strategies and economic impact. *Supportive Care in Cancer*. 2010;18(8):1061-1079. doi:10.1007/s00520-010-0837-6
78. Jasmer KJ, Gilman KE, Muñoz Forti K, Weisman GA, Limesand KH. Radiation-Induced Salivary Gland Dysfunction: Mechanisms, Therapeutics and Future Directions. *J Clin Med*. 2020;9(12). doi:10.3390/jcm9124095
79. Nutting CM, Morden JP, Harrington KJ, et al. Parotid-sparing intensity modulated versus conventional radiotherapy in head and neck cancer (PARSPORT): a phase 3 multicentre randomised controlled trial. *Lancet Oncol*. 2011;12(2):127-136. doi:10.1016/S1470-2045(10)70290-4
80. Li Y, Taylor JMG, Ten Haken RK, Eisbruch A. The impact of dose on parotid salivary recovery in head and neck cancer patients treated with radiation therapy. *Int J Radiat Oncol Biol Phys*. 2007;67(3):660-669. doi:10.1016/j.ijrobp.2006.09.021
81. Eisbruch A, Ten Haken RK, Kim HM, Marsh LH, Ship JA. Dose, volume, and function relationships in parotid salivary glands following conformal and intensity-modulated irradiation of head and neck cancer. *Int J Radiat Oncol Biol Phys*. 1999;45(3):577-587. doi:10.1016/s0360-3016(99)00247-3

82. Deasy JO, Moiseenko V, Marks L, Chao KSC, Nam J, Eisbruch A. Radiotherapy dose-volume effects on salivary gland function. *Int J Radiat Oncol Biol Phys*. 2010;76(3 Suppl):S58-63. doi:10.1016/j.ijrobp.2009.06.090
83. Pedersen AML, Sørensen CE, Proctor GB, Carpenter GH, Ekström J. Salivary secretion in health and disease. *J Oral Rehabil*. 2018;45(9):730-746. doi:10.1111/joor.12664
84. Jensen SB, Vissink A, Limesand KH, Reyland ME. Salivary Gland Hypofunction and Xerostomia in Head and Neck Radiation Patients. *J Natl Cancer Inst Monogr*. 2019;2019(53). doi:10.1093/jncimonographs/lgz016
85. Pinna R, Campus G, Cumbo E, Mura I, Milia E. Xerostomia induced by radiotherapy: an overview of the physiopathology, clinical evidence, and management of the oral damage. *Ther Clin Risk Manag*. 2015;11:171-188. doi:10.2147/TCRM.S70652
86. Patterson JM, Lu L, Watson LJ, et al. Associations between markers of social functioning and depression and quality of life in survivors of head and neck cancer: Findings from the Head and Neck Cancer 5000 study. *Psychooncology*. 2022;31(3):478-485. doi:10.1002/pon.5830
87. Riley P, Glenny AM, Hua F, Worthington H V. Pharmacological interventions for preventing dry mouth and salivary gland dysfunction following radiotherapy. *Cochrane Database Syst Rev*. 2017;7(7):CD012744. doi:10.1002/14651858.CD012744
88. Sung SY, Kim YS, Kim SH, Lee SJ, Lee SW, Kwak YK. Current Evidence of a Deintensification Strategy for Patients with HPV-Related Oropharyngeal Cancer. *Cancers (Basel)*. 2022;14(16). doi:10.3390/cancers14163969
89. Mensour EA, Alam S, Mawani S, et al. What is the future of treatment de-escalation for HPV-positive oropharyngeal cancer? A review of ongoing clinical trials. *Front Oncol*. 2022;12:1067321. doi:10.3389/fonc.2022.1067321
90. Mehanna H, Robinson M, Hartley A, et al. Radiotherapy plus cisplatin or cetuximab in low-risk human papillomavirus-positive oropharyngeal cancer (De-ESCALaTE HPV): an open-label randomised controlled phase 3 trial. *Lancet*. 2019;393(10166):51-60. doi:10.1016/S0140-6736(18)32752-1
91. Gillison ML, Trotti AM, Harris J, et al. Radiotherapy plus cetuximab or cisplatin in human papillomavirus-positive oropharyngeal cancer (NRG Oncology RTOG 1016): a randomised, multicentre, non-inferiority trial. *Lancet*. 2019;393(10166):40-50. doi:10.1016/S0140-6736(18)32779-X
92. Rasmussen JH, Grønhøj C, Håkansson K, et al. Risk profiling based on p16 and HPV DNA more accurately predicts location of disease relapse in patients with oropharyngeal squamous cell carcinoma. *Ann Oncol*. 2019;30(4):629-636. doi:10.1093/annonc/mdz010
93. Grønhøj C, Jensen JS, Wagner S, et al. Impact on survival of tobacco smoking for cases with oropharyngeal squamous cell carcinoma and known human papillomavirus and p16-status: a multicenter retrospective study. *Oncotarget*. 2019;10(45):4655-4663. doi:10.18632/oncotarget.27079

94. Joura EA, Giuliano AR, Iversen OE, et al. A 9-Valent HPV Vaccine against Infection and Intraepithelial Neoplasia in Women. *N Engl J Med*. 2015;372(8):711-723. doi:10.1056/NEJMoa1405044
95. Lehtinen M, Luostarinen T, Vänskä S, et al. Gender-neutral vaccination provides improved control of human papillomavirus types 18/31/33/35 through herd immunity: Results of a community randomized trial (III). *Int J Cancer*. 2018;143(9):2299-2310. doi:10.1002/ijc.31618
96. Chaturvedi AK, Graubard BI, Broutian T, et al. Effect of Prophylactic Human Papillomavirus (HPV) Vaccination on Oral HPV Infections Among Young Adults in the United States. *J Clin Oncol*. 2018;36(3):262-267. doi:10.1200/JCO.2017.75.0141
97. Nielsen KJ, Jakobsen KK, Jensen JS, Grønhøj C, Von Buchwald C. The Effect of Prophylactic HPV Vaccines on Oral and Oropharyngeal HPV Infection-A Systematic Review. *Viruses*. 2021;13(7). doi:10.3390/v13071339
98. Lehtinen T, Elfström KM, Mäkitie A, et al. Elimination of HPV-associated oropharyngeal cancers in Nordic countries. *Prev Med (Baltim)*. 2021;144:106445. doi:10.1016/j.ypmed.2021.106445
99. Karp EE, Yin LX, Moore EJ, et al. Barriers to Obtaining a Timely Diagnosis in Human Papillomavirus-Associated Oropharynx Cancer. *Otolaryngol Head Neck Surg*. 2021;165(2):300-308. doi:10.1177/0194599820982662
100. Jerjes W. Comments on “Epidemiological trends and survival of oropharyngeal cancer in a high HPV-prevalent area: A Danish population-based study from 2000 to 2020”. *Int J Cancer*. Published online September 2024. doi:10.1002/ijc.35178
101. Sharma SJ, Schartinger VH, Wuerdemann N, et al. Awareness of Human Papillomavirus (HPV) and HPV Vaccination amongst the General Population in Germany: Lack of Awareness and Need for Action. *Oncol Res Treat*. 2022;45(10):561-566. doi:10.1159/000525697
102. Verhees F, Demers I, Schouten LJ, Lechner M, Speel EJM, Kremer B. Public awareness of the association between human papillomavirus and oropharyngeal cancer. *Eur J Public Health*. 2021;31(5):1021-1025. doi:10.1093/eurpub/ckab081
103. <https://www.altinget.dk/artikel/drenge-mister-retten-til-kraeftvaccine-i-ny-finanslov>.
104. Qu W, Hua C, Wang Y, et al. Lineage Replacement and Genetic Changes of Four HR-HPV Types during the Period of Vaccine Coverage: A Six-Year Retrospective Study in Eastern China. *Vaccines (Basel)*. 2024;12(4):1-13. doi:10.3390/vaccines12040411
105. Schutte HW, Heutink F, Wellenstein DJ, et al. Impact of Time to Diagnosis and Treatment in Head and Neck Cancer: A Systematic Review. *Otolaryngol Head Neck Surg*. 2020;162(4):446-457. doi:10.1177/0194599820906387
106. Yin LX, Karp EE, Elias A, et al. Disease Profile and Oncologic Outcomes After Delayed Diagnosis of Human Papillomavirus-Associated Oropharyngeal Cancer. *Otolaryngol Head Neck Surg*. 2021;165(6):830-837. doi:10.1177/01945998211000426
107. Garland SM, Kjaer SK, Muñoz N, et al. Impact and Effectiveness of the Quadrivalent Human Papillomavirus Vaccine: A Systematic Review of 10 Years of Real-world Experience. *Clin Infect Dis*. 2016;63(4):519-527. doi:10.1093/cid/ciw354

108. Drolet M, Bénard É, Pérez N, Brisson M. Population-level impact and herd effects following the introduction of human papillomavirus vaccination programmes: updated systematic review and meta-analysis. *Lancet*. 2019;394(10197):497-509. doi:10.1016/S0140-6736(19)30298-3
109. Kreimer AR, Pierce Campbell CM, Lin HY, et al. Incidence and clearance of oral human papillomavirus infection in men: the HIM cohort study. *Lancet*. 2013;382(9895):877-887. doi:10.1016/S0140-6736(13)60809-0
110. Agalliu I, Gapstur S, Chen Z, et al. Associations of Oral α -, β -, and γ -Human Papillomavirus Types With Risk of Incident Head and Neck Cancer. *JAMA Oncol*. 2016;2(5):599-606. doi:10.1001/jamaoncol.2015.5504
111. D'Souza G, Clemens G, Strickler HD, et al. Long-term Persistence of Oral HPV Over 7 Years of Follow-up. *JNCI Cancer Spectr*. 2020;4(5):pkaa047. doi:10.1093/jncics/pkaa047
112. Fakhry C, Rosenthal BT, Clark DP, Gillison ML. Associations between oral HPV16 infection and cytopathology: evaluation of an oropharyngeal "pap-test equivalent" in high-risk populations. *Cancer Prev Res (Phila)*. 2011;4(9):1378-1384. doi:10.1158/1940-6207.CAPR-11-0284
113. D'Souza G, Clemens G, Troy T, et al. Evaluating the Utility and Prevalence of HPV Biomarkers in Oral Rinses and Serology for HPV-related Oropharyngeal Cancer. *Cancer Prev Res (Phila)*. 2019;12(10):689-700. doi:10.1158/1940-6207.CAPR-19-0185
114. Kreimer AR, Johansson M, Waterboer T, et al. Evaluation of human papillomavirus antibodies and risk of subsequent head and neck cancer. *J Clin Oncol*. 2013;31(21):2708-2715. doi:10.1200/JCO.2012.47.2738
115. Lehtinen M, Butt J, Gray P, et al. HPV16 E6-antibody associated risk of oropharyngeal cancer increases by calendar-time: A nested case-control study. *Int J Cancer*. 2023;153(6):1313-1315. doi:10.1002/ijc.34614
116. Friedenstein AJ, Petrakova K V, Kurolesova AI, Frolova GP. Heterotopic of bone marrow. Analysis of precursor cells for osteogenic and hematopoietic tissues. *Transplantation*. 1968;6(2):230-247.
117. Singer NG, Caplan AI. Mesenchymal Stem Cells: Mechanisms of Inflammation. *Annu Rev Pathol Mech Dis*. 2011;6:457-478. doi:10.1146/annurev-pathol-011110-130230
118. Kern S, Eichler H, Stoeve J, Klüter H, Bieback K. Comparative analysis of mesenchymal stem cells from bone marrow, umbilical cord blood, or adipose tissue. *Stem Cells*. 2006;24(5):1294-1301. doi:10.1634/stemcells.2005-0342
119. Pittenger MF, Mackay AM, Beck SC, et al. Multilineage potential of adult human mesenchymal stem cells. *Science*. 1999;284(5411):143-147. doi:10.1126/science.284.5411.143
120. Dominici M, Le Blanc K, Mueller I, et al. Minimal criteria for defining multipotent mesenchymal stromal cells. The International Society for Cellular Therapy position statement. *Cytotherapy*. 2006;8(4):315-317. doi:10.1080/14653240600855905
121. Von Bahr L, Batsis I, Moll G, et al. Analysis of tissues following mesenchymal stromal cell therapy in humans indicates limited long-term engraftment and no ectopic tissue formation. *Stem Cells*. 2012;30(7):1575-1578. doi:10.1002/stem.1118

122. Krampera M, Le Blanc K. Mesenchymal stromal cells: Putative microenvironmental modulators become cell therapy. *Cell Stem Cell*. 2021;28(10):1708-1725. doi:10.1016/j.stem.2021.09.006
123. Panés J, García-Olmo D, Van Assche G, et al. Expanded allogeneic adipose-derived mesenchymal stem cells (Cx601) for complex perianal fistulas in Crohn's disease: a phase 3 randomised, double-blind controlled trial. *The Lancet*. 2016;388(10051):1281-1290. doi:10.1016/S0140-6736(16)31203-X
124. Toyserkani NM, Jørgensen MG, Tabatabaeifar S, Jensen CH, Sheikh SP, Sørensen JA. Concise Review: A Safety Assessment of Adipose-Derived Cell Therapy in Clinical Trials: A Systematic Review of Reported Adverse Events. *Stem Cells Transl Med*. 2017;6(9):1786-1794. doi:10.1002/sctm.17-0031
125. Rangatchew F, Vester-Glowinski P, Rasmussen BS, et al. Mesenchymal stem cell therapy of acute thermal burns: A systematic review of the effect on inflammation and wound healing. *Burns*. 2021;47(2):270-294. doi:10.1016/j.burns.2020.04.012
126. Ankrum JA, Ong JF, Karp JM. Mesenchymal stem cells: Immune evasive, not immune privileged. *Nat Biotechnol*. 2014;32(3):252-260. doi:10.1038/nbt.2816
127. Di Nicola M, Carlo-Stella C, Magni M, et al. Human bone marrow stromal cells suppress T-lymphocyte proliferation induced by cellular or nonspecific mitogenic stimuli. *Blood*. 2002;99(10):3838-3843. doi:10.1182/blood.v99.10.3838
128. Jakobsen K, Carlander ALF, Todsén T, et al. Mesenchymal Stem/Stromal Cell Therapy for Radiation-Induced Xerostomia in Previous Head and Neck Cancer Patients: A Phase 2 Randomised, Placebo-Controlled Trial. *Clin Cancer Res*. Published online March 2024. doi:10.1158/1078-0432.CCR-23-3675
129. Lynggaard CD, Grønhøj C, Christensen R, et al. Intraglandular Off-the-Shelf Allogeneic Mesenchymal Stem Cell Treatment in Patients with Radiation-Induced Xerostomia: A Safety Study (MESRIX-II). *Stem Cells Transl Med*. 2022;11(5):478-489. doi:10.1093/stcltm/szac011
130. Grønhøj C, Jensen DH, Vester-Glowinski P, et al. Safety and Efficacy of Mesenchymal Stem Cells for Radiation-Induced Xerostomia: A Randomized, Placebo-Controlled Phase 1/2 Trial (MESRIX). *Int J Radiat Oncol Biol Phys*. 2018;101(3):581-592. doi:10.1016/j.ijrobp.2018.02.034
131. Lynggaard CD, Grønhøj C, Jensen SB, et al. Long-term Safety of Treatment with Autologous Mesenchymal Stem Cells in Patients with Radiation-Induced Xerostomia: Primary Results of the MESRIX Phase I/II Randomized Trial. *Clin Cancer Res*. 2022;28(13):2890-2897. doi:10.1158/1078-0432.CCR-21-4520
132. Jakobsen KK, Carlander ALF, Grønhøj C, et al. Effectiveness and safety of mesenchymal stem/stromal cell for radiation-induced hyposalivation and xerostomia in previous head and neck cancer patients (MESRIX-III): a study protocol for a single-centre, double-blinded, randomised, placebo-controlled, pha. *Trials*. 2023;24(1):1-9. doi:10.1186/s13063-023-07594-5
133. Blitzer GC, Glazer T, Burr A, et al. Marrow-Derived Autologous Stromal Cells for the Restoration of Salivary Hypofunction (MARSH): A pilot, first-in-human study of interferon gamma-stimulated marrow mesenchymal stromal cells for treatment of radiation-induced xerostomia. *Cytotherapy*. 2023;25(11):1139-1144. doi:10.1016/j.jcyt.2023.07.009

134. Wilkins MR, Sanchez JC, Gooley AA, et al. Progress with proteome projects: why all proteins expressed by a genome should be identified and how to do it. *Biotechnol Genet Eng Rev.* 1996;13:19-50. doi:10.1080/02648725.1996.10647923
135. Thomas SN, French D, Jannetto PJ, Rappold BA, Clarke WA. Liquid chromatography–tandem mass spectrometry for clinical diagnostics. *Nature Reviews Methods Primers.* 2022;2(1):1-14. doi:10.1038/s43586-022-00175-x
136. Denny P, Hagen FK, Hardt M, et al. The proteomes of human parotid and submandibular/sublingual gland salivas collected as the ductal secretions. *J Proteome Res.* 2008;7(5):1994-2006. doi:10.1021/pr700764j
137. Dijkema T, Terhaard CHJ, Roesink JM, et al. MUC5B levels in submandibular gland saliva of patients treated with radiotherapy for head-and-neck cancer: a pilot study. *Radiat Oncol.* 2012;7:91. doi:10.1186/1748-717X-7-91
138. Lynggaard CD, Jersie-Christensen R, Juhl M, et al. Intraglandular mesenchymal stem cell treatment induces changes in the salivary proteome of irradiated patients. *Communications medicine.* 2022;2(1):160. doi:10.1038/s43856-022-00223-3
139. Brandt E, Keskin M, Räisänen IT, et al. Induction of Collagenolytic MMP-8 and -9 Tissue Destruction Cascade in Mouth by Head and Neck Cancer Radiotherapy: A Cohort Study. *Biomedicines.* 2023;12(1). doi:10.3390/biomedicines12010027
140. Hynne H, Aqrabi LA, Jensen JL, et al. Proteomic Profiling of Saliva and Tears in Radiated Head and Neck Cancer Patients as Compared to Primary Sjögren’s Syndrome Patients. *Int J Mol Sci.* 2022;23(7). doi:10.3390/ijms23073714
141. <https://health.usnews.com/best-hospitals/rankings/cancer>.
142. Jansson PM, Lynggaard CD, Carlander AF, et al. Mesenchymal stromal/stem cell therapy for radiation-induced salivary gland hypofunction in animal models: a protocol for a systematic review and meta-analysis. *Syst Rev.* 2022;11(1):72. doi:10.1186/s13643-022-01943-2
143. Percie du Sert N, Hurst V, Ahluwalia A, et al. The ARRIVE guidelines 2.0: Updated guidelines for reporting animal research. *PLoS Biol.* 2020;18(7):e3000410. doi:10.1371/journal.pbio.3000410
144. Hooijmans CR, Rovers MM, de Vries RBM, Leenaars M, Ritskes-Hoitinga M, Langendam MW. SYRCLE’s risk of bias tool for animal studies. *BMC Med Res Methodol.* 2014;14:43. doi:10.1186/1471-2288-14-43
145. Carlander ALF, Jakobsen KK, Jersie-Christensen R, et al. Exploring the salivary proteome following intraglandular mesenchymal stromal cell therapy for radiation-induced hyposalivation in previous head and neck cancer patients: a secondary study protocol for the MESRIX-III, randomised, controlled trial. :In review.
146. Kwok J, Langevin SM, Argiris A, Grandis JR, Gooding WE, Taioli E. The impact of health insurance status on the survival of patients with head and neck cancer. *Cancer.* 2010;116(2):476-485. doi:10.1002/cncr.24774

147. Gupta A, Sonis ST, Schneider EB, Villa A. Impact of the insurance type of head and neck cancer patients on their hospitalization utilization patterns. *Cancer*. 2018;124(4):760-768. doi:10.1002/cncr.31095
148. Lassen P, Huang SH, Su J, et al. Treatment outcomes and survival following definitive (chemo)radiotherapy in HPV-positive oropharynx cancer: Large-scale comparison of DAHANCA vs PMH cohorts. *Int J Cancer*. 2022;150(8):1329-1340. doi:10.1002/ijc.33876
149. Machiels JP, René Leemans C, Golusinski W, Grau C, Licitra L, Gregoire V. Squamous cell carcinoma of the oral cavity, larynx, oropharynx and hypopharynx: EHNS–ESMO–ESTRO Clinical Practice Guidelines for diagnosis, treatment and follow-up†. *Annals of Oncology*. 2020;31(11):1462-1475. doi:10.1016/j.annonc.2020.07.011
150. McGrath K, Wolf M, Bishop J, et al. Transient platelet and HLA antibody formation in multitransfused patients with malignancy. *Br J Haematol*. 1988;68(3):345-350. doi:10.1111/j.1365-2141.1988.tb04212.x
151. Ziemann M, Altermann W, Angert K, et al. Preformed Donor-Specific HLA Antibodies in Living and Deceased Donor Transplantation: A Multicenter Study. *Clin J Am Soc Nephrol*. 2019;14(7):1056-1066. doi:10.2215/CJN.13401118
152. Hiraoka S, Yoshimura M, Nakajima A, Nakashima R, Mizowaki T. Long-term outcomes of stimulated salivary flow and xerostomia after definitive intensity-modulated radiation therapy for patients with head and neck cancer†. *J Radiat Res*. 2024;65(1):71-77. doi:10.1093/jrr/rrad087
153. Huang SH, Wu CH, Chen SJ, Sytwu HK, Lin GJ. Immunomodulatory effects and potential clinical applications of dimethyl sulfoxide. *Immunobiology*. 2020;225(3):151906. doi:10.1016/j.imbio.2020.151906
154. Hoang C, Nguyen AK, Nguyen TQ, et al. Application of Dimethyl Sulfoxide as a Therapeutic Agent and Drug Vehicle for Eye Diseases. *J Ocul Pharmacol Ther*. 2021;37(8):441-451. doi:10.1089/jop.2021.0043
155. Karim M, Boikess RS, Schwartz RA, Cohen PJ. Dimethyl sulfoxide (DMSO): a solvent that may solve selected cutaneous clinical challenges. *Arch Dermatol Res*. 2023;315(6):1465-1472. doi:10.1007/s00403-022-02494-1
156. Mohamed-Ahmed S, Fristad I, Lie SA, et al. Adipose-derived and bone marrow mesenchymal stem cells: A donor-matched comparison. *Stem Cell Res Ther*. 2018;9(1):1-15. doi:10.1186/s13287-018-0914-1
157. Gholami N, Hosseini Sabzvari B, Razzaghi A, Salah S. Effect of stress, anxiety and depression on unstimulated salivary flow rate and xerostomia. *J Dent Res Dent Clin Dent Prospects*. 2017;11(4):247-252. doi:10.15171/joddd.2017.043
158. Elishoov H, Wolff A, Kravel LS, Shiperman A, Gorsky M. Association between season and temperature and unstimulated parotid and submandibular/sublingual secretion rates. *Arch Oral Biol*. 2008;53(1):75-78. doi:10.1016/j.archoralbio.2007.08.002
159. Lynge Pedersen AM, Belstrøm D. The role of natural salivary defences in maintaining a healthy oral microbiota. *J Dent*. 2019;80 Suppl 1:S3-S12. doi:10.1016/j.jdent.2018.08.010

160. Azzam EI, Jay-Gerin JP, Pain D. Ionizing radiation-induced metabolic oxidative stress and prolonged cell injury. *Cancer Lett.* 2012;327(1-2):48-60. doi:10.1016/j.canlet.2011.12.012
161. Laheij AMGA, Rasch CN, Brandt BW, et al. Proteins and peptides in parotid saliva of irradiated patients compared to that of healthy controls using SELDI-TOF-MS Oral Health. *BMC Res Notes.* 2015;8(1):1-7. doi:10.1186/s13104-015-1641-7
162. Gomar-Vercher S, Simón-Soro A, Montiel-Company JM, Almerich-Silla JM, Mira A. Stimulated and unstimulated saliva samples have significantly different bacterial profiles. *PLoS One.* 2018;13(6):e0198021. doi:10.1371/journal.pone.0198021
163. Foratori-Junior GA, Le Guennec A, Fidalgo TK da S, et al. Comparison of the Metabolic Profile between Unstimulated and Stimulated Saliva Samples from Pregnant Women with/without Obesity and Periodontitis. *J Pers Med.* 2023;13(7):1-21. doi:10.3390/jpm13071123
164. Ährlund-Richter A, Holzhauser S, Dalianis T, Näsman A, Mints M. Whole-Exome Sequencing of HPV Positive Tonsillar and Base of Tongue Squamous Cell Carcinomas Reveals a Global Mutational Pattern along with Relapse-Specific Somatic Variants. *Cancers (Basel).* 2021;14(1). doi:10.3390/cancers14010077
165. Matas J, Orrego M, Amenabar D, et al. Umbilical Cord-Derived Mesenchymal Stromal Cells (MSCs) for Knee Osteoarthritis: Repeated MSC Dosing Is Superior to a Single MSC Dose and to Hyaluronic Acid in a Controlled Randomized Phase I/II Trial. *Stem Cells Transl Med.* 2019;8(3):215-224. doi:10.1002/sctm.18-0053
166. Petrou P, Kassis I, Ginzberg A, et al. Long-Term Clinical and Immunological Effects of Repeated Mesenchymal Stem Cell Injections in Patients With Progressive Forms of Multiple Sclerosis. *Front Neurol.* 2021;12:639315. doi:10.3389/fneur.2021.639315
167. Chihaby N, Orliaguet M, Le Pottier L, Pers JO, Boisramé S. Treatment of Sjögren's Syndrome with Mesenchymal Stem Cells: A Systematic Review. *Int J Mol Sci.* 2021;22(19). doi:10.3390/ijms221910474
168. Wolff A, Joshi RK, Ekström J, et al. A Guide to Medications Inducing Salivary Gland Dysfunction, Xerostomia, and Subjective Sialorrhea: A Systematic Review Sponsored by the World Workshop on Oral Medicine VI. *Drugs in R and D.* 2017;17(1):1-28. doi:10.1007/s40268-016-0153-9

Appendix I – 8th edition of the TNM AJCC/UICC staging manual for OPSCC⁵³

p16+ OPSCC

T category	N category	M category	Overall stage
T0, T1 or T2	N0 or N1	M0	I
T0, T1 or T2	N2	M0	II
T3	N0, N1 or N2	M0	II
T0, T1, T2, T3 or T4	N3	M0	III
T4	N0, N1, N2 or N3	M0	III
Any T	Any N	M1	IV

p16- OPSCC

T category	N category	M category	Overall stage
Tis	N0	M0	0
T1	N0	M0	I
T2	N0	M0	II
T3	N0	M0	III
T1, T2 or T3	N1	M0	III
T4a	N0 or N1	M0	IVA
T T1, T2, T3 or T4a	N2	M0	IVA
Any T	N3	M0	IVB
T4b	Any N	M0	IVB
Any T	Any N	M1	IVC

Abbreviations: AJCC, American Joint Committee on Cancer; M, metastases; N, nodes; p16, p16^{Ink4a} tumor suppressor protein; T, tumor; UICC, Union of International Cancer Control.

Appendix II – Xerogenic medicine¹⁶⁸

Anatomical group	Therapeutic group	Chemical substance
Alimentary tract/metabolism	Anticholinergics	Propantheline
		Atropine
		Scopolamine
Cardiovascular system	Antihypertensives	Clonidine
		Methyldopa
	Diuretics	Bendroflumethiazide
		Furosemide
		Tolvaptan
	Beta-blockers	Timolol
		Metoprolol
		Atenolol
	Calcium channel blockers	Isradipine
		Verapamil
	ACE inhibitors	Enalapril
		Lisinopril
Musculoskeletal system	Bisphosphonates	Alendronate
Nervous system	Analgesics	Fentanyl
		Morphine
		Buprenorphine
		Butorphanol
		Tramadol
		Clonidine
	Antipsychotics	Chlorpromazine
		Haloperidol
		Clozapine
		Olanzapine
		Quetiapine
		Lithium

		Risperidone Zolpidem
	Antidepressants	Amitriptyline Citalopram Sertraline Escitalopram Venflaxine Duloxetine Bupropion
Respiratory system	Drugs for obstructive airway diseases	Tiotropium
	Antihistamines	Cetirizine Doxylamine
Sensory organs	Ophthalmologicals	Atropine Brimonidine

*Modified table from Wolff et al., Drug R D, 2017¹⁶⁸

Abbreviations: ACE, angiotensin-converting enzyme.



EORTC QLQ - H&N35

Patienter fortæller undertiden, at de har følgende symptomer eller problemer. Anfør venligst, i hvilket omfang De har haft disse symptomer eller problemer inden for den forløbne uge. Besvar spørgsmålene ved at sætte en ring omkring det tal, som passer bedst til Dem.

I den forløbne uge:	Slet ikke	Lidt	En del	Meget
31. Har De haft smerter i munden?	1	2	3	4
32. Har De haft smerter i kæben?	1	2	3	4
33. Har De været øm i munden?	1	2	3	4
34. Har De haft ondt i halsen?	1	2	3	4
35. Har De haft svært ved at synke væske?	1	2	3	4
36. Har De haft svært ved at synke most, blendet eller pureret mad?	1	2	3	4
37. Har De haft svært ved at synke fast føde?	1	2	3	4
38. Har De fået noget galt i halsen, når De har sunket?	1	2	3	4
39. Har De haft problemer med tænderne?	1	2	3	4
40. Har De haft svært ved at åbne munden helt?	1	2	3	4
41. Har De været tør i munden?	1	2	3	4
42. Har Deres spyt virket klæbende?	1	2	3	4
43. Har De haft problemer med lugtesansen?	1	2	3	4
44. Har De haft problemer med Deres smagssans?	1	2	3	4
45. Har De hostet?	1	2	3	4
46. Har De været hæs?	1	2	3	4
47. Har De følt Dem syg?	1	2	3	4
48. Har Deres udseende generet Dem?	1	2	3	4

Gå venligst videre til næste side

I den forløbne uge:		Slet ikke	Lidt	En del	Meget
49.	Har De haft svært ved at spise?	1	2	3	4
50.	Har De haft svært ved at spise i Deres families nærvær?	1	2	3	4
51.	Har De haft svært ved spise i andre folks nærvær?	1	2	3	4
52.	Har De haft svært ved at nyde Deres måltider?	1	2	3	4
53.	Har De haft svært ved at tale til andre?	1	2	3	4
54.	Har De haft svært ved at tale i telefon?	1	2	3	4
55.	Har De haft svært ved at være sammen med Deres familie?	1	2	3	4
56.	Har De haft svært ved at være sammen med venner?	1	2	3	4
57.	Har De haft svært ved at gå ud blandt andre?	1	2	3	4
58.	Har De haft svært ved at have fysisk kontakt med familie eller venner?	1	2	3	4
59.	Har De haft mindre lyst til seksuelt samvær?	1	2	3	4
60.	Har De følt mindre seksuel nydelse?	1	2	3	4

I den forløbne uge:		Nej	Ja
61.	Har De taget smertestillende medicin?	1	2
62.	Har De taget nogen form for kosttilskud (bortset fra vitaminpiller)?	1	2
63.	Har De anvendt en ernæringssonde?	1	2
64.	Har De tabt Dem?	1	2
65.	Har De taget på?	1	2

Appendix V – Scientific papers I-IV



Clinical and prognostic differences in oropharyngeal squamous cell carcinoma in USA and Denmark, two HPV high-prevalence areas

Amanda-Louise Fenger Carlander^{a,*}, Simone Kloch Bendtsen^{a,1}, Jacob H. Rasmussen^a, Kathrine Kronberg Jakobsen^a, Martin Garset-Zamani^a, Christian Grønhoj^a, Jeppe Friberg^b, Katherine Hutcheson^c, Faye M. Johnson^{d,g}, Clifton D. Fuller^e, Amy C. Moreno^e, Toyin Babarinde^f, Neil D. Gross^c, Jeffrey N. Myers^{f,g}, Christian von Buchwald^a

^a Department of Otolaryngology, Head and Neck Surgery & Audiology, Copenhagen University Hospital - Rigshospitalet, Copenhagen, Denmark

^b Department of Oncology, Copenhagen University Hospital - Rigshospitalet, Copenhagen, Denmark

^c Department of Head and Neck Surgery, Division of Surgery, The University of Texas M.D. Anderson Cancer Center, UTMDACC, TX, USA

^d Department of Thoracic Head and Neck Medical Oncology, The University of Texas M.D. Anderson Cancer Center, UTMDACC, TX, USA

^e Department of Radiation Oncology, The University of Texas M.D. Anderson Cancer Center, UTMDACC, TX, USA

^f Department of Head and Neck Surgery, The University of Texas M.D. Anderson Cancer Center, UTMDACC, TX, USA

^g The University of Texas Graduate School of Biomedical Sciences; UTMDACC, TX, USA

ARTICLE INFO

Keywords:

Human papillomavirus
Oropharyngeal cancer
Squamous cell carcinoma
Survival
Recurrence
Prognosis
Prevalence
Demographics

ABSTRACT

Background: Uncertainty persists regarding clinical and treatment variations crucial to consider when comparing high human papillomavirus (HPV)-prevalence oropharyngeal squamous cell carcinoma (OPSCC) cohorts for accurate patient stratification and replicability of clinical trials across different geographical areas.

Methods: OPSCC patients were included from The University of Texas MD Anderson Cancer Center (UTMDACC), USA and from The University Hospital of Copenhagen, Denmark from 2015–2020, (n = 2484). Outcomes were 3-year overall survival (OS) and recurrence-free interval (RFI). Subgroup analyses were made for low-risk OPSCC patients (T1–2N0M0) and high-risk patients (UICC8 III-IV).

Results: There were significantly more HPV-positive (88.2 % vs. 63.1 %), males (89.4 % vs. 74.1 %), never-smokers (52.1 % vs. 23.7 %), lower UICC8-stage (I/II: 79.3 % vs. 68 %), and fewer patients treated with radiotherapy (RT) alone (14.8 % vs. 30.3 %) in the UTMDACC cohort. No difference in the adjusted OS was observed (hazard ratio [HR] 1.21, p = 0.23), but a significantly increased RFI HR was observed for the Copenhagen cohort (HR: 1.74, p = 0.003). Subgroup analyses of low- and high-risk patients revealed significant clinical and treatment differences. No difference in prognosis was observed for low-risk patients, but the prognosis for high-risk patients in the Copenhagen cohort was worse (OS HR 2.20, p = 0.004, RFI HR 2.80, p = 0.002).

Conclusions: We identified significant differences in clinical characteristics, treatment modalities, and prognosis between a Northern European and Northern American OPSCC population. These differences are important to consider when comparing outcomes and for patient stratification in clinical trials, as reproducibility might be challenging.

1. Introduction

Human papillomavirus (HPV) is a key factor in the rising incidence of oropharyngeal squamous cell carcinoma (OPSCC), but with

geographical variation [1–4]. How HPV-positivity is defined varies, but the expression of p16 is widely accepted as a surrogate marker for HPV-positivity. However, double p16/HPV-positivity more accurately classify biologically active HPV infection, better predicts patient

* Correspondence to: Department of Otolaryngology, Head and Neck Surgery & Audiology, Copenhagen University Hospital - Rigshospitalet, Inge Lehmanns vej 8, 2100 Copenhagen Oe, Denmark.

E-mail address: amanda-louise.fenger.carlander@regionh.dk (A.-L.F. Carlander).

¹ Shared first authorship.

<https://doi.org/10.1016/j.ejca.2024.113983>

Received 21 November 2023; Received in revised form 15 January 2024; Accepted 28 February 2024

Available online 2 March 2024

0959-8049/© 2024 The Author(s). Published by Elsevier Ltd. This is an open access article under the CC BY license (<http://creativecommons.org/licenses/by/4.0/>).

outcomes, and standardized HPV testing has been recommended [4,5].

Given the remarkable treatment response [6] and the toxicities associated with treatment [7], attempts have been made to identify low-risk HPV-positive (HPV+) patients suitable for treatment de-intensification [8,9]. Some well described factors associated with low-risk OPSCC include limited smoking history, p16 + status, and low tumor burden [6].

Both the U.S. and Denmark are HPV high-prevalence areas [10,11]. However, the healthcare systems in Denmark and the U.S. exhibit significant differences. Denmark operates under a public, universal healthcare system and all cancer diagnoses, treatments, and follow-ups are conducted with the standardized cancer care packages available within the public health care system. The healthcare system in the U.S. is larger and more complex, with patients being treated at private or public care centers based on a variety of factors, including access to health insurance.

The consideration of potential clinical and demographic variations is crucial when interpreting the prognosis across different patient populations, for accurate patient stratification and management and to ensure reproducibility and replicability of clinical trials in diverse geographical areas.

The aim of this study was to identify potential differences in clinical characteristics, treatment modalities given, and prognosis among OPSCC patients in two HPV high-prevalence areas with distinct populations and health care systems: The Copenhagen Oropharyngeal Cancer Database (COHOC) at the University Hospital of Copenhagen, Denmark and the Stiefel OPSCC Database at The University of Texas MD Anderson Cancer Center, USA (UTMDACC). Second, we wished to identify if such differences impact the prognosis for a subgroup of low-risk (T1T2N0M0) and high-risk (III-IV) patients potentially eligible for de-escalated or intensified therapy, respectively.

2. Materials and methods

2.1. Study design, setting and population

This retrospective cohort study was approved by The Regional Scientific Ethical Committee (H-20072877) and the Danish Data Protection Agency and followed the Strengthening the Reporting of Observational Studies in Epidemiology (STROBE) reporting guideline [12]. The Stiefel OPSCC Database is approved by The Institutional Review Board of the University of Texas MD Anderson Cancer Center (UTMDACC) (protocol PA 14-0947).

The study cohort consisted of all patients with a new OPSCC diagnosed from 2015–2020 treated with curative intent from two independent OPSCC cohorts: *The COHOC Database* is a population-based, retrospective cohort, comprising patients diagnosed with OPSCC from 2000–2020 in Eastern Denmark and was previously well described [13–15]. *The Stiefel OPSCC Database* is a prospective, longitudinal cohort initiated at UTMDACC. Since 2015, 1671 OPSCC patients have been enrolled with detailed characterization of exposures, HPV-status, diagnostic/staging, treatment, disease control, and longitudinal collection of validated clinician- and patient-reported survivorship outcomes from diagnosis through 5-years of follow-up.

2.2. Variables

The primary outcomes were 3-year overall survival (OS) and 3-year recurrence-free interval (RFI). OS was defined as time from diagnosis to death of any cause and patients alive were censored at the last date of follow-up or 3 years after diagnosis. RFI was defined as the time from primary diagnosis to the diagnosis of recurrence (locoregional and/or distant) and patients without recurrence were censored at last day of follow-up, date of death or 3-years after diagnosis. HPV-positivity was defined as being positive for both HPV DNA and p16 in the Copenhagen cohort, while HPV-positivity was defined as being positive to HPV or

Table 1

Characteristics of 2484 patients with OPSCC in Eastern Denmark and UTM-DACC, USA from 2015–2020.

Variable	Eastern Denmark. n = 1216		UTMDACC, USA. n = 1268		P-value ^a
	no.	%	no.	%	
Gender					< 0.001 ^a
Male	901	74.1	1133	89.4	
Female	315	25.9	135	10.6	
Median age (IQR)	62 (56-70)		60 (54-67)		< 0.001 ^a
no.	1216		1268		
Median follow-up. years (IQR)	2.3 (1.7-3.3)		2.1 (1-3.6)		< 0.001 ^a
no.	1216		1210		
Smoking					< 0.001 ^a
Current	464	38.2	66	5.2	
Former	458	37.7	535	42.2	
Never	288	23.7	661	52.1	
Unknown	6	0.5	6	0.5	
HPV-status					< 0.001 ^a
HPV+	767	63.1	1119	88.2	
HPV-	420	34.5	98	7.7	
Unknown	29	2.4	51	4	
HPV/p16-status					< 0.001 ^a
HPV+/p16+	767	63.1	345	27.2	
HPV+/p16-	49	4.0	8	0.6	
HPV-/p16+	35	2.9	18	1.4	
HPV-/p16-	49	4.0	8	0.6	
Unknown	327	26.9	23	1.8	
Unknown	38	3.1	874	68.9	
HPV genotype					< 0.001 ^a
HPV16	662	87.8	109	87.9	
HPV33	59	7.8	1	0.87	
Other genotypes	33	4.4	14	11.3	
Tumor location					< 0.001 ^a
BOT	374	30.8	617	48.7	
Tonsil	628	51.6	599	47.2	
Other	214	17.6	52	4.1	
TNM Stage (UICC/AJCC 8), HPV+ T-class					0.005 ^a
T1	241	31.4	400	35.7	
T2	289	37.7	433	38.7	
T3	116	15.1	168	15.0	
T4	119	15.5	118	10.5	
Unknown	2	0.3	0		
N-class					< 0.001 ^a
N0	121	15.8	125	11.2	
N1	514	67.0	768	68.6	
N2	103	13.4	203	18.1	
N3	29	3.8	23	2.1	
Unknown	0		0		
M-class					0.01 ^a
M0	762	99.3	1093	97.7	
M1	2	0.3	14	1.8	
Unknown	3	0.4	12	1.1	
Overall Stage (UICC/AJCC 8), HPV+					0.001 ^a
I	462	60.2	709	63.4	
II	220	28.7	259	23.1	
III	81	10.6	124	11.1	
IV	1	0.1	15	1.3	
Unknown	3	0.4	12	1.1	
TNM Stage (UICC/AJCC 8), HPV- T-class					0.74
T1	105	25.0	30	30.6	
T2	138	32.9	32	32.7	
T3	92	21.9	20	20.4	
T4	84	20.0	16	16.3	

(continued on next page)

Table 1 (continued)

Variable	Eastern Denmark, n = 1216		UTMDACC, USA, n = 1268		P-value ^a
	no.	%	no.	%	
Unknown	1	0.2	0		
N-class					0.001 ^a
N0	162	38.6	21	21.4	
N1	76	18.1	34	34.7	
N2	156	37.1	37	37.8	
N3	24	5.7	6	6.1	
Unknown	2	0.5	0		
M-class					0.001 ^a
M0	412	98.1	93	94.9	
M1	6	1.4	1	1.0	
Unknown	2	0.5	4	4.1	
Overall Stage (UICC/AJCC 8), HPV-					0.22
I	74	17.6	18	18.4	
II	71	16.9	9	9.2	
III	72	17.1	18	18.4	
IV	198	47.1	50	51.0	
Unknown	5	1.2	3	3.1	
Treatment					< 0.001 ^a
RT	368	30.3	188	14.8	
CRT	620	51.0	631	49.8	
Neoadjuvant chemotherapy + RT/ CRT	-	-	212	16.7	
Surgery	195	16.0	110	8.7	
Surgery + RT/CRT	33	2.7	124	9.8	
Systemic therapy alone	0	-	2	0.2	
Unspecified curative treatment	0	-	1	0.1	

Note: Frequency (%) is provided for categorical variables, median (IQR) are provided for continuous variables. Chi Square test was used for categorical variables, while t-test was used for continuous variables. Fisher's exact test was used for small sample sizes. Abbreviations: UTMDACC, The University of Texas MD Anderson Cancer Center, USA; IQR, interquartile range; HPV, human papillomavirus; BOT, base of tongue; RT, radiotherapy; CRT, chemoradiotherapy.

^a Significant value.

p16 in the UTMDACC cohort. Details on HPV detection, p16 immunohistochemistry, and measurements are provided in Supplement 1.

2.3. Treatment

Therapeutic decisions were made during a multidisciplinary

treatment planning conference based on factors such as disease stage, anatomical subsite, comorbidity, functional considerations, and the preferences of both patients and clinicians. In Denmark, treatment decisions are in accordance with the Danish Head and Neck Cancer Group (DAHANCA) national guidelines [16,17], while treatment decisions generally follow the National Comprehensive Cancer Network (NCCN) guidelines at UTMDACC [18]. Details on treatment guidelines are provided in Supplement 1.

2.4. Statistics

Statistical analysis was performed in R statistics version 4.1.3. Continuous variables were reported as median values with an interquartile range (IQR) and categorical variables as frequencies. To test for significance, Pearson's chi-square test was used for the binomial categorical covariables, Fisher's exact test was used for small sample sizes, and a t-test was used for the quantitative covariables distributed in two groups. We considered a p-value < 0.05 to be statistically significant. Age and follow-up time were included as continuous variables and the other variables were considered categorical variables.

OS and RFI were evaluated by Kaplan–Meier curves and with the log-rank method and by uni- and multivariable Cox regression analyses (packages Survminer and Survival). For the total OPSCC cohort, the OS (372 events) and RFI (267 events) multivariable analyses were adjusted for treatment center, age (cut-off >60 years), gender, HPV-status, smoking status, tumor location, T-, N- and M-class and treatment modality. For model control, proportionality was tested with Score residuals. Details on subgroup analyses are provided in Supplement 1.

3. Results

3.1. Patient characteristics

A total of 2484 patients were included, 1216 patients from Eastern Denmark and 1268 from UTMDACC. The median follow-up time was 2.1 years (IQR:1.0–3.6) in the UTMDACC cohort and 2.3 years (IQR: 1.7–3.3) in the Copenhagen cohort, p < 0.001. The UTMDACC cohort included significantly more men (89.4 % vs. 74.1 %), younger age (median age 60 vs. 62 years), and never smokers (52.1 % vs. 23.7 %). See Table 1.

3.2. Clinical tumor characteristics

In the UTMDACC cohort, the predominant tumor site was the base of

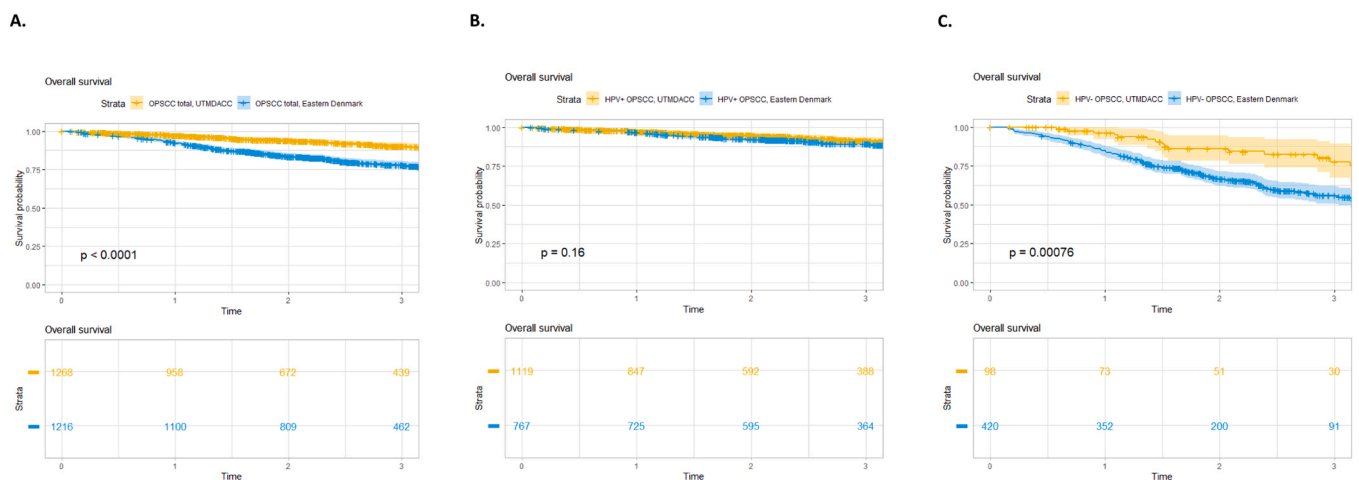


Fig. 1. Kaplan-Meier curves depicting the overall survival probability stratified by center and HPV status from 2015–2020. A. All OPSCC 2015–2020 stratified by center. B. HPV+ OPSCC stratified by center. C. HPV- OPSCC stratified by center. Abbreviations: UTMDACC, The University of Texas MD Anderson Cancer Center, USA, HPV, human papillomavirus; OPSCC, oropharyngeal squamous cell carcinoma.

Table 2

Multi- and univariable analysis for overall survival for OPSCC in Eastern Denmark and UTMDACC from 2015–2020.

Variable	Univariable			Multivariable ^c		
	HR	95 % CI	p-value ^b	HR	95 % CI	p-value ^b
Center (UTMDACC ref)						
Eastern Denmark	2.20	1.77-2.75	< 0.001 ^b	1.21	0.89-1.65	0.23
Age (<60 years ref) > 60 years						
	1.87	1.51-2.32	< 0.001 ^b	1.22	0.97-1.55	0.09
Gender (female ref)						
Male	1.02	0.79-1.32	0.90	1.30	0.99-1.72	0.06
HPV status (HPV+ ref)						
HPV-	4.93	4.00-6.07	< 0.001 ^b	2.56	1.92-3.41	< 0.001 ^b
Smoking status (never ref)						
previous	1.78	1.33-2.39	< 0.001 ^b	1.31	0.96-1.79	0.09
current	5.08	3.83-6.73	< 0.001 ^b	2.26	1.58-3.22	< 0.001 ^b
Tumor location (tonsil ref)						
BOT	0.97	0.76-1.22	0.77	0.95	0.74-1.22	0.69
Other	2.62	2.02-3.41	< 0.001 ^b	0.86	0.64-1.16	0.33
T-class (T1 ref)						
T2	1.42	1.04-1.93	0.03 ^b	1.28	0.93-1.77	0.14
T3	3.37	2.46-4.61	< 0.001 ^b	2.63	1.86-3.71	< 0.001 ^b
T4	3.87	2.84-5.29	< 0.001 ^b	2.52	1.78-3.57	< 0.001 ^b
N-class (N0 ref)						
N1	0.48	0.36-0.64	< 0.001 ^b	1.17	0.85-1.65	0.34
N2	1.60	1.21-2.12	0.001 ^b	1.88	1.38-2.57	< 0.001 ^b
N3	2.92	1.94-4.39	< 0.001 ^b	3.79	2.42-5.95	< 0.001 ^b
M-class (M0 ref)						
M1	3.93	2.26-6.84	< 0.001 ^b	4.06	2.26-7.30	< 0.001 ^b
Treatment regimen (CRT ref)						
RT	2.7	2.15-3.39	< 0.001 ^b	2.21	1.71-2.86	< 0.001 ^b
Neoadjuvant chemotherapy + RT/CRT	1.49	1.04-2.15	0.03 ^b	1.31	0.83-2.07	0.24
Surgery	0.70	0.46-1.07	0.10	1.24	0.78-1.98	0.36
Surgery + RT/CRT	0.56	0.31-1.01	0.05	0.89	0.48-1.65	0.71

Abbreviations: UTMDACC, The University of Texas MD Anderson Cancer Center, USA; ref, reference; HPV, human papillomavirus; BOT, base of tongue; RT, radiotherapy; CRT, chemoradiotherapy.

^b Significant value.

^c Adjusted for center, age group, gender, HPV status, smoking status, T-site location, stage UICC8 and treatment regimen.

tongue (BOT) (48.6 %) followed by the palatine tonsils (47.2 %) while the palatine tonsils were the predominant tumor site in the Copenhagen cohort (51.6 %). OPSCC patients presented at the UTMDACC at significantly lower T-stage (T1-T2: 72.9 % vs. 64.9 %), lower N-stage (N0-N1: 76.8 % vs. 73.3 %) and lower UICC stage (I-II: 78.4 % vs. 69.2 %) than in Copenhagen. See [Table 1](#).

3.3. HPV status

Overall, 63.1 % of the OPSCC patients were HPV+ in the Copenhagen cohort, while 88.2 % were HPV+ in the UTMDACC cohort. In the UTMDACC cohort, 68.9 % did not have both HPV and p16 status available, while this was the case in 3.1 % of the patients in the Copenhagen cohort. In the Copenhagen cohort, 0.4 % and 10.8 % in the UTMDACC were tested for HPV DNA only, while 1.9 % in the Copenhagen cohort and 46.7 % in the UTMDACC cohort were tested for p16 only. See [Table 1](#).

3.4. Treatment modalities

Radiation-based treatment was predominant in both cohorts, but significantly more patients received radiotherapy (RT) alone in the Copenhagen cohort (30.3 % vs. 14.8 %). In the Copenhagen cohort, 51.0 % received concurrent chemotherapy (CRT) vs. 49.8 % in the UTMDACC cohort. Neoadjuvant chemotherapy + RT/CRT was given to 16.7 % of patients in the UTMDACC cohort. Significantly more patients received surgery alone in the Copenhagen cohort (16.0 % vs. 8.7 %), while significantly more received surgery and post-operative adjuvant therapy in the UTMDACC cohort (9.8 % vs. 2.7 %). See [Table 1](#).

3.5. Overall survival

Kaplan-Meier OS models showed significant differences in the 3-year OS for the UTMDACC cohort (91 % [95 % CI: 89–93 %]) compared to the Copenhagen cohort (83 %, [95 % CI: 80–85 %], log-rank $p < 0.001$) which was also noted for HPV-negative (HPV-) patients. See [Fig. 1](#) and [supplementary Table 2](#) (Supplement 1).

The multivariable OS analysis revealed no significant difference between the two cohorts (hazard ratio [HR] 1.21, $p = 0.23$). A significantly increased HR was observed for HPV-, current smoking, T3- and T4-class, N2 and N3-class, M1-class, and RT. See [Table 2](#).

3.6. Recurrence-free interval

Overall, 83 OPSCC patients had a recurrence in the UTMDACC cohort, with a median time to recurrence of 1.00 years (IQR: 0.63–1.91 years). In the Copenhagen cohort, 184 had a recurrence with a median time to recurrence of 1.04 years (IQR: 0.69–1.61). The 3-year RFI was significantly better in the UTMDACC cohort (91.3 % [95 % CI 89.3–93.3 %]) compared to the Copenhagen cohort (82.7 % [95 % CI 80.3–85.1 %]), log-rank $p < 0.001$, which was also found when stratifying for HPV-status. See [Fig. 2](#).

The multivariable recurrence analysis showed a significantly increased HR in the Danish cohort (HR: 1.74, $p = 0.003$). Also, HPV-, T2-T4-class, N1-N3-class, M1-class, RT and neoadjuvant chemotherapy + RT/CRT had a significantly increased HR. See [Table 3](#). The analysis was also performed for patients with either progression or second primary tumors, which did not significantly impact the conclusions; data not shown.

3.7. Subgroup analysis of low-risk patients with T1/T2N0M0 OPSCC

The UTMDACC T1T2N0M0 cohort exhibited significantly more men, younger ages, more never smokers, more HPV+, fewer OPSCC from other locations, and more received CRT than the Danish T1T2N0M0 cohort. See [Table 3](#) ([Supplement 1](#)). In total, 26 had a recurrence with a median time to recurrence of 1.07 years (IQR 0.82–1.47) and 0.76 years (IQR 0.63–1.75) in the Copenhagen and the UTMDACC T1T2N0M0 cohorts, respectively.

Kaplan-Meier models showed a significantly higher 3-year OS in the UTMDACC T1T2N0M0 cohort (89.9 % vs. 79.3 %, log-rank $p = 0.01$) and 3-year RFI (96.2 % vs. 86.5 %, log-rank $p = 0.011$). See [Fig. 1](#) (Supplement 1). These differences were not reproduced in the

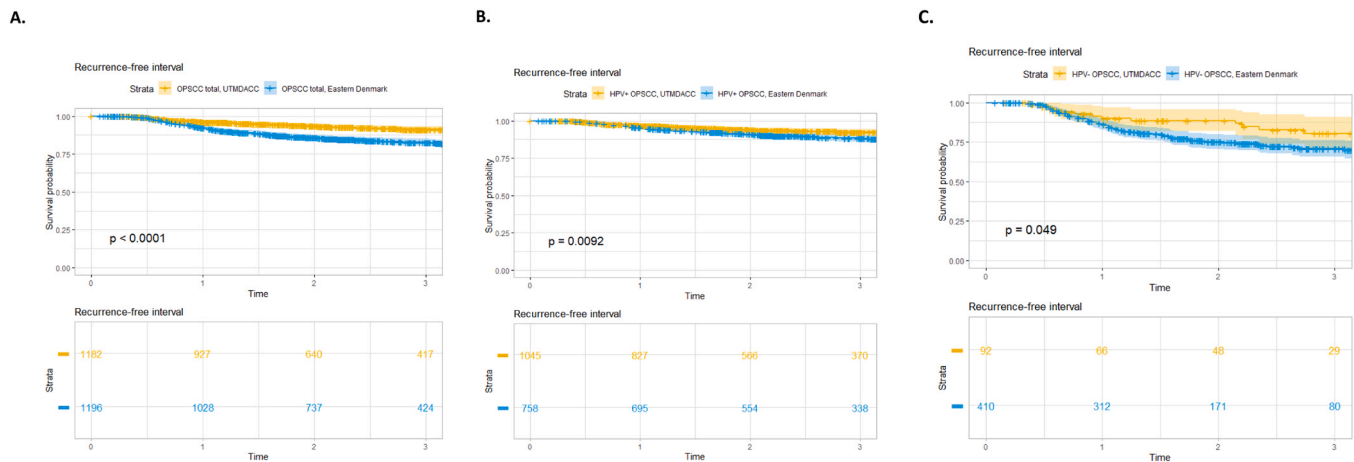


Fig. 2. Kaplan-Meier curves depicting the recurrence-free interval stratified by center and HPV status from 2015–2020. A. All OPSCC 2015–2020 stratified by center. B. HPV+ OPSCC stratified by center. C. HPV- OPSCC stratified by center. *Abbreviations:* UTMDACC, The University of Texas MD Anderson Cancer Center, USA, HPV, human papillomavirus; OPSCC, oropharyngeal squamous cell carcinoma.

multivariable analysis for both OS and RFI. See Table 4 and 5 (Supplement 1).

3.8. Subgroup analysis of high-risk patients with stage III-IV OPSCC

The UTMDACC III-IV cohort exhibited significantly more men, younger age, more never smokers, more HPV+, fewer other OPSCC locations, higher UICC-stage for HPV+ and fewer received RT. See Table 6 (Supplement 1). In total, 126 had a recurrence with a median time to recurrence of 0.88 years (IQR 0.59–1.32) and 1.19 (IQR: 0.54–1.71) in the Copenhagen and the UTMDACC III-IV cohorts, respectively.

Kaplan-Meier models showed a significantly higher 3-year OS in the UTMDACC III-IV cohort (77.1 % vs. 52.4 %, log-rank $p < 0.001$) and 3-year RFI (78.6 % vs. 65 %, log-rank $p = 0.001$). See Fig. 2 (Supplement 1). These differences were reproduced in the multivariable analysis for both OS (HR 2.20, $p = 0.004$) and RFI (HR 2.80, $p = 0.002$). See Table 7 and 8 (Supplement 1).

3.9. Subgroup analysis of RT single modality patients

See Results (Supplement 1).

4. Discussion

This study, which included 2484 patients with OPSCC from Eastern Denmark and UTMDACC treated between 2015–2020, revealed significant differences in OS and RFI driven by clinical and treatment differences.

Although RT was predominant in both cohorts, significantly more patients received RT alone in the Copenhagen cohort, which is noteworthy, since the Copenhagen cohort presented at higher UICC-stage, T- and N-class. In line with this, RT remains the predominant treatment modality in most European centers [19].

The UTMDACC cohort included more clinical characteristics associated with a better prognosis: more HPV+ OPSCC patients, lower T- and N-burden and less smoking [20] but also more men which are associated with a worse prognosis [21]. When adjusting for these differences, an independent increased OS HR was not observed, but the risk of recurrence was significantly higher in the Copenhagen cohort (RFI HR 1.74, $p = 0.003$). Discrepancies in follow-up regimens and divergent definitions of recurrence may also attribute to the observed disparities. Although speculative, it could also indicate a different biology of HPV+ OPSCCs. In line with our findings, a recent study including a Danish and a Toronto cohort observed significant clinical and treatment

differences, which were reflected in differences in locoregional failure and OS [22].

In low-risk patients, significant clinical and treatment differences were observed with 41.7 % in the Copenhagen cohort receiving RT alone versus 19.6 % in the UTMDACC cohort, yet the prognosis remained equally good. Several studies have investigated de-escalated treatment to minimize treatment-related toxicities for a subgroup of patients with HPV+ OPSCC [8,9,23–25]. It is important to keep in mind that substantial clinical differences exist when selecting patients and validating de-escalating trials in various geographical areas. This study underlines the importance of including multiple factors like TNM-classification, HPV- and smoking status, but also indicates that factors like age and tumor location could be considered for stratification.

Conversely, a sub-analysis of high-risk patients revealed significant clinical and treatment differences influencing the prognosis. The UTMDACC cohort had significantly better OS and RFI. OS was also significantly associated with current smoking, stage, and treatment modality given. RT alone was given to a substantial part of the Copenhagen patients (37.1 % vs 7.2 %), despite that CRT is recommended for this group [16,17]. The difference in use of concurrent chemotherapy is likely a result of factors impacting treatment selection not included in this study, e.g., comorbidities, as the Copenhagen cohort is older and include more smokers. But a difference in practice patterns cannot be excluded. Neoadjuvant chemotherapy is not used in Denmark but was given to 16.7 % of the patients in the UTMDACC cohort (corresponding to 39.1 % of UTMDACC high-risk patients), which is in line with data from The National Cancer Database [26]. The role of neoadjuvant therapy in locally advanced OPSCC cancer remains controversial [27], however, it may be associated with a decreased risk of distant metastasis [28]. A recent study suggests that neoadjuvant therapy is associated with an improved OS and lower risk of distant metastasis in patients with OPSCC [29].

A notable difference in treatment between MDA and Denmark is the use of hypoxic modification with nimorazole. The DAHANCA 5 randomized trial, investigated the effect of nimorazole, showed a significantly better loco-regional control rate and lower cancer-related death in patients receiving nimorazole compared to placebo [30]. Nimorazole has since been standard of care for all Danish OPSCC patients. The benefit of nimorazole may primarily be present in HPV- OPSCC, as reanalysis showed no significant benefit in HPV/p16-positive tumors [31].

Disparities in the healthcare system might introduce a selection bias contributing to the differences observed in this study, with more HPV- OPSCC patients and more high-risk patients in the Copenhagen cohort.

Table 3
Multi- and univariable analysis for recurrence-free interval for OPSCC in Eastern Denmark and UTMDACC from 2015–2020.

Variable	Univariable			Multivariable ^e		
	HR	95 % CI	p-value ^d	HR	95 % CI	p-value ^d
Center (UTMDACC ref)						
Eastern Denmark	2.11	1.63–2.74	< 0.001 ^d	1.74	1.21–2.50	0.003 ^d
Age (<60 years ref)						
> 60 years	1.53	1.20–1.96	< 0.001 ^d	1.08	0.83–1.42	0.56
Gender (female ref)						
Male	1.01	0.75–1.38	0.93	1.19	0.86–1.64	0.30
HPV-status (HPV+ ref)						
HPV-	3.30	2.58–4.24	< 0.001 ^d	2.10	1.50–2.96	< 0.001 ^d
Smoking status (never ref)						
previous	1.55	1.13–2.12	0.01 ^d	1.11	0.80–1.55	0.53
current	3.12	2.27–4.28	< 0.001 ^d	1.31	0.88–1.95	0.19
Tumor location (tonsil ref)						
BOT	1.04	0.79–1.37	0.76	0.96	0.72–1.28	0.78
Other	2.44	1.77–3.36	< 0.001 ^d	1.12	0.78–1.63	0.53
T-class (T1 ref)						
T2	1.73	1.20–2.48	0.003 ^d	1.71	1.17–2.49	0.005 ^d
T3	3.11	2.12–4.58	< 0.001 ^d	2.68	1.75–4.09	< 0.001 ^d
T4	4.16	2.86–6.04	< 0.001 ^d	3.12	2.05–4.75	< 0.001 ^d
N-class (N0 ref)						
N1	0.73	0.52–1.04	0.08	1.67	1.13–2.47	0.01 ^d
N2	2.07	1.45–2.96	< 0.001 ^d	2.49	1.66–3.72	< 0.001 ^d
N3	3.17	1.85–5.43	< 0.001 ^d	4.07	2.27–7.32	< 0.001 ^d
M-class (M0 ref)						
M1	3.49	1.65–7.41	0.001 ^d	3.22	1.46–7.10	0.004 ^d
Treatment regimen (CRT ref)						
RT	2.19	1.66–2.89	< 0.001 ^d	1.98	1.45–2.70	< 0.001 ^d
Neoadjuvant chemotherapy + RT/CRT	1.86	1.24–2.77	0.002 ^d	2.18	1.32–3.60	0.002 ^d
Surgery	0.90	0.58–1.40	0.64	1.64	0.99–2.71	0.05
Surgery + RT/CRT	0.60	0.30–1.18	0.14	0.96	0.46–2.01	0.91

Abbreviations: UTMDACC, The University of Texas MD Anderson Cancer Center, USA; ref, reference; HPV, human papillomavirus; BOT, base of tongue; RT, radiotherapy; CRT, chemoradiotherapy.

^d Significant value.

^e Adjusted for center, age group, gender, HPV status, smoking status, T-site location, stage UICC8 and treatment regimen.

Studies have shown that insurance status is predictive of clinical characteristics such as tumor stage and comorbidities at diagnosis as well as oncological outcome [32–34]. A recent Copenhagen study comprising a part of the Copenhagen cohort suggests that low socioeconomic status negatively impact the OS, largely due to differences in clinical characteristics at diagnosis, including smoking status, comorbidities, and clinical stage [35]. Future studies including socioeconomic status,

comorbidities, and performance status would provide further insight into this matter. Disparities in the healthcare system might also contribute to the differences observed in treatment modalities, availability, and regimen of follow-up care.

4.1. Limitations

A recent study, where the Copenhagen cohort comprised the majority of the study cohort, has proven double HPV/p16-positivity to be superior in prognostication and has led to the recommendation of including both p16 and HPV-status in areas with high discordance, which is also supported by the ESMO guidelines [4,5]. In this study, HPV-positivity was defined in the UTMDACC as p16 + or HPV+ , which might overestimate the HPV-positive prevalence, although only 1.9 % had discordant p16/HPV status [4]. The included cohorts might not be representative of the Northern American and European cohorts overall, and the results may not apply to other regions. Socioeconomic status, performance score, and comorbidities were not included which potentially impacts treatment decisions and have shown to influence both OS and RFI and which we hypothesize differs greatly in the two cohorts.

5. Conclusion

Significant demographic, clinical, and treatment differences influencing the prognosis were identified between two high HPV-prevalence OPSCC cohorts representing two different health care systems: a Copenhagen cohort from a public, universal health care system and a UTMDACC cohort from an insurance-based health care system. Despite significant clinical and treatment differences, the prognosis for low-risk OPSCC patients was equally good, while the prognosis was significantly better for patients with high-risk OPSCC in the UTMDACC cohort.

Our study suggests that significant clinical differences exist between a Northern European and Northern American OPSCC population, which is important to consider when comparing outcomes and for patient stratification in clinical trials, as reproducibility might be challenging. However, to fully understand possible intercontinental variations, more data are needed.

Role of the funding source

The funder had no role in study design, in the collection, analysis or interpretation of data, in the writing of the report or in the decision to submit this article for publication.

Declaration of Interest statement

None declared.

Funding

ALFC is support by the private fund Candys Foundation, Denmark (reference: 2020-352).

CRedit authorship contribution statement

Christian Grønhoj: Conceptualization, Writing – review & editing. **Martin Gasset-Zamani:** Data curation, Investigation, Writing – review & editing. **Faye M. Johnson:** Writing – review & editing. **Katherine Hutcheson:** Writing – review & editing. **Amy C. Moreno:** Conceptualization, Resources, Supervision, Writing – review & editing. **Clifton D. Fuller:** Project administration, Writing – review & editing. **Neil D. Gross:** Writing – review & editing. **Simone Kloch Bendtsen:** Project administration, Writing – review & editing. **Toyin Babarinde:** Conceptualization, Writing – review & editing. **Amanda-Louise F. Carlander:** Conceptualization, Data curation, Formal analysis, Investigation, Methodology, Project administration, Validation, Visualization,

Writing – original draft, Writing – review & editing. **Christian von Buchwald**: Conceptualization, Resources, Supervision, Writing – review & editing. **Kathrine Kronberg Jakobsen**: Methodology, Writing – review & editing. **Jeffrey N. Myers**: Conceptualization, Resources, Supervision, Writing – review & editing. **Jacob Høyer Rasmussen**: Conceptualization, Methodology, Writing – review & editing. **Jeppe Friberg**: Conceptualization, Writing – review & editing.

Declaration of Competing Interest

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

Appendix A. Supporting information

Supplementary data associated with this article can be found in the online version at [doi:10.1016/j.ejca.2024.113983](https://doi.org/10.1016/j.ejca.2024.113983).

References

- Ferlay J, Colombet M, Soerjomataram I, et al. Estimating the global cancer incidence and mortality in 2018: GLOBOCAN sources and methods. *Int J Cancer* 2019;144(8):1941–53. <https://doi.org/10.1002/ijc.31937>.
- Chaturvedi AK, Anderson WF, Lortet-Tieulent J, et al. Worldwide trends in incidence rates for oral cavity and oropharyngeal cancers. *J Clin Oncol* 2013;31(36):4550–9. <https://doi.org/10.1200/JCO.2013.50.3870>.
- Carlander AF, Jakobsen KK, Bendtsen SK, et al. A contemporary systematic review on repartition of HPV-positivity in oropharyngeal cancer worldwide. *Viruses* 2021; 13(7). <https://doi.org/10.3390/v13071326>.
- Mehanna H, Taberna M, von Buchwald C, et al. Prognostic implications of p16 and HPV discordance in oropharyngeal cancer (HNCIG-EPIC-OPC): a multicentre, multinational, individual patient data analysis. *Lancet Oncol* 2023;239–51. [https://doi.org/10.1016/S1470-2045\(23\)00013-X](https://doi.org/10.1016/S1470-2045(23)00013-X).
- Machiels JP, René Leemans C, Golusinski W, Grau C, Licitra L, Gregoire V. Squamous cell carcinoma of the oral cavity, larynx, oropharynx and hypopharynx: EHSN-ESMO-ESTRO Clinical Practice Guidelines for diagnosis, treatment and follow-up. *Ann Oncol* 2020;31(11):1462–75. <https://doi.org/10.1016/j.annonc.2020.07.011>.
- Ang KK, Harris J, Wheeler R, et al. Human papillomavirus and survival of patients with oropharyngeal cancer. *N Engl J Med* 2010;363(1):24–35. <https://doi.org/10.1056/NEJMoa0912217>.
- Buchberger AMS, Strzelczyk EA, Wollenberg B, Combs SE, Pickhard A, Pigorsch SU. Report on late toxicity in head-and-neck tumor patients with long term survival after radiochemotherapy. *Cancers* 2021;13(17). <https://doi.org/10.3390/cancers13174292>.
- Gillison ML, Trotti AM, Harris J, et al. Radiotherapy plus cetuximab or cisplatin in human papillomavirus-positive oropharyngeal cancer (NRG Oncology RTOG 1016): a randomised, multicentre, non-inferiority trial. *Lancet (Lond Engl)* 2019; 393(10166):40–50. [https://doi.org/10.1016/S0140-6736\(18\)32779-X](https://doi.org/10.1016/S0140-6736(18)32779-X).
- Palma DA, Prisman E, Berthelet E, et al. Assessment of toxic effects and survival in treatment deescalation with radiotherapy vs transoral surgery for HPV-associated oropharyngeal squamous cell carcinoma: the ORATOR2 Phase 2 Randomized Clinical Trial. *JAMA Oncol* 2022;8(6):1–7. <https://doi.org/10.1001/jamaoncol.2022.0615>.
- Garset-Zamani M, Carlander AF, Jakobsen KK, et al. Impact of specific high-risk human papillomavirus genotypes on survival in oropharyngeal cancer. *Int J Cancer* 2022;150(7):1174–83. <https://doi.org/10.1002/ijc.33893>.
- Chaturvedi AK, Engels EA, Pfeiffer RM, et al. Human papillomavirus and rising oropharyngeal cancer incidence in the United States. *J Clin Oncol* 2011;29(32): 4294–301. <https://doi.org/10.1200/JCO.2011.36.4596>.
- von Elm E, Altman DG, Egger M, Pocock SJ, Gøtzsche PC, Vandenbroucke JP. The Strengthening of Reporting of Observational Studies in Epidemiology (STROBE) statement: guidelines for reporting observational studies. *J Clin Epidemiol* 2008;61(4):344–9. <https://doi.org/10.1016/j.jclinepi.2007.11.008>.
- Garnaes E, Kiss K, Andersen L, et al. A high and increasing HPV prevalence in tonsillar cancers in Eastern Denmark, 2000-2010: the largest registry-based study to date. *Int J Cancer J Int du Cancer* 2014;00(July):1–8. <https://doi.org/10.1002/ijc.29254>.
- Carlander ALF, Grønhoj Larsen C, Jensen DH, et al. Continuing rise in oropharyngeal cancer in a high HPV prevalence area: a Danish population-based study from 2011 to 2014. *Eur J Cancer* 2017;70:75–82. <https://doi.org/10.1016/j.ejca.2016.10.015>.
- Zamani M, Grønhoj C, Jensen DH, et al. The current epidemic of HPV-associated oropharyngeal cancer: an 18-year Danish population-based study with 2,169 patients. *Eur J Cancer* 2020;134(2020):52–9. <https://doi.org/10.1016/j.ejca.2020.04.027>.
- Dansk Selskab for Hoved- og Hals Onkologi (DSHHO) DDHHC (DAHANCA). Nationale Retningslinjer for Udredning, Behandling, Rehabilitering og Kontrolforløb for Patienter med Pharynx- og Larynx-cancer i Danmark; 2011; (april):1–69.
- Jensen K, Friberg J, Hansen CR, et al. The Danish Head and Neck Cancer Group (DAHANCA) 2020 radiotherapy guidelines. *Radiother Oncol: J Eur Soc Ther Radiol Oncol* 2020;151:149–51. <https://doi.org/10.1016/j.radonc.2020.07.037>.
- National Comprehensive Cancer Network Clinical Practice Guidelines in Oncology: Head and Neck Cancers; 2023;(Version 2.2023).
- Culié D, Garrel R, Viotti J, et al. Impact of HPV-associated p16-expression and other clinical factors on therapeutic decision-making in patients with oropharyngeal cancer: A GETTEC multicentric study. *Eur J Surg Oncol: J Eur Soc Surg Oncol Br Assoc Surg Oncol* 2018;44(12):1908–13. <https://doi.org/10.1016/j.ejso.2018.05.022>.
- Larsen C, Jensen D, Carlander AF, et al. Novel nomograms for survival and progression in HPV+ and HPV- oropharyngeal cancer: a population-based study of 1,542 consecutive patients. *Oncotarget* 2016;7(33). <https://doi.org/10.18632/oncotarget.12335>.
- Licitra L, Zigon G, Gatta G, Sánchez MJ, Berrino F. Human papillomavirus in HNSCC: a European epidemiologic perspective. *Hematol Oncol Clin N Am* 2008;22(6):1143–53. <https://doi.org/10.1016/j.hoc.2008.10.002>.
- Lassen P, Huang SH, Su J, et al. Treatment outcomes and survival following definitive (chemo)radiotherapy in HPV-positive oropharynx cancer: Large-scale comparison of DAHANCA vs PMH cohorts. *Int J Cancer* 2022;150(8):1329–40. <https://doi.org/10.1002/ijc.33876>.
- Yom SS, Torres-Saavedra P, Caudell JJ, et al. Reduced-dose radiation therapy for HPV-associated oropharyngeal carcinoma (NRG Oncology HN002). *J Clin Oncol: J Am Soc Clin Oncol* 2021;39(9):956–65. <https://doi.org/10.1200/JCO.20.03128>.
- Rischin D, King M, Kenny L, et al. Randomized trial of radiation therapy with weekly cisplatin or cetuximab in low-risk hpv-associated oropharyngeal cancer (TROG 12.01) - a trans-tasman radiation oncology group study. *Int J Radiat Oncol Biol Phys* 2021;111(4):876–86. <https://doi.org/10.1016/j.ijrobp.2021.04.015>.
- Mehanna H, Robinson M, Hartley A, et al. Radiotherapy plus cisplatin or cetuximab in low-risk human papillomavirus-positive oropharyngeal cancer (De-ESCALaTE HPV): an open-label randomised controlled phase 3 trial. *Lancet (Lond Engl)* 2019; 393(10166):51–60. [https://doi.org/10.1016/S0140-6736\(18\)32752-1](https://doi.org/10.1016/S0140-6736(18)32752-1).
- Sher DJ, Rusthoven CG, Khan SA, Fidler MJ, Zhu H, Koshy M. National patterns of care and predictors of neoadjuvant and concurrent chemotherapy use with definitive radiotherapy in the treatment of patients with oropharyngeal squamous cell carcinoma. *Cancer* 2017;123(2):273–82. <https://doi.org/10.1002/cncr.30255>.
- Pignon JP, le Maître A, Maillard E, Bourhis J. Meta-analysis of chemotherapy in head and neck cancer (MACH-NC): an update on 93 randomised trials and 17,346 patients. *Radiother Oncol: J Eur Soc Ther Radiol Oncol* 2009;92(1):4–14. <https://doi.org/10.1016/j.radonc.2009.04.014>.
- Ma J, Liu Y, Huang XL, et al. Induction chemotherapy decreases the rate of distant metastasis in patients with head and neck squamous cell carcinoma but does not improve survival or locoregional control: a meta-analysis. *Oral Oncol* 2012;48(11): 1076–84. <https://doi.org/10.1016/j.oraloncology.2012.06.014>.
- Guo TW, Saiyed F, Yao CMKL, et al. Outcomes of patients with oropharyngeal squamous cell carcinoma treated with induction chemotherapy followed by concurrent chemoradiation compared with those treated with concurrent chemoradiation. *Cancer* 2021;127(16):2916–25. <https://doi.org/10.1002/cncr.33491>.
- Overgaard J, Hansen HS, Overgaard M, et al. A randomized double-blind phase III study of nimorazole as a hypoxic radiosensitizer of primary radiotherapy in supraglottic larynx and pharynx carcinoma. Results of the Danish Head and Neck Cancer Study (DAHANCA) Protocol 5-85. *Radiol Oncol* 1998;46(2):135–46. [https://doi.org/10.1016/s0167-8140\(97\)00220-x](https://doi.org/10.1016/s0167-8140(97)00220-x).
- Lassen P, Eriksen JG, Hamilton-Dutoit S, Tramm T, Alsner J, Overgaard J. HPV-associated p16-expression and response to hypoxic modification of radiotherapy in head and neck cancer. *Radiol Oncol* 2010;94(1):30–5. <https://doi.org/10.1016/j.radonc.2009.10.008>.
- Kwok J, Langevin SM, Argiris A, Grandis JR, Gooding WE, Taioli E. The impact of health insurance status on the survival of patients with head and neck cancer. *Cancer* 2010;116(2):476–85. <https://doi.org/10.1002/cncr.24774>.
- Rohlfing ML, Mays AC, Isom S, Waltonen JD. Insurance status as a predictor of mortality in patients undergoing head and neck cancer surgery. *Laryngoscope* 2017;127(12):2784–9. <https://doi.org/10.1002/lary.26713>.
- Shin JY, Yoon JK, Shin AK, Blumenfeld P, Mai M, Diaz AZ. Association of insurance and community-level socioeconomic status with treatment and outcome of squamous cell carcinoma of the pharynx. *JAMA Otolaryngol- Head Neck Surg* 2017;143(9):899–907. <https://doi.org/10.1001/jamaoto.2017.0837>.
- Olsen MH, Frederiksen K, Lassen P, et al. Association of smoking, comorbidity, clinical stage, and treatment intent with socioeconomic differences in survival after oropharyngeal squamous cell carcinoma in Denmark. *JAMA Netw Open* 2022;5(12):e2245510. <https://doi.org/10.1001/jamanetworkopen.2022.45510>.

Supplement 1

1. Methods and Materials

Variables

The last day of follow-up for the Copenhagen cohort was defined as the last clinical visit, death, or recurrence; and for the UTMDACC cohort as the last imaging, death, or recurrence. Recurrence was defined as local, regional and/or distant. Head and neck cancer within the first 6 months after primary OPSCC was considered progression, i.e., not a recurrence in both cohorts. For the Copenhagen cohort, a new diagnosis of OPSCC identified from six months to five years after primary diagnosis and treatment was considered a recurrence and not a second primary unless specifically stated otherwise in the medical record. For the UTMDACC cohort, the distinction between recurrence or second primary was determined by the treating team based on proximity to the initial cancer and time from treatment (<5 years).

Measurement data

Age was grouped as <60 years and >60 years. Patient-reported smoking status at the time of diagnosis was grouped as never smoking/former smoking/current smoking. HPV-positivity was defined as being positive to both p16 immunohistochemistry and HPV DNA in the Copenhagen cohort, while HPV-positivity was defined as being positive to either p16 or HPV DNA/RNA. Clinical stage was defined according to the American Joint Committee on Cancer/Union for International Cancer Control (AJCC/UICC) TNM classification system (8th edition). For p16- tumors, stages IVA-C were merged to IV. Treatment regimen as assigned at a multidisciplinary treatment planning conference and grouped as radiotherapy (RT)/chemoradiotherapy (CRT)/neoadjuvant chemotherapy + RT/CRT/surgery/surgery +RT/CRT. Tumor location was grouped as palatine tonsils/base of tongue/other OPSCC location. Other oropharyngeal sites included in the Copenhagen cohort were pharyngeal wall, soft palate, uvula or pharyngeal arch, while sites included in the UTMDACC cohort were soft palate, pharyngeal wall and glossopharyngeal sulcus.

HPV detection and p16 immunohistochemistry (IHC)

For the Copenhagen cohort, HPV DNA was detected with PCR as previously described from 2015-2017¹³⁻¹⁵. From 2018-2020 the presence of HPV DNA and HPV genotypes was evaluated by VisionArray HPV chip 1.0 (ZytoVision). p16 IHC was carried out using Ventana Benchmark Ultra auto-stainer with the UltraView detection kit and the p16 monoclonal antibody E6H4 ready-to-use with CC1 as pretreatment (Roche,

Tuscon, USA). The p16-positivity cutoff was set at >70% for both nuclear and cytoplasmic staining. HPV genotypes was detected with next generation sequencing from 2000-2017 as previously described¹³⁻¹⁵.

For the Stiefel Database, the presence of HPV virus was detected by HPV DNA in situ hybridization, HPV RNA in situ hybridization, cytology genotyping, or HPV DNA PCR. The p16-positivity was assessed by IHC.

Treatment

In Denmark, curative radiation-based therapy is the primary treatment and consists of moderately accelerated radiotherapy (RT) given in 33-34 fractions 6 days a week with or without concurrent chemotherapy (CRT). The standard dose levels are 66-68 Gray (Gy) to the clinical target volume (CTV) 1, and 60 Gy to the CTV2. Dose level to elective targets is 50 Gy. According to guidelines, CRT is given based on an evaluation of performance status and comorbidities and consists mainly of weekly cisplatin (40mg/m²). Carboplatin is given if cisplatin is not tolerated. Very few patients received cetuximab. Induction therapy is not used in Denmark. Trans-oral robotic surgery (TORS) is considered for patients in clinical trials or if primary RT cannot be performed. In Denmark, all patients were also offered concurrent nimorazole, a radiosensitizer, unless there were contraindications. Unilateral RT could be offered for well-lateralized tonsil carcinomas without involvement of midline structures^{16,17}.

At UTMDACC, radiation therapy for OPSCC is generally delivered alone or with concurrent systemic therapy over the course of 6 to 7 weeks (i.e., 33-34 fractions) to a curative total dose of 60-70 Gy. The standard dose levels include 66-70 Gy to CTV1 (i.e., gross tumor with anatomically restricted expansion), 60-63 Gy to adjacent at-risk primary sites, involved neck levels or postoperative volumes, and 54-57 Gy to elective clinical targets (i.e., elective nodal irradiation volumes). Unilateral RT is reserved for well-lateralized clinical T1-T2 tonsil carcinomas with no tongue base involvement, <1cm soft palate involvement, and minimal adenopathy. Concurrent chemotherapy is primarily given as weekly cisplatin (40 mg/m²), which can be switched to carboplatin if cisplatin is not tolerated. Currently, few patients who are ineligible to cisplatin receive cetuximab. TORS is considered for all patients with unilateral, low-volume disease (T0-2, N0-1) OPSCC¹⁸.

Statistics

Subgroup analyses were performed for both low-risk patients (T1/T2N0M0) and high-risk patients (stage III-IV). For the T1T2N0M0 analyses the multivariable OS analysis was adjusted for center, HPV-status, smoking

status, T-site location, and stage UICC8 based on clinical relevance and due a limited number of events (50). Likewise, the multivariable RFI analysis was adjusted for center, HPV status and stage UICC8 based on clinical relevance and due a limited number of events (26). For the III-IV analyses, the multivariable OS analysis (192 events) was adjusted for center, HPV-status, smoking status, stage UICC8 and treatment regimen while the multivariable RFI (126 events) analysis was adjusted for center, HPV-status, stage UICC8, treatment regimen based on clinical relevance.

2. Results

Subgroup analysis of patients receiving RT as single modality

The Copenhagen RT single modality cohort comprised significantly more men (27.2% vs 12.2%, $p<0.001$), more HPV- OPSCC (47.6% vs 9.0%, $p<0.001$), more other OPSCC location (23.1% vs 5.9%, $p<0.001$), older age (mean age 68.4 vs 61.3 years, $p<0.001$), more current smokers (45.1% vs 7.5%, $p<0.001$) and more stage III-IV (35.9% vs 8.3%, $p<0.001$). 3-year OS in the Copenhagen RT single modality cohort was 61.6% (95% CI: 56.2-67.5%) and in the UTMDACC RT single modality cohort 88.6% (95% CI: 83.4-94.1%), log-rank $p=0.001$. Data not shown.

Table 2. 1- and 3-year overall survival estimates stratified by center and HPV-status from 2015-2020.

Overall survival 2015-2020	Eastern Denmark	UTMDACC, USA
OS total	% (95% CI)	% (95% CI)
1-year	92% (91-94%)	96% (95-97%)
3-year	83% (80-85%)	91% (89-93%)
OS HPV+		
1-year	96% (94-97%)	97% (96-98%)
3-year	88% (86-91%)	93% (91-95%)
OS HPV-		
1-year	85% (81-88%)	96% (92-100%)
3-year	56% (51-62%)	78% (68-89%)

Abbreviations: UTMDACC, The University of Texas MD Anderson Cancer Center, USA; OS, overall survival; HPV, Human Papillomavirus.

Table 3. Characteristics of 299 patients with T1/T2N0M0 OPSCC in Eastern Denmark and UTMDACC, Texas, USA from 2015-2020.

Variable	Eastern Denmark, n=192		UTMDACC, Texas USA, n=107		p-value*
	no.	%	no.	%	
Gender					<0.001*
Male	126	65.6	80	74.8	
Female	66	34.4	27	25.2	
Median age (IQR)	65 (59-72)		61 (55-67)		<0.001*
no.	192		107		
Median follow-up. years (IQR)	2.2 (1.4-3.2)		1.9 (0.8-3.5)		0.34
no.	192		107		
Smoking					<0.001*
Current	108	56.3	5	4.7	
Former	61	31.8	40	37.4	
Never	22	11.5	60	56.1	
Unknown	1	0.5	2	1.9	
HPV-status					<0.001*
HPV+	69	35.9	91	85.0	
HPV-	121	63.0	14	13.1	
Unknown	2	1.0	2	1.9	
Tumor location					<0.001*
BOT	24	12.5	33	30.8	
Tonsils	84	43.8	64	59.8	
Other	84	43.8	10	9.3	
TNM Stage (UICC/AJCC 8), HPV+					
T-class					1
T1	20	29.0	27	29.7	
T2	49	71.0	64	70.3	
TNM Stage (UICC/AJCC 8), HPV-					
T-class					0.68
I	55	45.5	5	35.7	
II	66	54.5	9	64.3	
Overall stage (UICC/AJCC8), HPV-					0.48
I	60	49.6	5	35.7	
II	61	50.4	9	64.3	
Treatment					<0.001*
CRT	9	4.7	25	23.4	
RT	80	41.7	21	19.6	
Neoadjuvant chemotherapy + RT/CRT	-	-	1	0.9	
Surgery	101	52.6	45	42.1	

Surgery + RT/CRT

2

1

15

14

Note: Frequency (%) is provided for categorical variables, median (IQR) are provided for continuous variables. Chi Square test were used for categorical variables, while t-test was used for continuous variables. Fisher's exact test was used for small sample sizes.

Abbreviations: UTMDACC, The University of Texas MD Anderson Cancer Center, USA; IQR, interquartile range; HPV, Human Papillomavirus; BOT, base of tongue; RT, radiotherapy; CRT, concurrent RT + systemic therapy.

*Significant p-value

Table 4. Multi- and univariable analysis for overall survival for T1/T2N0M0 OPSCC in Eastern Denmark and UTMDACC from 2015-2020.

Variable	Univariable			Multivariable**		
	HR	95% CI	p-value*	HR	95% CI	p-value*
Center (UTMDACC ref)						
Eastern Denmark	2.63	1.23-5.62	0.01*	0.94	0.36-2.42	0.90
Age (<60 years ref)						
>60 years	1.83	0.55-0.99	0.06	-		
Gender (female ref)						
Male	1.19	0.64-2.21	0.58	-		
HPV-status (HPV+ ref)						
HPV-	3.96	2.11-7.40	<0.001*	1.92	0.70-5.21	0.20
Smoking status (never ref)						
Previous	0.96	0.36-2.58	0.93	0.74	0.26-2.11	0.58
Current	3.97	1.75-9.01	<0.001*	2.11	0.71-6.27	0.18
Tumor location (tonsils ref)						
BOT	0.81	0.30-2.19	0.67	0.72	0.26-2.0	0.53
Other	2.99	0.163-5.49	<0.001*	1.40	0.65-3.0	0.39
UICC stage (I ref)						
UICC II	2.56	1.45-4.49	0.001*	0.98	0.49-1.94	0.94
Treatment (CRT ref)						
RT	1.73	0.66-4.54	0.26	-		
Surgery	0.78	0.29-2.1	0.62			
Surgery + RT/CRT	0.85	0.16-4.36	0.84			

Abbreviations: UTMDACC, The University of Texas MD Anderson Cancer Center, USA; ref, reference; HPV, Human Papillomavirus; BOT, base of tongue; RT, radiotherapy; CRT, concurrent RT + systemic therapy.

*Significant p-value. **Adjusted for center, HPV-status, smoking status, tumor location and stage UICC8.

Table 5. Multi- and univariable analysis for recurrence-free interval for T1/T2N0M0 OPSCC in Eastern Denmark and UTMDACC from 2015-2020.

Variable	Univariable			Multivariable**		
	HR	95% CI	p-value*	HR	95% CI	p-value*
Center (UTMDACC ref)						
Eastern Denmark	5.32	1.25-22.6	0.02*	3.14	0.67-14.61	0.15
Age (<60 years ref)						
>60 years	2.21	0.88-5.53	0.09	-		
Gender (female ref)						
Male	1.30	0.54-3.12	0.56	-		
HPV-status (HPV+ ref)						
HPV-	4.10	1.63-10.33	0.002*	2.08	0.66-6.60	0.21
Smoking status (never ref)						
Previous	1.08	0.65-14.51	0.15	-		
Current	5.51	1.26-24.17	0.02*			
Tumor location (tonsils ref)						
BOT	1.50	0.44-5.13	0.52	-		
Other	3.27	1.31-8.13	0.01*			
UICC stage (I ref)						
UICC II	3.19	1.45-7.0	0.004*	1.73	0.68-4.39	0.25

Abbreviations: UTMDACC, The University of Texas MD Anderson Cancer Center, USA; ref, reference; HPV, Human Papillomavirus; BOT, base of tongue. *Significant p-value. ** Adjusted for HPV-status, center and stage UICC8.

Table 6. Characteristics of 560 patients with OPSCC stage III-IV in Eastern Denmark and UTMDACC, Texas, USA from 2015-2020.

Variable	Eastern Denmark, n=353		UTMDACC, Texas USA, n=207		p-value*
	no.	%	no.	%	
Gender					<i><0.001*</i>
Male	260	73.7	190	91.8	
Female	93	26.3	17	8.2	
Median age (IQR)	64 (58-70)		63 (57-68)		<i>0.02*</i>
no.	353		207		
Median follow-up. years (IQR)	1.9 (1.2-2.8)		2.1 (1.1-3.6)		<i>0.02*</i>
no.	192		207		
Smoking					<i><0.001*</i>
Current	221	62.6	19	9.2	
Former	104	29.5	106	51.2	
Never	28	7.9	81	39.1	
Unkown	0		1	0.5	
HPV-status					<i><0.001*</i>
HPV+	82	23.2	139	67.1	
HPV-	270	76.5	68	32.9	
Unknown	1	0.3	0	-	
Tumor location					<i><0.001*</i>
BOT	137	38.8	127	61.4	
Tonsils	114	32.3	66	31.9	
Other	102	28.9	14	6.8	
TNM Stage (UICC/AJCC 8), HPV+					
T-class					<i>0.86</i>
T1	8	9.8	10	7.2	
T2	9	11.0	13	9.4	
T3	30	36.6	56	40.3	
T4	35	42.7	60	43.2	
N-class					<i>0.002*</i>
N0	0	-	1	0.7	
N1	0	-	4	2.9	
N2	53	64.6	112	80.6	
N3	29	35.4	22	15.8	
M-class					<i>0.003*</i>
M0	79	96.3	125	89.9	
M1	1	1.2	14	10.1	

Unkown	2	2.4	0	-	
Overall stage (UICC/AJCC8), HPV+					0.007*
III	81	98.8	124	89.2	
IV	1	1.2	15	10.8	
TNM Stage (UICC/AJCC 8), HPV-					
T-class					0.57
T1	45	16.7	11	16.2	
T2	63	23.3	22	32.4	
T3	83	30.7	19	27.9	
T4	79	29.3	16	5.9	
N-class					0.44
N0	37	13.7	6	8.8	
N1	54	20	19	27.9	
N2	155	57.4	37	54.4	
N3	24	8.9	6	8.8	
M-class					0.23
M0	264	97.8	66	97.1	
M1	6	2.2	1	1.5	
Unknown	0	-	1	1.5	
Overall stage (UICC/AJCC8), HPV-					1
III	72	26.7	18	26.5	
IV	198	73.3	50	73.5	
Treatment					<0.001*
CRT	201	56.9	103	49.8	
RT	132	37.4	15	7.2	
Neoadjuvant chemotherapy + RT/CRT			81	39.1	
Surgery	12	3.4	1	0.5	
Surgery RT/CRT	8	2.3	7	3.4	

Note: Frequency (%) is provided for categorical variables, median (IQR) are provided for continuous variables. Chi Square test were used for categorical variables, while t-test was used for continuous variables. Fisher's exact test was used for small sample sizes.

Abbreviations: UTMDACC, The University of Texas MD Anderson Cancer Center, USA; IQR, interquartile range; HPV, Human Papillomavirus; BOT, base of tongue; RT, radiotherapy; CRT, concurrent RT + systemic therapy.

*Significant p value

Table 7. Multi- and univariable analysis for overall survival for OPSCC UICC8 Stage III/IV in Eastern Denmark and UTMDACC from 2015-2020.

Variable	Univariable			Multivariable**		
	HR	95% CI	p-value*	HR	95% CI	p-value*
Center (UTMDACC ref)						
Eastern Denmark	2.51	1.76-3.57	<0.001*	2.20	1.29-3.75	0.004*
Age (<60 years ref)						
<60 years	1.07	0.79-1.43	0.68	-		
Gender (female ref)						
Male	0.93	0.66-1.31	0.67	-		
HPV-status (HPV+ ref)						
HPV-	1.91	1.39-2.63	<0.001*	0.76	0.46-1.26	0.29
Smoking status (never ref)						
Previous	1.88	1.14-3.12	0.01*	1.52	0.89-2.58	0.23
Current	3.00	1.85-4.85	<0.001*	1.85	1.03-3.30	0.04*
Tumor location (tonsils ref)						
BOT	0.74	0.53-1.03	0.07	-		
Other	1.06	0.74-1.52	0.74			
UICC stage (III ref)						
UICC IV	1.85	1.39-2.48	<0.001*	1.80	1.24-2.62	0.002*
Treatment (CRT ref)						
RT	2.34	1.72-3.19	<0.001*	2.20	1.59-3.04	<0.001*
Neoadjuvant chemotherapy + RT/CRT	0.84	0.52-1.35	0.47	1.84	0.96-3.51	0.07
Surgery	0.83	0.30-2.32	0.72	0.75	0.26-2.14	0.59
Surgery + RT/CRT	0.86	0.35-2.11	0.74	1.07	0.42-2.72	0.88

Abbreviations: UTMDACC, The University of Texas MD Anderson Cancer Center, USA; ref, reference; HPV, Human Papillomavirus; BOT, base of tongue; RT, radiotherapy; CRT, concurrent RT + systemic therapy.
*significant p-value. **Adjusted for center, HPV-status, smoking status and treatment regimen.

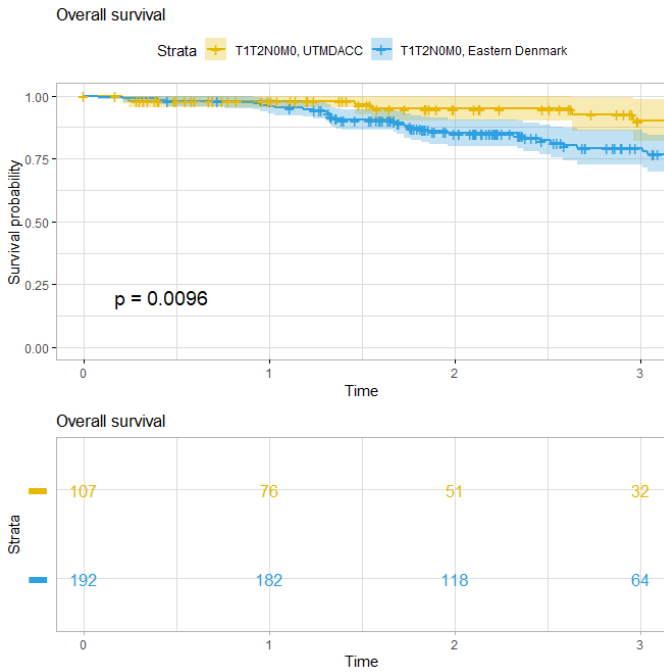
Table 8. Multi- and univariable analysis for recurrence-free interval for OPSCC UICC8 Stage III/IV in Eastern Denmark and UTMDACC from 2015-2020.

Variable	Univariable			Multivariable**		
	HR	95% CI	p-value*	HR	95% CI	p-value*
Center (UTMDACC ref)						
Eastern Denmark	1.95	1.30-2.93	0.001*	2.80	1.46-5.38	0.002*
Age (<60 years ref)						
>60 years	0.97	0.67-1.38	0.85	-		
Gender (female ref)						
Male	0.95	0.62-1.46	0.81	-		
HPV-status (HPV+ ref)						
HPV-	1.52	1.04-2.22	0.03*	0.74	0.41-1.33	0.32
Smoking status (never ref)						
Previous	1.11	0.67-1.88	0.67	-		
Current	1.48	0.90-2.44	0.12	-		
Tumor location (tonsils ref)						
BOT	0.92	0.61-1.40	0.70	-		
Other	1.46	0.94-2.270	0.09	-		
UICC stage (III ref)						
UICC IV	1.92	1.34-2.74	<0.001*	2.20	1.35-3.57	0.001*
Treatment (CRT ref)						
RT	1.90	1.28-2.81	0.001*	1.84	1.22-2.76	0.004*
Neoadjuvant chemotherapy + RT/CRT	1.29	0.77-2.15	0.33	3.10	1.45-6.65	0.004*
Surgery	0.66	0.16-2.71	0.57	0.72	0.17-2.98	0.65
Surgery + RT/CRT	0.81	0.25-2.60	0.73	1.13	0.34-3.70	0.85

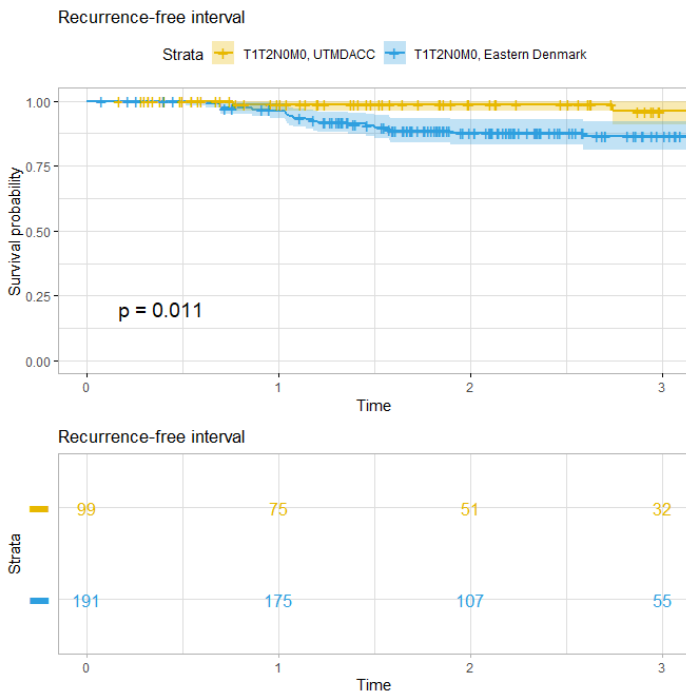
Abbreviations: UTMDACC, The University of Texas MD Anderson Cancer Center, USA; ref, reference; HPV, Human Papillomavirus; BOT, base of tongue; RT, radiotherapy; CRT, concurrent RT + systemic therapy.
*Significant p-value. ** Adjusted for center, HPV-status, stage UICC8, treatment regimen.

Figure 1. Kaplan-Meier curves depicting the overall survival and recurrence-free interval stratified by center for T1/T2N0M0 OPSCC from 2015-2020. A. Overall survival. B. Recurrence-free interval.

A.



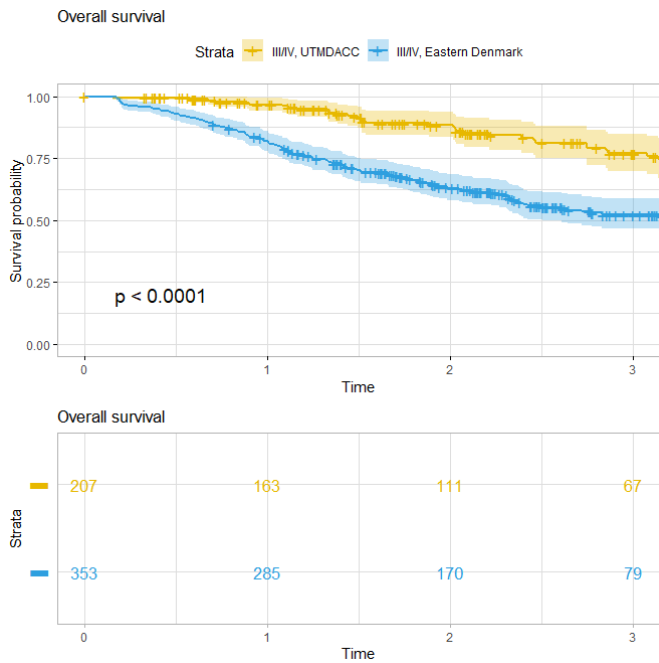
B.



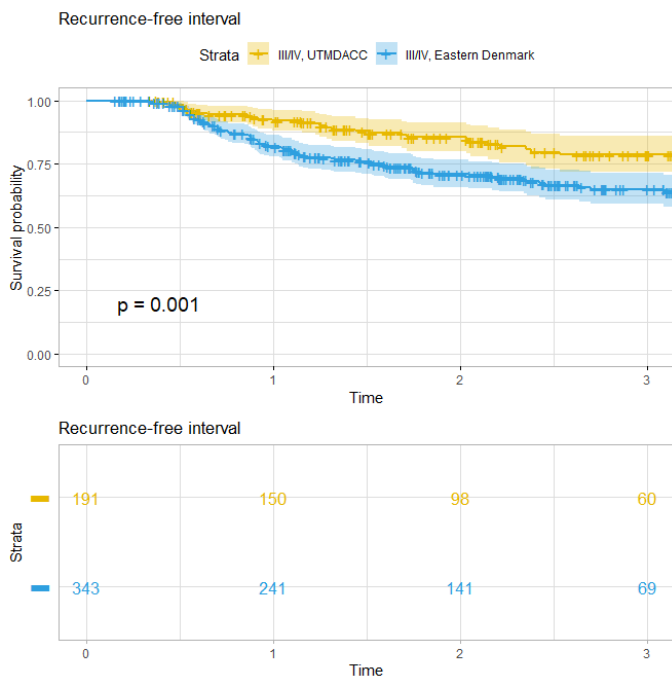
Abbreviations: UTMDACC, The University of Texas MD Anderson Cancer Center, USA, HPV, Human Papillomavirus; OPSCC, oropharyngeal squamous cell carcinoma.

Figure 2. Kaplan-Meier curves depicting the overall and recurrence-free interval stratified by center for OPSCC UICC8 stage III-IV from 2015-2020. A. Overall survival. B. Recurrence-free interval.

A.



B.



Abbreviations: UTMDACC, The University of Texas MD Anderson Cancer Center, USA, HPV, Human Papillomavirus; OPSCC, oropharyngeal squamous cell carcinoma.



Mesenchymal Stromal/Stem Cell Therapy Improves Salivary Flow Rate in Radiation-Induced Salivary Gland Hypofunction in Preclinical in vivo Models: A Systematic Review and Meta-Analysis

Amanda-Louise Fenger Carlander^{1,8} · Anders Kierkegaard Gundestrup¹ · Per Marcus Jansson¹ · Bjarke Follin² · Cecilie Hoeeg² · Birgitte Saima Kousholt³ · Rasmus Tolstrup Larsen^{4,5} · Kathrine Kronberg Jakobsen¹ · Susie Rimborg⁶ · Anne Fischer-Nielsen⁷ · Christian Grønhoj¹ · Christian von Buchwald¹ · Charlotte Duch Lynggaard¹

Accepted: 17 February 2024 / Published online: 2 March 2024
© The Author(s) 2024

Abstract

Background Mesenchymal stromal/stem cells (MSCs) have been suggested for salivary gland (SG) restoration following radio-induced salivary gland damage. This study aimed to determine the safety and effectiveness of MSC therapy on radio-induced SG damage and hypofunction in preclinical in vivo studies.

Methods PubMed and EMBASE were systematically searched for preclinical in vivo interventional studies evaluating efficacy and safety of MSC treatment following radio-induced salivary gland damage published before 10th of January 2022. The primary endpoint was salivary flow rate (SFR) evaluated in a meta-analysis. The study protocol was published and registered on PROSPERO (www.crd.ac.uk/prospero), registration number CRD42021227336.

Results A total of 16 preclinical in vivo studies were included for qualitative analysis (858 experimental animals) and 13 in the meta-analysis (404 experimental animals). MSCs originated from bone marrow (four studies), adipose tissue (10 studies) and salivary gland tissue (two studies) and were administered intravenously (three studies), intra-glandularly (11 studies) or subcutaneously (one study). No serious adverse events were reported. The overall effect on SFR was significantly increased with a standardized mean difference (SMD) of 6.99 (95% CI: 2.55–11.42). Studies reported improvements in acinar tissue, vascular areas and paracrine factors.

Conclusion In conclusion, this systematic review and meta-analysis showed a significant effect of MSC therapy for restoring SG functioning and regenerating SG tissue following radiotherapy in preclinical in vivo studies without serious adverse events. MSC therapy holds significant therapeutic potential in the treatment of radio-induced xerostomia, but comprehensive, randomized, clinical trials in humans are required to ascertain their efficacy in a clinical setting.

Keywords Xerostomia · Radiotherapy · Cell Therapy · Mesenchymal stem Cells · Systematic Review

✉ Amanda-Louise Fenger Carlander
amanda-louise.fenger.carlander@regionh.dk

¹ Department of Otolaryngology and Audiology, Head and Neck Surgery, Rigshospitalet, Copenhagen University Hospital, Copenhagen, Denmark

² Cardiology Stem Cell Centre, Rigshospitalet, Copenhagen University Hospital, Copenhagen, Denmark

³ Department of Clinical Medicine, Aarhus University Group for Understanding Systematic Reviews and Meta analyses in Translational Preclinical Science, Aarhus University, Copenhagen, Denmark

⁴ Department of Occupational Therapy and Physiotherapy, Rigshospitalet, Copenhagen University Hospital, Copenhagen, Denmark

⁵ Section of Social Medicine, Department of Public Health, University of Copenhagen, Copenhagen, Denmark

⁶ The Royal Danish Library, Copenhagen University Library, Copenhagen, Denmark

⁷ Department of Immunology, Rigshospitalet, Copenhagen University Hospital, Copenhagen, Denmark

⁸ Department of Otolaryngology, Head and Neck Surgery and Audiology, Rigshospitalet, Copenhagen University hospital, Copenhagen, Denmark

Introduction

Most patients with head and neck cancer (HNC) are treated with radiotherapy [1]. Salivary gland (SG) hypofunction and xerostomia, the subjective feeling of dry mouth, are common and long-term side effects following radiotherapy in the head and neck area [2]. Despite the emergence of intensity-modulated radiation-therapy (IMRT), that to some extent spare the surrounding tissue due to a more precise delivery to target tissue, the SG are often damaged [3, 4]. Radio-induced SG damage is dose-dependent and leads to gland degeneration and progressive decline in saliva production, followed by complications such as xerostomia, problems with speech and swallowing, oral infections and dental caries thus reducing quality of life. Currently, only symptomatic treatments are available, and there is a lack of regenerative and restorative therapeutic options [2, 5–7].

Mesenchymal stromal/stem cells (MSCs) are multipotent adult progenitor cells that in vitro can differentiate into mesodermal lineages with abilities for tissue regeneration and which can be isolated from numerous connective tissues, e.g. bone marrow (MSCs[M]) and adipose tissue (MSCs[AT]) [8–10]. Aside from being easily accessible, MSCs encompass various advantages such as proliferative and differentiating capacities; but also, immunomodulatory, and trophic properties such as anti-inflammatory, anti-fibrotic, anti-apoptotic, angiogenic and immunosuppressive effects [11–13]. Thus, MSCs have a therapeutic and disease-modifying potential to repair and/or restore radio-induced SG damage. Recent preclinical in vivo studies have focused on mesenchymal stem cell (MSC) transplantation to repair radiation damaged SGs as a potentially curative treatment for SG hypofunction [14, 15]. Also, MSC therapy have shown to improve salivary flow rate (SFR) in humans [16–19].

Nevertheless, while MSC therapy shows potential as a treatment option for radio-induced SG damage, existing studies have been limited in size, characterized by high heterogeneity in relation to MSC origin, and only a few have been conducted in both preclinical in vivo models and humans. The use of MSCs therapy for radio-induced SG hypofunction alone has not yet been evaluated in a systematic review and meta-analysis. The aim of this study was therefore to review the safety and effectiveness of MSC therapy for restoring SG function after radiation-induced damage in preclinical in vivo studies. This is of great importance to optimize clinical trials and to assess the prospective implication in the curative treatment of SG hypofunction caused by radiotherapy.

Method

The study adheres to the Preferred Reporting Items for Systematic Reviews and Meta-Analyses Protocols (PRISMA-P) statement and the study protocol was published and registered on PROSPERO (www.crd.ac.uk/prospéro), registration number CRD42021227336 [20].

We considered preclinical in vivo models that assessed MSC therapy following experimentally induced radiation injury of major SGs. Inclusion criteria were: (1) preclinical in vivo intervention studies of both sexes and all ages (2) exposure of SGs to ionizing radiation (3) MSC therapy of all administration routes. There were no restrictions regarding induction of radiation damage, but MSCs administration should be after induction of radiation injury. MSC secretome, exosomes, and treatment with parts of MSCs were also included. In vitro models, treatments other than MSCs, SG damage other than ionizing radiation and non-relevant outcome were excluded. Human studies were not included in the formal analysis but described and included the discussion. The primary outcome was efficacy measured by SFR and secondary outcomes was SG morphology, SG histology, changes in saliva composition, circulating immune cells, SG paracrine effects, mode of action and safety in terms of objective adverse events as previously described [20].

Systematic Search

In January 2022 two authors (ALFC, CH) systematically searched PubMed and Embase for preclinical in vivo interventional studies assessing the efficacy and safety of MSC therapy for radiation-induced SG hypofunction. The search was performed using Medical Subject Headings (MeSH), Emtree and text words relating to MSCs, SG hypofunction, SG damage, SG dysfunction, radiation-induced SG damage or xerostomia. The specific search string for PubMed and Embase was previously described [20]. The search of databases reference lists was evaluated for additional relevant studies.

Data Extraction

Two authors (ALFC, CH) independently screened all articles for eligibility and disagreement was solved by consensus or by discussion with a third reviewer (BM or CDL). The following information was extracted from each study: (1) article information (author, publication year), (2) details on preclinical in vivo model (species, sex, sample size, age), (3) study design (controlled, uncontrolled, randomized and/or blinded), (4) irradiation details (dose, Gy, quality assurance and days from irradiation to MSC therapy), (5) MSC therapy (type, concentration, administration route,

follow-up time), 7) statistical analysis, 8) outcomes (functional and molecular outcomes).

Quality Assessment and Risk of Bias

We assessed the quality of reporting in the included studies according to the latest Animal Research: reporting of in vivo Experiments (ARRIVE) guidelines [21]. One point was given for evidence of each quality criterion. The methodological quality was assessed using the SYRCLE (Systematic review Center for Laboratory Animal Experimentation) risk of bias tool in domains related to selection bias, performance bias, detection bias, attrition bias and reporting bias [22].

Data Analysis

A descriptive summary of all outcomes was performed. The efficacy of MSC therapy was evaluated by a random effect meta-analysis adjusted to Hedge's g on SFR. If there were multiple time points, only the last one was included in the meta-analysis. A standardized mean difference (SMD) with 95% confidence intervals (CIs) was used to evaluate the effect on SFR. The SMD was calculated by dividing the difference in mean outcome between the groups with the standard deviation of outcome among participants as per recommendation by the Cochrane Handbook [23]. Heterogeneity of the study results was investigated using the Cochrane Q test and quantified with I^2 values. Subgroup analyses were performed for species, strain, sex, administration route, age in weeks, radiation duration, frequency of treatment, radiation dose and time between radiation to MSC treatment. Sub analysis on frequency of radiation was not performed since radiation was administered as a single treatment in all studies.

Results

Sixteen preclinical studies published between 2011 and 2019 met the inclusion criteria, see Fig. 1 [24–39]. One human study [19] ($n=30$, intervention group $n=15$) were identified. All preclinical in vivo studies were included in the quantitative (858 experimental animals) and 13 (404 experimental animals) were also included in the qualitative analyses, see Fig. 1.

Description of the Preclinical in vivo Models

All studies investigated the safety and efficacy of MSC-based therapy for xerostomia and SG hypofunction following radiation in the head and neck area. A total of 858

animals were included of which 341 received intervention with MSC therapy. Ten studies included mice, two studies rats and one study miniature pigs. The irradiation dose varied from 10 to 25 Gy, and all were administered as a single dose. See Table 1.

MSCs were originating from bone marrow (4 studies) [24, 27, 36, 37], adipose tissue (10 studies) [25, 28–33, 35, 38, 39], and salivary gland tissue (2 studies) [26, 34] and were administered intravenously (3 studies) [28, 31, 39], intra-glandularly (11 studies) [24–27, 29, 32–36, 38] or subcutaneously (1 study) [30]. One study did not specify administration route [37]. One study used secretomes originating from adipose tissue [39]. The mean follow-up time was 12 weeks after MSC treatment, ranging from 7 days to 24 weeks. See Table 1.

Safety of MSC

All included preclinical in vivo studies described the MSC-based cell therapy as safe with no reported serious adverse events [24–39].

The Effect of MSCs on Efficacy, Salivary Flow Rate

13 studies included SFR as primary functional outcome [24–29, 31–35, 38, 39]. Eleven studies found a significant increase in SFR after intervention with MSCs compared to other groups (placebo/sham or IR only) [24–29, 32–35, 38], with an overall effect of SMD 6.99 (95% CI: 2.55–11.42), ranging from 1.68 (95% CI: 0.63–2.74) [31] to 25.41 (95% CI: 15.22–35.59) [33]. Heterogeneity was $X^2=177.37$ ($p<.001$) and $I^2=93%$ (95% CI [90–95%]). See Fig. 2.

Subgroup analyses investigating the effect of species, strain, sex, administration route, age in weeks, radiation duration, frequency of treatment, radiation dose and time between radiation to MSC treatment revealed significant differences regarding strain ($p<.001$) and administration route ($p=.01$). The most prominent differences were observed in the strains NOD.SCID-PrckSCID (SMD 22.92, 95% CI: 18.99–26.86) and Wistar (SMD 25.41, 95% CI: 15.22–35.59) compared to C57BL/6 (SMD 2.90, 95% CI: 0.08–5.52), CH3 (SMD 4.99, 95% CI: 1.58–8.39) and Sprague-Dawley (SMD 2.49, 95% CI: 1.81–3.18), while the effect following intraglandular injection was greater than following intravenously injection (SMD 8.59, 95% CI: 2.94–14.24 versus 2.12, 95% CI: 1.20–3.05, respectively). There was no significant effect of species, sex, age in weeks, radiation dose, frequency of treatment or time from radiation to first treatment. Effect of duration of radiation was not possible to assess due to insufficient data. See **Supplementary Results**.

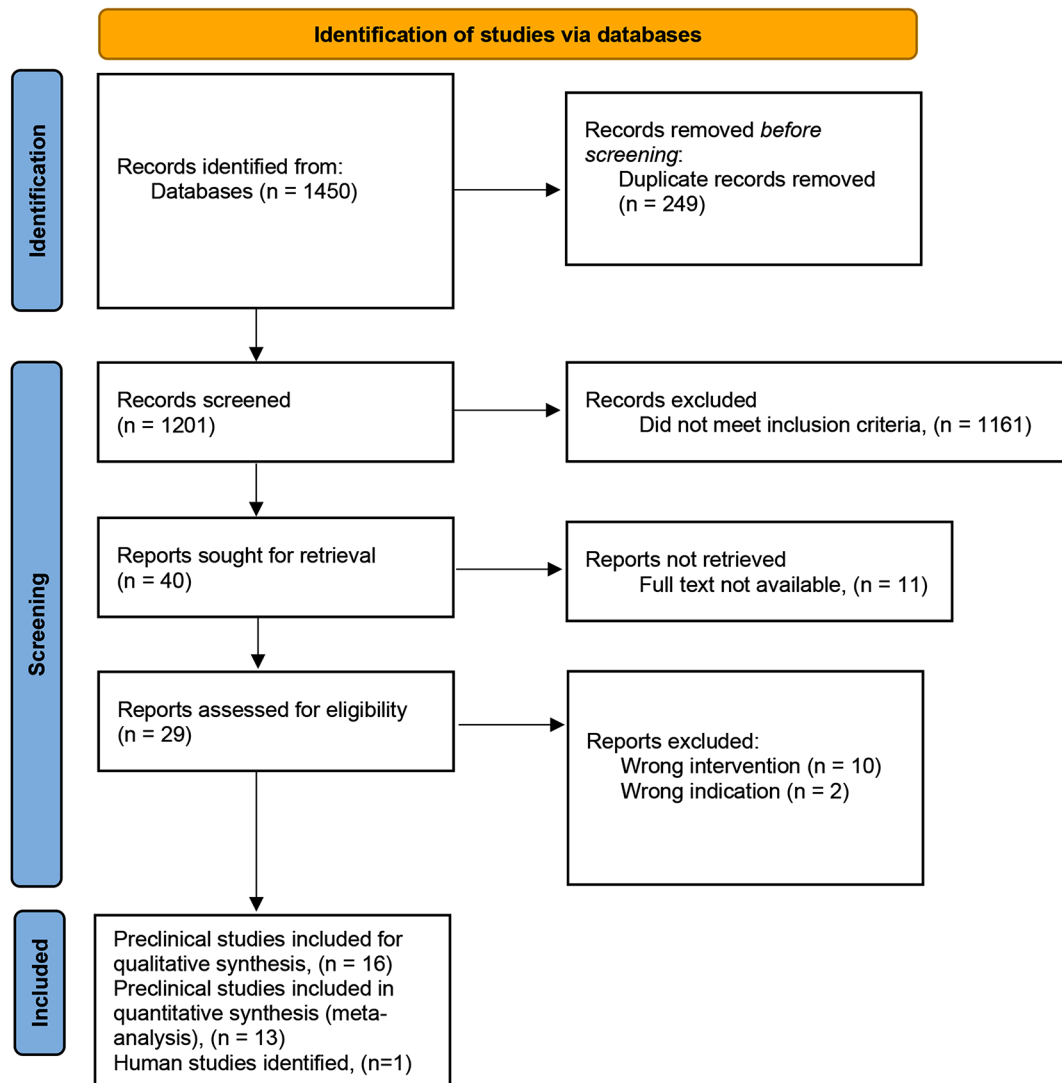


Fig. 1 PRISMA 2020 Statement flow chart of the screening process for the selection of eligible studies

The Effect of MSCs on Salivary Gland Regeneration and Apoptosis

Seven studies reported improvements in acinar tissue [25–27, 29, 31, 32, 35] with more acinar cells [25, 29, 32, 35] and more compact acinar structure [26]. Also, several studies reported less inflammation and fibrosis [25, 28, 30, 33, 35, 36] and increased amylase [27, 28, 31–34, 37–39]. Li et al. and Wang et al. reported intact cellular ultra-microstructure with healthy cell membrane and almost undamaged cytoplasmic organelles [31, 38]. They also found significantly higher proliferative activity, but this was not found by others [27, 28]. Shin et al. found greater expression of SG epithelial cell markers (KRT7 and KRT18) and upregulated structure-related genes (SMR3A, AMY2A5, PRB1, AMY1, CLDN22, PRPMP, AMY1A and AQP5) [35]. Similarly, Choi et al. found higher expressions of AQP5, alfa-SMA

and CD31 [33]. Mulyani et al. found increased expression of SDF1-CXCR4 and Bcl-2 genes [37]. Nine studies reported reduction in apoptotic cells [26–33, 38], though Wang et al. did not specify the results [32].

Lim et al. observed anti-alfa-amylase signaling in transplanted MSC(M) suggesting a transdifferentiation into SG epithelial cells [27], but this was not found by Kojima et al. [25].

The Effect of MSCs on Vascular Areas

Several studies reported an increase in vascular areas [25, 27, 29–31]. Furthermore, Kojima et al. localized MSCs in vessel endothelial cells post transplantation five- and ten-weeks post transplantation [25]. Wang et al. also found

Table 1 Study characteristics

Author (Year)	Animal: species, strain and age, gender	Study design	Groups	Irradiation	Days from radiation to MSC treatment	MSC type and concentration	Administration route	Statistical analysis	Functional outcome	Molecular outcome
Lin, C et al. (2011)	Mice, NOD.SCID- <i>Prck^{scid}</i> , four weeks old	Pro-spective, controlled trial	1. Control group, no IR, no treatment, <i>n</i> = 35. 2. IR, no treatment, <i>n</i> = 35. 3. IR, treated with MSC(M), <i>n</i> = 35. 4. IR, treated with acinar-like cells, <i>n</i> = 35	Single dose, 15 Gy, from ⁶⁰ Co-source. No QA on radiation delivery was performed.	11 days	10 ⁶ MSC(M) and acinar-like cells, co-cultured from MSC(M).	Intracanalicular injection (submandibular gland)	One-way ANOVA and marginal linear regression analysis. Significance level was <i>P</i> < .05.	Saliva production in group 3 and 4 significantly increased. Body weight and gland weight significantly higher in groups 3 and 4. Significantly faster rate of recovery in gland weight in group 4, compared to group 3.	No molecular data. However, a significantly higher expression of α -amylase in acinar cells and co-cultured MSC(M) compared to MSC(M) which was not been co-cultured
Kojima, T. et al. (2011)	Mice, C57BL/6, nine weeks old	Pro-spective, controlled trial	1. Control group, no IR, no treatment, <i>n</i> = 24. 2. IR, treated with ADSCs in PBS, <i>n</i> = 31. 3. IR, treated with PBS only, <i>n</i> = 31.	Single dose, 10 Gy gamma irradiation from ¹³⁷ C source. No QA on radiation delivery was performed.	70 days (10 weeks)	ADSCs 0.1 mL solution of PBS containing 500,000 cells.	Intracanalicular injection (submandibular glands)	One-way and two-way ANOVA followed by Fishers PLSD or students <i>t</i> -test. Significance level was <i>P</i> < .01 or <i>P</i> < .05.	Significant increase in SFR in ADSC group compared to sham. 75% function compared to normal group.	Group 2 had more acinar cells and less inflammatory infiltration compared to group 3. Group 2 had significantly more CD31-positivity, and showed a significant increase in levels of HGF, VEGF and several proteins related to angiogenesis.
Jeong, J. et al. (2013)	Rats, Wistar rats, six weeks old, male	Pro-spective, controlled trial	1. Control group, no IR, no treatment. 2. IR, treated with PBS 3. IR, treated with hSGSCs	Single dose, 25 Gy, dose rate of 2 Gy min ⁻¹ . No QA on radiation delivery was performed.	1 day	5 × 10 ⁵ hSGSCs	Intracanalicular injection	One-way ANOVA. Significance level of < 0.05 was used.	At 60 days, significant increase in SFR and body weight in group 3 compared to group 4.	Group 3 had compact acinar and ductal structure like undamaged rat tissue, whereas group 2 had disrupted acinar structure and numerous vacuoles. Apoptotic cells were observed only in group 2 and not in group 3.
Lim, J. et al. (2013)	Mice, C57BL/6, eight to nine weeks old	Pro-spective, randomized, controlled trial	1. Control group, no IR, no treatment, <i>n</i> = 8. 2. IR, treated with PBS, <i>n</i> = 8. 3. IR, treated with BM-cMSCs, <i>n</i> = 8.	Single dose, 15 Gy. No QA on radiation delivery was performed.	1 day	1 × 10 ⁵ cMSC(M) in 15 μ L of PBS.	Intracanalicular injection (submandibular glands)	Mann-Whitney test for differences between the two groups. Kruskal-Wallis followed by post hoc Dunns for three groups. Significance level was <i>P</i> < .05.	Significant increase in SFR in group 3 compared to group 2. Body weight, gland weight and salivary lag time showed no significant differences.	Significant reduction of apoptosis and increase in microvessel density in group 3 compared to group 2. No significant difference in proliferation activity. Significantly higher mucopolysaccharide and tissue amylase upon histological examination.

Table 1 (continued)

Author (Year)	Animal: species, strain and age, gender	Study design	Groups	Irradiation	Days from radiation to MSC treatment	MSC type and concentration	Administration route	Statistical analysis	Functional outcome	Molecular outcome
Lim, J. et al. (2013)	Mice, CH3, eight to nine weeks old	Prospective, randomized, controlled trial.	1. Control group, no IR, $n=20$. 2. IR, no treatment, $n=20$. 3. IR, treated with hAdMSCs, $n=20$.	Single dose, 15 Gy No QA on radiation delivery was performed.	0 days (once weekly for 3 consecutive weeks)	1×10^6 MSC(AT)h.	Intravenously (tail vein)	Mann-Whitney test for differences between two groups. Kruskal-Wallis followed by post hoc Dunns for three groups. Significance level was $P < .05$.	At 12 weeks significant increase in SFR in group 3 compared to group 2. Salivary lag time was significantly longer in group 2 compared to group 1.	A significantly higher fraction of mucin and amylose and reduction in apoptosis in group 3 compared to group 2. No significant differences in proliferative activity.
Xiong, X. et al. (2014)	Rats, Spraque-Dawley, 12 weeks old	Prospective, randomized, controlled trial.	1. Control group, no IR, no treatment, $n=30$. 2. IR, treated with PBS, $n=30$. 3. IR, treated with hADSCs, $n=30$.	Single dose, 18 Gy, dose rate of 300 cGy min^{-1} No QA on radiation delivery was performed.	0 days	1×10^6 MSC(AT)h in 0.1 mL PBS solution.	Intralandular injection (submandibular glands)	One-way ANOVA and Student-Newman-Keuls analysis. Significance level was $P < .05$.	At 24 weeks, significant increase in SFR in group 3 compared to group 2. Group 3 recovered to 71% SFR compared to group 1.	Group 3 had a significantly larger number of acinar cells and PAS-positive acini, significantly higher area of blood vessels and significantly fewer apoptotic cells compared to group 2. Significant increase in mRNA levels of VEGF, HGF, COX-2 and MMP-2 in group 3 compared to group 2.
Chen, Y. and Niu, Z. et al. (2014)	Minipigs, eight months old, female	Prospective, randomized, controlled trial.	1. IR, treated with ADSCs + PRF, $n=5$. 2. IR, treated with ADSCs, $n=5$. 3. IR, treated with PRF, $n=5$. 4. IR, treated with PBS, $n=5$.	Single dose, 20 Gy, dose rate of 3 Gy min^{-1} . Only right parotid gland was radiated. No QA on radiation delivery was performed.	28 days (four weeks)	4×10^6 cells ADSCs (autologous),	40 subcutaneous injections (0.2 mL/point), with 2 cm interval between neighbouring points.	One-way ANOVA and Kruskal-Wallis H test. Significance level was $P < .05$.	Group 1 showed significantly more fat cells, less fibrosis and inflammation compared to group 4. Group 2 and 3 were significantly better compared to group 4. Group 1–3 showed significantly less apoptotic activity compared to group 4 and group 1 showed the least. Group 1–3 showed significantly more neovascular capillary area, compared to group 4 and group 1 showed the most.	Group 1 showed significantly more fat cells, less fibrosis and inflammation compared to group 4. Group 2 and 3 were significantly better compared to group 4. Group 1–3 showed significantly less apoptotic activity compared to group 4 and group 1 showed the least. Group 1–3 showed significantly more neovascular capillary area, compared to group 4 and group 1 showed the most.

Table 1 (continued)

Author (Year)	Animal: species, strain and age, gender	Study design	Groups	Irradiation	Days from radiation to MSC treatment	MSC type and concentration	Administration route	Statistical analysis	Functional outcome	Molecular outcome
An, H. et al. (2015)	Mice, CH3, eight to nine weeks old	Prospective, randomized, controlled trial	1. Control group, no IR, no treatment, $n = 35$. 2. IR, treated with PBS, $n = 35$. 3. IR, treated with hADMSC SEC, $n = 35$	Single dose, 15 Gy. No QA on radiation delivery was performed.	0 days (once daily for seven consecutive days)	500 μ L MSC(AT)h. Secretome stemmed from 1×10^5 cell.	Intravenously (tail vein)	Mann-Whitney test, one-way ANOVA followed by Tukey's post hoc test, two-way ANOVA followed by Bonferroni post hoc test and linear regression. Significance level was $P < .05$.	At 16 weeks, significant increase in SFR in group 3 compared to group 1, while group 3 was not. EGF content in saliva was significantly higher in group 3 compared to group 2. Salivary lag time was significantly longer in group 3 compared to the other groups. Body and gland weight was significantly higher in group 3 compared to group 2.	Saliva amylase activity was significantly lower in group 2 compared to group 1, while group 3 was not. EGF content in saliva was significantly higher in group 3 compared to group 2. PAS stain for mucin showed significantly larger area of mucin in group 3 compared to group 2.
Li, Z. et al. (2015)	Mice, C57BL/6, eight weeks old	Prospective, randomized, controlled trial	1. IR, treated with ADSCs, $n = 10$ 2. IR, treated with PBS, $n = 10$ 3. Control, no IR, no treatment, $n = 10$	Single dose, 18 Gy No QA on radiation delivery was performed.	0 days (twice a week for six weeks)	1×10^6 cells ADSCs.	Intravenously (tail vein)	ANOVA followed by Tukey's honestly significant difference test. Significance level was $P < .05$.	At eight weeks significant increase in SFR in group 1 compared to group 2. Gland weight was significantly higher in group 1 compared to group 2.	PAS staining of glands showed significant increase in production of mucopolysaccharide, C31 staining showed significantly higher density of microvessels, and significantly higher levels of amylase in group 1 compared to group 2. Significantly higher proliferation activity and reduction of apoptosis activity in group 1 compared to group 2.
Wang, Z. et al. (2016)	Mice, C3H, 8–12 weeks old, female	Prospective, randomized, controlled trial	1. Control group, no IR, no treatment, $n = 10$ 2. IR, treated with PBS, $n = 10$ 3. IR, treated with ADSCs, $n = 10$ 4. IR, treated with PR, $n = 10$ 5. IR, treated with ADSCs + PRF, $n = 10$	Single dose, 18 Gy No QA on radiation delivery was performed.	84 days (12 weeks)	2×10^5 ADSCs in 100 μ L solution of PBS/PRF.	Intralandular injection (submandibular glands). Weekly injections for 3 consecutive weeks.	Mann-Whitney test and one-way ANOVA followed by Tukey's post hoc test. Significance level was $P < .05$.	At 12 weeks. Significant increase in SFR, body weight and gland weight in group 3, 4 and 5 compared to group 2.	Group 5 showed significantly more acinar cells, more amylase area compared to groups 2,3 and 4. Group 5 showed significantly more microvessels, compared to group 2 and 4, but not group 3. Group 5 showed less apoptotic activity and more proliferation activity compared to group 2.

Table 1 (continued)

Author (Year)	Animal: species, strain and age, gender	Study design	Groups	Irradiation	Days from radiation to MSC treatment	MSC type and concentration	Administration route	Statistical analysis	Functional outcome	Molecular outcome
Wang, Z. et al. (2017)	Miniature pigs, eight months old, female	Prospective, randomized, controlled trial	1. Control group, no IR, $n=5$ 2. IR, treated with ADSCs, $n=5$ 3. IR, treated with PBS, $n=5$	Single dose, 20 Gy, dose rate was 3.2 Gy min ⁻¹ . Only right parotid gland was irradiated. No QA on radiation delivery was performed.	0 days (un-clear?)	4×10^6 ADSCs	Intralandular injection at 40 different injection points, 0.2 mL per injection. Repeated twice per week for 6 weeks.	ANOVA followed by Tukey's honestly significant difference test. Significance level was $P < .05$.	At 3 months significant increase in SFR and gland weight in group 2 compared to group 3. There was no significant difference in gland weight between group 1 and 2.	Amylase levels was significantly higher in group 2 compared to group 3. The number of PAS-positive acinar cells was significantly higher in group 2 compared to group 3. Group 2 showed less fibrosis and more vascularization than group 3.
Choi, J. et al. (2018)	Mice, C3H, eight to nine weeks old.	Prospective, randomized, controlled trial	1. Control group, no IR, $n=28$ 2. IR, treated with PBS, $n=28$ 3. IR, treated with porcine SIS, $n=28$ 4. IR, treated with AdMSC, $n=28$ 5. IR, treated with AdMSC + SIS, $n=28$	Single dose, 15 Gy No QA on radiation delivery was performed.	0 days	1×10^5 cells MSC(AT) in 20 μ L solution of respective carrier.	Intralandular injection (submandibular glands)	T-test, one-way ANOVA followed by Tukey's post hoc test. Significance level was $P < .05$.	At 16 weeks, significant increase in SFR in group 5 compared to group 2. Group 4 and 5 showed significantly less fibrosis than group 2 and 3. Only group 5 showed increased mucin production, compared to group 2 and 3. Group 4 and 5 showed significant anti-apoptotic effects and increased ROS scavenging effects compared to group 2.	Group 4 and 5 showed significantly higher levels of EGF and amylase compared to group 2. Group 4 and 5 showed significantly less fibrosis than group 2 and 3. Only group 5 showed increased mucin production, compared to group 2 and 3. Group 4 and 5 showed significant anti-apoptotic effects and increased ROS scavenging effects compared to group 2.
Shin, H. et al. (2018)	Mice, C3H, six weeks old, female	Prospective, randomized, controlled trial	1. Control group, no IR, $n=6$ 2. IR, treated with PBS, $n=6$ 3. IR, treated with SGSC ^{2D} , $n=6$ 4. IR, treated with SGSC ^{3D} , $n=6$	Single dose, 15 Gy No QA on radiation delivery was performed.	28 days (four weeks)	2×10^5 SGSC ^{3D} in 10 μ L solution of PBS.	Intralandular injection (submandibular glands)	Mann-Whitney test, one-way ANOVA followed by Tukey's post hoc test, two-way ANOVA followed by Bonferroni post hoc test, as well as linear regression. Significance level was $P < .05$.	At 16 weeks, significant increase in SFR, body and gland weight and lower lag time in group 3 and 4 compared to group 2. Significant increase in SFR, body and gland weight and lower lag time in group 4 compared to group 3.	Group 3 and 4 showed significantly higher amylase activity and higher levels of EGF compared to group 2. Group 4 showed significantly higher amylase activity and levels of EGF compared to group 3. Group 3 and 4 showed significantly higher expression of genes related to salivary gland function and growth factors compared to group 2. Group 4 showed significantly higher expression compared to group 3, however by a smaller relative margin.

Table 1 (continued)

Author (Year)	Animal: species, strain and age, gender	Study design	Groups	Irradiation	Days from radiation to MSC treatment	MSC type and concentration	Administration route	Statistical analysis	Functional outcome	Molecular outcome
Shin, H. et al. (2018)	Mice, C3H, six weeks old.	Prospective, randomized, controlled trial	1. Control group, no IR, $n=4$ 2. IR, treated with PBS, $n=4$ 3. IR, treated with hADSC ^{NMX} , $n=4$ 4. IR, treated with hADSC ^{HPX} , $n=4$	Single dose, 15 Gy. No QA on radiation delivery was performed.	28 days (four weeks)	2×10^5 MSC(AT) ^h ^{HPX} and MSC(AT) ^h ^{NMX} in 10 μ L solution of PBS.	Intracranial injection (submandibular glands)	Mann-Whitney test, one-way ANOVA followed by Tukey's post hoc test, two-way ANOVA followed by Bonferroni post hoc test. Significance level was $P < .05$.	At 12 weeks significant increase in SFR, amylase activity , body and gland weight in group 3 and 4 compared to group 2, and significantly higher in group 2, and significantly higher in group 3 compared to group 4. Lag time was significantly shorter in group 3 & 4 compared to group 2.	Significantly more fluorescently labelled MSC(AT) ^h ^{HPX} cells compared to MSC(AT) ^h ^{NMX} cells in random fields. The density of PAS-positive acinar cells was significantly higher in group 3 and 4 compared to group 2, and significantly higher in group 4 compared to group 3. Significantly less fibrosis in group 2. EGF levels were higher in group 3 and 4 compared to group 2.

Table 1 (continued)

Author (Year)	Animal: species, strain and age, gender	Study design	Groups	Irradiation	Days from radiation to MSC treatment	MSC type and concentration	Administration route	Statistical analysis	Functional outcome	Molecular outcome
Elsaadany, B. et al. (2019)	Rats, albino, three to four months old, male	Pro-spective, randomized, controlled trial	1. Control group, no IR, no treatment, $n=6$ 2. IR, treated with PBS, $n=6$ 3. IR, treated with MSC(M), $n=6$	Single dose, 1.3 Gy. No QA on radiation delivery was performed.	0 days	1×10^5 MSC(M) in 0.2 mL PBS solution.	Intracranial injection.	ANOVA followed by Tukey's post hoc test. Significance level was $P < .05$.	No functional outcome parameter.	The acini diameter was significantly larger in group 3 compared to group 2. Group 3 showed significantly less fibrosis compared to group 2. Group 2 showed necrosis of the ductal lining epithelium, which was less commonly observed in group 3, however no significantly different data was reported.
Mulyani, S. et al. (2019)	Rats, Wistar, three to four months old, male	Pro-spective, randomized, controlled trial	1. Control group, no IR, no treatment, $n=10$ 2. IR, no treatment, $n=10$ 3. IR, treated with MSC(M) ^{NMX} , $n=10$ 4. IR, treated with MSC(M) ^{HPX} , $n=10$	Single dose, 1.5 Gy. No QA on radiation delivery was performed.	1 day	MSC(M) ^{HPX} and MSC(M) ^{NMX} . Dose not specified.	Not specified	Normality test and MANOVA test. Significance level was $P < .05$.	At 7 days, group 4 showed a significantly higher expression of amylase compared to group 3. Group 3 and 4 showed significantly higher levels compared to group 2.	The expression of SDF1-CXCR4 and Bcl-2 was significantly higher in group 4 compared to group 3.

Abbreviations: ADSC, adipose-derived stromal cells; ANOVA, analysis of variance; Bcl-2, B-cell lymphoma 2; COX-2, cyclooxygenase-2; EGF, epidermal growth factor; HGF, hepatocyte growth factor; hADSC, human adipose-derived stromal cells; hSGCs, human salivary gland stem cells; IR, irradiation; MANOVA, multivariate analysis of variance; MMP-2, matrix-metalloproteinase-2; MSC(AT)h, human adipose-derived mesenchymal stem cells; MSC(AT)h SEC, human adipose-derived mesenchymal cell secretome; MSC(AT)h^{HPX}, human adipose-derived mesenchymal stem cells incubated in a hypoxic environment; MSC(AT)h^{NMX}, human adipose-derived mesenchymal stem cells incubated in a normoxic environment; MSC(M), bone marrow-derived clonal mesenchymal stem cells; MSC(M)^{HPX}, bone marrow-derived mesenchymal stem cells incubated in a hypoxic environment; MSC(M)^{NMX}, bone marrow-derived mesenchymal stem cells incubated in a normoxic environment; PBS, phosphate-buffered saline (PBS) solution; PAS, Periodic acid Schiff; PRF, platelet-rich fibrin; SGSC2D, human parotid gland 2D monolayer cultured human salivary gland stem cells; SGSC3D, 3D spheroid microwell cultured human salivary gland stem cells; SDF1-CXCR4, stromal cell-derived factor 1-C-X-C chemokine receptor type 4; SFR, salivary flow rate; SIS, small intestine submucosa; VEGF, vascular endothelial growth factor; QA, quality assurance

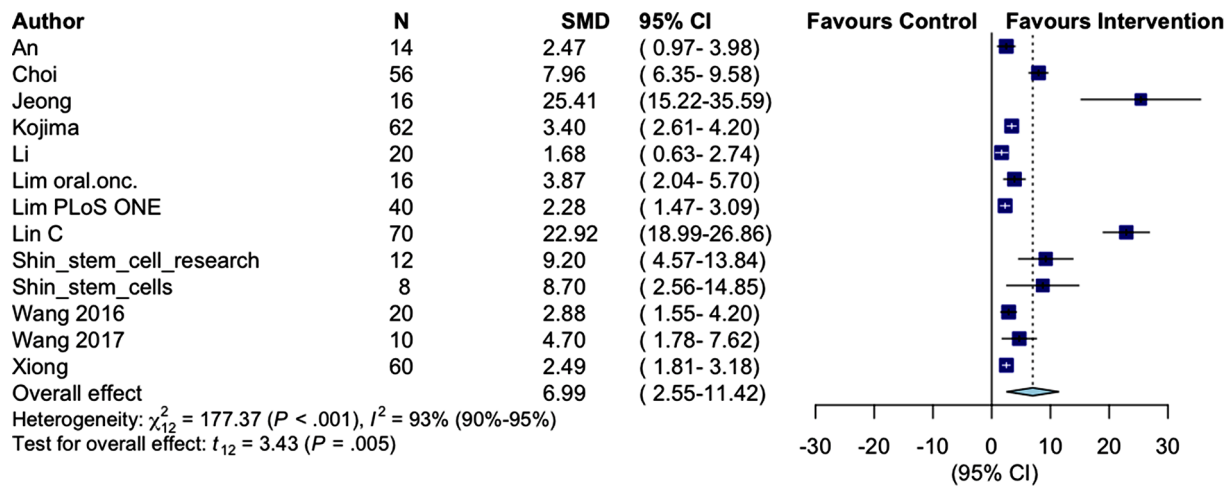


Fig. 2 Random-effects meta-analysis of the overall effect on SFR following MSC therapy. SFR: salivary flow rate; SMD: standardized mean difference; MSC: mesenchymal stem cell

improvements in vascular areas, but only for the intervention group receiving both MSCs and platelet-rich-fibrin (PRF) [38].

The Paracrine Effects of MSCs

Five studies reported on the paracrine effect of MSC treatment [25, 29, 33–35]: Xiong et al. found increased mRNA levels VEGF, HGF and COX-2 [29]; Shin et al. found higher expression of the paracrine factors BDNF, GDNF, EGF, IGF1 and NGF [34]; Shin et al. found greater expression of the growth factor FGF10 [35]; Kojima et al., found increased expressions of HGF, VEGF, COX-2 and MMP-2 [25] and Choi et al., found increased levels of EGF [33].

Homing of Systemically Transplanted MSCs

Both Li et al. and Lim et al. found that systematically transplanted MSCs could be identified in the salivary glands post transplantation [28, 31]. An et al. also administered the therapy intravenously but did not report on homing to the SG post transplantation [39].

Platelet-Rich Fibrin in Addition to MSCs

Two studies also investigated MSC+PRF [30, 38]. Chen et al. found the MSC+PRF group had significantly improvement on soft tissue defects [30], while Wang et al. found that MSC+PRF had increased levels of acinar cells, amylase, microvessels and proliferative activity and reduced levels of apoptotic cells [38].

The Effect of MSCs on Efficacy and Safety in Human Studies

One human study investigated the intervention of intraglandular autologous MSCs(AT) in a randomized, placebo-controlled phase I/II study ($n = 30$) [19]. The study found MSC treatment to be safe and reported a significant increased UWS (50% after four months, $p = .003$) including improvements in patient-reported outcomes in the group receiving MSCs. Also, a significant increase in serous gland tissue, improvements in saliva composition, and a decrease in connective were observed. See **Supplementary Results**.

Risk of Bias and Quality Assessment

The preclinical in vivo studies were assessed using SYRCLEs risk of bias assessment tool with nine questions to determine potential biases [22]. All studies involved a risk of bias, especially regarding selection bias as none of the studies reported how the randomization of intervention/control groups was performed [24–39]. Also, all studies revealed a high risk of performance bias, since none of the included studies used random housing or blinding of interventions/investigators [24–39]. All studies, except Mulyani et al., had a low risk of bias concerning attrition bias and reporting bias. Seven studies had a low risk of detection bias since they reported blinding of the outcome assessment [27, 28, 32, 33]: (1) histological examination [26, 35] (2) immunohistochemical evaluation [27, 31, 32, 37] (3) evaluation of apoptotic cells [26, 27, 32, 37, 38] (4) evaluation of cytoprotective effects (AQP5, CD31, alfa-SMA, c-Kit) [38]. Nine studies did not report blinding [24–26, 29–31, 34, 35, 37]. See Fig. 3.

	Lin C, 2011	Kojima T, 2011	Jeong J, 2013	Lim J, 2013	Lim J, 2013	Xiong X, 2014	Chen Y, 2014	An H, 2015	Li Z, 2015	Wang Z, 2016	Wang Z, 2017	Choi J, 2018	Shin H-S, 2018	Shin H-S, 2018	Elsaadany B, 2019	Mulyani S, 2019
Sequence generation (selection bias)	Yellow	Yellow	Yellow	Yellow	Yellow	Yellow	Yellow	Yellow	Yellow	Yellow	Yellow	Yellow	Yellow	Yellow	Yellow	Yellow
Baseline characteristics (selection bias)	Red	Green	Yellow	Green	Green	Yellow	Green	Green	Green	Green	Green	Green	Green	Green	Green	Green
Allocation concealment (selection bias)	Yellow	Yellow	Yellow	Yellow	Yellow	Yellow	Yellow	Yellow	Yellow	Yellow	Yellow	Yellow	Yellow	Yellow	Yellow	Yellow
Random housing (performance bias)	Red	Red	Red	Red	Red	Red	Red	Red	Red	Red	Red	Red	Red	Red	Red	Red
Blinding of interventions (performance bias)	Red	Red	Red	Red	Red	Red	Red	Red	Red	Red	Red	Red	Red	Red	Red	Red
Random outcome assessment (detection bias)	Yellow	Yellow	Yellow	Yellow	Yellow	Yellow	Yellow	Yellow	Yellow	Yellow	Yellow	Yellow	Yellow	Yellow	Yellow	Yellow
Blinding of outcome assessments (detection bias)	Red	Red	Red	Green	Green	Red	Red	Green	Red	Green	Green	Green	Red	Red	Green	Red
Incomplete outcome data (attrition bias)	Green	Green	Green	Green	Green	Green	Green	Green	Green	Green	Green	Green	Green	Green	Green	Yellow
Selective outcome reporting (reporting bias)	Green	Green	Green	Green	Green	Green	Green	Green	Green	Green	Green	Green	Green	Green	Green	Green

Fig. 3 SYRCLEs Risk of bias assessment tool. *Green* = low risk of bias, *Yellow* = unclear risk of bias, *Red* = high risk of bias

The quality of the included studies was assessed using the ARRIVE guidelines [21]. Highest score given was 9 [27, 28, 32, 33, 36, 38] and lowest score given was 5 [26]. Most studies revealed high quality with ≥ 8 points [25, 27–29, 31–36, 38, 39], but 4 studies had a total score of ≤ 7 [24, 26, 30, 37]. All studies except Jeong et al. reported sufficiently regarding study design and sample size [24, 25, 27–38]. All studies reported sufficiently on outcome measures and results and all studies were randomized, though none reported on the randomization method used and none reported how they included/excluded animals [24–39]. One study failed to report sufficiently on statistical method [24], one on experimental animals [30] and two studies failed to report sufficiently on experimental procedures [26, 37]. See **Supplementary Results**.

Discussion

In this systematic review and meta-analysis, MSC therapy demonstrated a significant effect on SG function following radiation-induced gland damage in preclinical in vivo studies. The treatment proved to be safe, with no reported adverse reactions.

We found that MSC therapy had a significant impact on the SG functioning with a significant increase in the SFR. However, there was a high heterogeneity among included

studies with differences in MSC origin, species, strains, age, radiation dose, administration route of MSC therapy, frequency of treatment and time between radiation and first treatment. The impact on SFR was significantly associated with strain and administration route. The most pronounced effect on SFR was observed when MSCs were administered intraglandular compared to systemic transplantation. Whether the effect of MSC therapy varies by strain is difficult to conclude, as only one study included Wistar [26], and one study NOD.SCID-PrckSCID [24], which both exhibited the most significant effect.

All studies but one, by Lin et al., reported elements of SG remodeling properties such as: increased density of acinar cells, more compact acinar structure, increased levels of amylase, decreased inflammation and fibrosis. If MSC therapy induces a higher proliferative activity remains uncertain. While Shin et al. reported such an effect, it was not confirmed by other studies [26, 27]. As possible modes of action, upregulation in epithelial markers (KRT7 and KRT18) and structure-related genes (SMR3A, AMY2A5, PRB1, AMY1, CLDN22, PRPMP, AMY1A, AQP5, AQP5, alfa-SMA and CD31) was identified [33, 35], while another study by Mulyani et al. found upregulation in genes encoding for proteins involved in cell migration, survival and differentiation (SDF1-CXCR4 and Bcl-2) [37].

This study also indicates that increased blood vessel regeneration and paracrine functioning participate in the

tissue repair and restoration of gland damage. Several studies reported an increase in vascular areas [24, 26, 28–30, 32]. A wide range of growth factors (VEGF, HGF, COX-2, BDNF, GDNF, EGF, IGF1, NGF, FGF10 and MMP-2) contributing to various aspects of regeneration including angiogenesis, neural regeneration and cellular proliferation were identified – ultimately supporting the repair of damaged glandular tissues.

The severity of radio-induced salivary gland damage is influenced by several factors, with radiation mean dose being a critical one [40]. The delivery of radiotherapy was greatly standardized and administered as a single dose in all the included studies, which does not resemble the clinical radiotherapy regimens for head and neck cancer which are patient-specific and often fractionated across several weeks. This is important to keep in mind, when translating the findings from this systematic review to a clinical setting. Prolonged exposure to radiation leads to cumulative damage, but unfortunately, we could not further investigate the relation between radiation duration and effect of MSC therapy due to insufficient data. The time from radiotherapy to administration of MSC might also be important, since especially the decreased levels of apoptotic cells found in several studies [26–28, 38], indicates that MSC therapy could be protective in the acute phase of radiotherapy. The timing of MSC administration varied across the studies from 0 days [28, 29, 31–33, 36, 39] to 12 weeks [25] post radiotherapy, but we observed no impact of the time from radiation to the initial MSC treatment.

Two studies that investigated the effect of intravenously administered MSC therapy also reported on homing to the SGs post transplantation, both identifying MSCs in the SGs [28, 31]. However, large-scale studies are required to further investigate the migration and homing following systemic transplantation.

PRF is a platelet-rich regenerative therapy containing a variety of growth factors and is known to promote cell proliferation [41]. Two studies investigated if additional PRF treatment improved SG function [30, 38]. Chen et al. found the MSC+PRF group had significantly improvement on soft tissue defects, but the groups were small, $n=5$ [30]. Wang et al. found that interventions with MSC, PRF or MSC+PRF improved SFR, gland and body weight, but MSC+PRF performed better regarding the regenerative outcomes [38].

In addition, the preclinical *in vivo* studies included in the meta-analysis, we identified one human, phase I/II, randomized, placebo-controlled clinical trial. The study found no serious adverse events and a significant effect on unstimulated SFR four months post MSC(AT) therapy in the treated group [19]. This is also supported by a recent study by Lynggaard et al. [16]. However, the long-term effects of

intraglandular MSC therapy in humans remain divergent [18, 42].

As a possible mode of action, Lynggaard et al. also investigated the regenerative effects of intraglandular allogeneic MSC(AT) therapy on the salivary proteome [43]. They observed an increase in proteins associated with tissue regeneration post transplantation, yet the salivary proteome did not return to a healthy state when compared to healthy controls [43].

This review is limited by the heterogeneity of methodologies and limited long-term data, hindering definite conclusions. The included studies varied in relation to included species, strains, origin of MSCs, delivering methods, radiation and follow-up regimen and study design. Prospects lie in optimizing challenges related to standardization of MSC therapy such as delivery methods, origin, and refining dosage protocols. Also, the radiotherapy regimens in the preclinical *in vivo* models were standardized and did not mirror those used in head and neck cancer patients. This lack of resemblance could potentially influence the effectiveness of MSC therapy in a clinical setting.

In conclusion, this systematic review and meta-analysis showed a significant effect of MSC therapy for restoring SG functioning and regenerating SG tissue following radiotherapy in preclinical *in vivo* studies. No serious adverse events were identified and intraglandular transplantation performed better effect than systemic transplantation. MSC therapy holds significant therapeutic potential in the treatment of radio-induced xerostomia and hypofunction, but comprehensive, randomized, clinical trials in humans are required to ascertain their efficacy in a clinical setting.

Supplementary Information The online version contains supplementary material available at <https://doi.org/10.1007/s12015-024-10700-y>.

Acknowledgements We thank Julia Menon from the Preclinical Trials.eu, The Netherlands and Siri Beier Jensen from the Department of Dentistry and Oral Health, Aarhus University, Denmark for their careful and meticulous assistance in designing the study protocol

Authors Contribution **ALFC**: investigation, data curation, writing original draft preparation, visualization **AG**: data curation, review and editing **CDL**: Conceptualization, methodology, review and editing **PMJ**: review and editing **BF**: investigation, review and editing **CH**: investigation, review and editing **BSK**: review and editing **RTL**: formal analysis, review and editing **CG**: conceptualization, methodology, review and editing **KKJ**: conceptualization, review and editing **AFN**: review and editing **CVB**: Conceptualization, methodology, review and editing.

Funding ALFC is support by the private fund Candys Foundation (reference: 2020–352–14024-1).

Open access funding provided by National Hospital

Data Availability Not applicable.

Code Availability Not applicable.

Declarations

Ethical Approval Not applicable.

Consent to Participate Not applicable.

Consent for publication Not applicable.

Conflict of interest C.D. Lynggaard, C. Grønhoj, and C. von Buchwald are co-inventors on a patent application PCT/EP2020/053878; the patent is owned by Rigshospitalet, Copenhagen University Hospital and the University of Copenhagen.

Role of the Funding Source The funder had no role in study design, in the collection, analysis or interpretation of data, in the writing of the report or in the decision to submit this article for publication. The sole responsibility for the content of this article rests with the authors.

Open Access This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit <http://creativecommons.org/licenses/by/4.0/>.

References

- Marur, S., & Forastiere, A. A. (2016). Head and Neck squamous cell carcinoma: Update on Epidemiology, diagnosis, and treatment. *Mayo Clinic Proceedings*, *91*, 386–396. <https://doi.org/10.1016/j.mayocp.2015.12.017>.
- Jensen, S. B., Vissink, A., Limesand, K. H., & Reyland, M. E. Salivary gland hypofunction and Xerostomia in Head and Neck Radiation patients. *Journal of the National Cancer Institute. Monographs* 2019;2019. <https://doi.org/10.1093/jncimonographs/lgz016>.
- Vissink, A., van Luijk, P., Langendijk, J. A., & Coppes, R. P. (2015). Current ideas to reduce or salvage radiation damage to salivary glands. *Oral Diseases*, *21*, e1–10. <https://doi.org/10.1111/odi.12222>.
- Nutting, C. M., Morden, J. P., Harrington, K. J., Urbano, T. G., Bhide, S. A., Clark, C., et al. (2011). Parotid-sparing intensity modulated versus conventional radiotherapy in head and neck cancer (PARSPORT): A phase 3 multicentre randomised controlled trial. *The Lancet Oncology*, *12*, 127–136. [https://doi.org/10.1016/S1470-2045\(10\)70290-4](https://doi.org/10.1016/S1470-2045(10)70290-4).
- Kakoei, S., Haghdoost, A. A., Rad, M., Mohammadalizadeh, S., Pourdanghan, N., Nakhaei, M., et al. (2012). Xerostomia after radiotherapy and its effect on quality of life in head and neck cancer patients. *Archives of Iranian Medicine*, *15*, 214–218.
- Memtsa, P. T., Tolia, M., Tzitzikas, I., Bizakis, J., Pisteovou-Gombaki, K., Charalambidou, M., et al. (2017). Assessment of xerostomia and its impact on quality of life in head and neck cancer patients undergoing radiation therapy. *Mol Clin Oncol*, *6*, 789–793. <https://doi.org/10.3892/mco.2017.1200>.
- Liu, X. K., Zeng, Z. Y., Hong, M. H., Zhang, A. L., Cui, N. J., & Chen, F. J. (2004). [Clinical analysis of xerostomia in patients with nasopharyngeal carcinoma after radiation therapy]. *Ai Zheng*, *23*, 593–596.
- El Agha, E., Kramann, R., Schneider, R. K., Li, X., Seeger, W., Humphreys, B. D., et al. (2017). Mesenchymal stem cells in Fibrotic Disease. *Cell Stem Cell*, *21*, 166–177. <https://doi.org/10.1016/j.stem.2017.07.011>.
- Ozdemir, T., Fowler, E. W., Hao, Y., Ravikrishnan, A., Harrington, D. A., Witt, R. L., et al. (2016). Biomaterials-based strategies for salivary gland tissue regeneration. *Biomater Sci*, *4*, 592–604. <https://doi.org/10.1039/c5bm00358j>.
- Galipeau, J., Krampera, M., Barrett, J., Dazzi, F., Deans, R. J., DeBrujin, J., et al. (2015). International Society for Cellular Therapy perspective on immune functional assays for mesenchymal stromal cells as potency release criterion for advanced phase clinical trials. *Cytotherapy*, *18*, 151–159. <https://doi.org/10.1016/j.jcyt.2015.11.008>.
- Singer, N. G., & Caplan, A. I. (2011). Mesenchymal stem cells: Mechanisms of inflammation. *Annu Rev Pathol Mech Dis*, *6*, 457–478. <https://doi.org/10.1146/annurev-pathol-011110-130230>.
- Caplan, A. I., & Dennis, J. E. (2006). Mesenchymal stem cells as trophic mediators. *Journal of Cellular Biochemistry*, *98*, 1076–1084. <https://doi.org/10.1002/jcb.20886>.
- Caplan, A. I., & Correa, D. (2011). The MSC: An injury drug-store. *Cell Stem Cell*, *9*, 11–15. <https://doi.org/10.1016/j.stem.2011.06.008>.
- Coppes, R. P., & Stokman, M. A. (2011). Stem cells and the repair of radiation-induced salivary gland damage. *Oral Diseases*, *17*, 143–153. <https://doi.org/10.1111/j.1601-0825.2010.01723.x>.
- Jensen, D. H., Oliveri, R. S., Trojahn Kølle, S. F., Fischer-Nielsen, A., Specht, L., Bardow, A., et al. (2014). Mesenchymal stem cell therapy for salivary gland dysfunction and xerostomia: A systematic review of preclinical studies. *Oral Surg Oral Med Oral Pathol Oral Radiol*, *117*, 335–342e1. <https://doi.org/10.1016/j.oooo.2013.11.496>.
- Lynggaard, C. D., Grønhoj, C., Christensen, R., Fischer-Nielsen, A., Melchior, J., Specht, L., et al. (2022). Intraglandular off-the-Shelf allogeneic mesenchymal stem cell treatment in patients with Radiation-Induced Xerostomia: A Safety Study (MESRIX-II). *Stem Cells Transl Med*, *11*, 478–489. <https://doi.org/10.1093/stcltm/szac011>.
- Blitzer, G. C., Glazer, T., Burr, A., Gustafson, S., Ganz, O., Meyers, R., et al. (2023). Marrow-derived autologous stromal cells for the restoration of salivary hypofunction (MARSH): A pilot, first-in-human study of interferon gamma-stimulated marrow mesenchymal stromal cells for treatment of radiation-induced xerostomia. *Cytotherapy*, *25*, 1139–1144. <https://doi.org/10.1016/j.jcyt.2023.07.009>.
- Lynggaard, C. D., Grønhoj, C., Jensen, S. B., Christensen, R., Specht, L., Andersen, E., et al. (2022). Long-term safety of treatment with autologous mesenchymal stem cells in patients with Radiation-Induced Xerostomia: Primary results of the MESRIX Phase I/II Randomized Trial. *Clinical Cancer Research*, *28*, 2890–2897. <https://doi.org/10.1158/1078-0432.CCR-21-4520>.
- Grønhoj, C., Jensen, D. H., Vester-Glowinski, P., Jensen, S. B., Bardow, A., Oliveri, R. S., et al. (2018). Safety and efficacy of mesenchymal stem cells for Radiation-Induced Xerostomia: A randomized, placebo-controlled phase 1/2 trial (MESRIX). *International Journal of Radiation Oncology Biology Physics*, *101*, 581–592. <https://doi.org/10.1016/j.ijrobp.2018.02.034>.
- Jansson, P. M., Lynggaard, C. D., Carlander, A. F., Jensen, S. B., Follin, B., Hoeg, C., et al. (2022). Mesenchymal stromal/stem cell therapy for radiation-induced salivary gland hypofunction in

- animal models: A protocol for a systematic review and meta-analysis. *Syst Rev*, 11, 72. <https://doi.org/10.1186/s13643-022-01943-2>.
21. Percie du Sert, N., Hurst, V., Ahluwalia, A., Alam, S., Avey, M. T., Baker, M., et al. (2020). The ARRIVE guidelines 2.0: Updated guidelines for reporting animal research. *Plos Biology*, 18, e3000410. <https://doi.org/10.1371/journal.pbio.3000410>.
 22. Hooijmans, C. R., Rovers, M. M., de Vries, R. B. M., Leenaars, M., Ritskes-Hoitinga, M., & Langendam, M. W. (2014). SYRCLÉ's risk of bias tool for animal studies. *Bmc Medical Research Methodology*, 14, 43. <https://doi.org/10.1186/1471-2288-14-43>.
 23. https://handbook-5-1.cochrane.org/chapter_9/9_2_3_2_the_standardized_mean_difference.htm. (n.d).
 24. Lin, C. Y., Chang, F. H., Chen, C. Y., Huang, C. Y., Hu, F. C., Huang, W. K., et al. (2011). Cell therapy for salivary gland regeneration. *Journal of Dental Research*, 90, 341–346. <https://doi.org/10.1177/0022034510386374>.
 25. Kojima, T., Kanemaru, S. I., Hirano, S., Tateya, I., Ohno, S., Nakamura, T., et al. (2011). Regeneration of radiation damaged salivary glands with adipose-derived stromal cells. *The Laryngoscope*, 121, 1864–1869. <https://doi.org/10.1002/lary.22080>.
 26. Jeong, J., Baek, H., Kim, Y. J., Choi, Y., Lee, H., Lee, E., et al. (2013). Human salivary gland stem cells ameliorate hyposalivation of radiation-damaged rat salivary glands. *Experimental & Molecular Medicine*, 45, e58. <https://doi.org/10.1038/emm.2013.121>.
 27. Lim, J. Y., Yi, T., Choi, J. S., Jang, Y. H., Lee, S., Kim, H. J., et al. (2013). Intraglandular transplantation of bone marrow-derived clonal mesenchymal stem cells for amelioration of post-irradiation salivary gland damage. *Oral Oncology*, 49, 136–143. <https://doi.org/10.1016/j.oraloncology.2012.08.010>.
 28. Lim, J. Y., Ra, J. C., Shin, I. S., Jang, Y. H., An, H. Y., Choi, J. S., et al. (2013). Systemic transplantation of human adipose tissue-derived mesenchymal stem cells for the regeneration of irradiation-induced salivary gland damage. *PLoS One*, 8, e71167. <https://doi.org/10.1371/journal.pone.0071167>.
 29. Xiong, X., Shi, X., & Chen, F. (2014). Human adipose tissue-derived stem cells alleviate radiation-induced xerostomia. *International Journal of Molecular Medicine*, 34, 749–755. <https://doi.org/10.3892/ijmm.2014.1837>.
 30. Chen, Y., Niu, Z., Xue, Y., Yuan, F., Fu, Y., & Bai, N. (2014). Improvement in the repair of defects in maxillofacial soft tissue in irradiated minipigs by a mixture of adipose-derived stem cells and platelet-rich fibrin. *British Journal of Oral and Maxillofacial Surgery*, 52, 740–745. <https://doi.org/10.1016/j.bjoms.2014.06.006>.
 31. Li, Z., Wang, Y., Xing, H., Wang, Z., Hu, H., An, R., et al. (2015). Protective efficacy of intravenous transplantation of adipose-derived stem cells for the prevention of radiation-induced salivary gland damage. *Archives of Oral Biology*, 60, 1488–1496. <https://doi.org/10.1016/j.archoralbio.2015.07.016>.
 32. Wang, Z., Ju, Z., He, L., Li, Z., Liu, Y., & Liu, B. (2017). Intraglandular transplantation of adipose-derived stem cells for the Alleviation of Irradiation-Induced parotid gland damage in Miniature pigs. *Journal of Oral and Maxillofacial Surgery*, 75, 1784–1790. <https://doi.org/10.1016/j.joms.2016.08.001>.
 33. Choi, J. S., An, H. Y., Shin, H. S., Kim, Y. M., & Lim, J. Y. (2018). Enhanced tissue remodelling efficacy of adipose-derived mesenchymal stem cells using injectable matrices in radiation-damaged salivary gland model. *Journal of Tissue Engineering and Regenerative Medicine*, 12, e695–706. <https://doi.org/10.1002/term.2352>.
 34. Shin, H. S., Lee, S., Hong, H. J., Lim, Y. C., Koh, W. G., & Lim, J. Y. (2018). Stem cell properties of human clonal salivary gland stem cells are enhanced by three-dimensional priming culture in nanofibrous microwells. *Stem Cell Research & Therapy*, 9, 74. <https://doi.org/10.1186/s13287-018-0829-x>.
 35. Shin, H. S., Lee, S., Kim, Y. M., & Lim, J. Y. (2018). Hypoxia-activated adipose mesenchymal stem cells prevents Irradiation-Induced Salivary Hypofunction by enhanced paracrine effect through fibroblast growth factor 10. *Stem Cells*, 36, 1020–1032. <https://doi.org/10.1002/stem.2818>.
 36. Elsaadany, B., Zakaria, M., & Mousa, M. R. (2019). Transplantation of bone marrow-derived mesenchymal stem cells preserve the salivary glands structure after Head and Neck Radiation in rats. *Open Access Maced J Med Sci*, 7, 1588–1592. <https://doi.org/10.3889/oamjms.2019.350>.
 37. Mulyani, S. W. M., Astuti, E. R., Wahyuni, O. R., Ernawati, D. S., & Ramadhani, N. F. (2019). Xerostomia Therapy due to Ionized Radiation using preconditioned bone marrow-derived mesenchymal stem cells. *Eur J Dent*, 13, 238–242. <https://doi.org/10.1055/s-0039-1694697>.
 38. Wang, Z., Xing, H., Hu, H., Dai, T., Wang, Y., Li, Z., et al. (2016). Intraglandular transplantation of adipose-derived stem cells combined with platelet-rich fibrin extract for the treatment of irradiation-induced salivary gland damage. *Exp Ther Med*, 15, 795–805. <https://doi.org/10.3892/etm.2017.5497>.
 39. A, H. Y., C, H. S. S. J. S., & K, H. J. (2015). Adipose mesenchymal stem cell secretome modulated in hypoxia for remodeling of radiation-induced salivary gland damage. *PLoS One*, 10, e0141862. <https://doi.org/10.1371/journal.pone.0141862>.
 40. Eisbruch, A., Ten Haken, R. K., Kim, H. M., Marsh, L. H., & Ship, J. A. (1999). Dose, volume, and function relationships in parotid salivary glands following conformal and intensity-modulated irradiation of head and neck cancer. *International Journal of Radiation Oncology Biology Physics*, 45, 577–587. [https://doi.org/10.1016/s0360-3016\(99\)00247-3](https://doi.org/10.1016/s0360-3016(99)00247-3).
 41. Dohan, D. M., Choukroun, J., Diss, A., Dohan, S. L., Dohan, A. J. J., Mouhyi, J., et al. (2006). Platelet-rich fibrin (PRF): A second-generation platelet concentrate. Part I: Technological concepts and evolution. *Oral Surgery, Oral Medicine, Oral Pathology, Oral Radiology and Endodontics*, 101, e37–44. <https://doi.org/10.1016/j.tripleo.2005.07.008>.
 42. Kronberg Jakobsen, K., Duch Lynggaard, C., Paaske, N., Fenger Carlander, A. L., Kastrup, J., Hauge Werner, A., et al. (2023). *Long-term outcome following treatment with allogeneic mesenchymal stem/Stromal cells for Radiation-Induced Hyposalivation and Xerostomia*. *Stem Cells Transl Med*.
 43. Lynggaard, C. D., Jersie-Christensen, R., Juhl, M., Jensen, S. B., Grønhoj, C., Melchior, J., et al. (2022). Intraglandular mesenchymal stem cell treatment induces changes in the salivary proteome of irradiated patients. *Communications Medicine*, 2, 160. <https://doi.org/10.1038/s43856-022-00223-3>.

Publisher's Note Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

Supplementary Results

Table 1. Random-effects model sub analyses on categorical heterogeneity factors.

Subgroup analysis	Classifications	No. of studies	SMD	95% CI	p-value*
Species	Mice	10	6.3	1.8-10.8	0.63
	Rats	2	13.4	-132.0-158.7	
	Miniature pigs	1	4.7	1.7-7.6	
Strain	NOD.SCID-PrckSCID	1	22.9	19.0-26.9	<0.001*
	C57BL/6	3	2.9	0.1-5.7	
	Whistar	1	25.4	15.2-35.6	
	CH3	6	5.0	1.6-8.4	
	Sprague-Dawlay	1	2.5	1.8-3.2	
	Missing	1	4.7	1.8-7.6	
Sex	Female	9	4.3	2.1-6.4	0.47
	Male	2	13.4	-132.0-158.7	
	Unknown	2	13.3	-107.7-134.3	
Administration route	Intraglandular	10	8.6	2.9-14.2	0.01*
	Intravenously	3	2.1	1.2-3.0	

*significant value.

Table 2. Meta-regression sub analyses on continuous heterogeneity factors

Subgroup	Estimate	95% CI	p-value*
Age, weeks	-0.3	-0.9-0.3	0.3
Radiation dose, Gy	-0.7	-0.7-2.1	0.3
Frequency of treatment	-0.6	-1.7-0.5	0.2
Time between radiation and first treatment	0.0	-0.2-0.1	0.7

*significant value.

Table 3. Human study characteristic.

Author (year)	Study design	Groups	Irradiation	Days from radiation to MSC treatment	MSC type, concentration and administration route	Statistical analysis	Functional outcome	Molecular outcome
Grønhoj, C. et al. (2018)	Prospective, randomized, blinded, controlled phase 1/2 trial	1. placebo, n = 30 2. MSCs(AT), n = 30	Mean Gy dose to single gland between 11.4 Gy and 71 Gy. 27 patients received both RT and chemotherapy, 3 received only RT.	Between 2.8 and 6.5 years. Median interval was 4.1 years.	MSC(AT)h (autologous). Dose was 2.8×10^6 cells x the volume of the gland (cm ³). Intraglandular injection, ultrasound guided (submandibular glands)	Sample size was calculated using a non-paired <i>t</i> test. Within-group comparisons were performed with the Wilcoxon signed-rank test, and between-group comparisons were performed with the Mann-Whitney U test. Nonparametric statistics was evaluated by Shapiro-Wilks tests. Significance level was $P < .05$.	Unstimulated whole SFR was significantly increased in group 2 compared to baseline at both one-month and four months after treatment. Control group did not see this increase. The net scores between the two groups were similar. Group 2 showed significant improvements in patient reported outcome measures such as VAS score and xerostomia questionnaire regarding thirst and oral dryness. No adverse events in group 2 at 1 year.	19 samples were evaluated, 11 were deemed not suitable for evaluation. A significant increase in serous gland tissue in group 2 was found compared to group 1. No significant differences was observed in the fractions of mucinous to serous tissue, mucinous tissue, glandular tissue adipose tissue.

Figure 2. Evaluation of reporting quality according to ARRIVE guidelines. *: 1=Study design; 2= Sample size; 3= Inclusion and exclusion criteria; 4= Randomization, **none of the studies reported randomization method; 5=Blinding; 6=Outcome measures; 7=Statistical methods; 8=Experimental animals; 9=Experimental procedures; 10=Results. One point was given if the criteria was sufficiently fulfilled, maximum total quality score possible was 10.

Author (Year)	1*	2*	3*	4*	5**	6*	7*	8*	9*	10*	Total quality score
Lin, C et al. (2011)	1	1	0	1**	0	1	0	1	1	1	7
Kojima, T. et al. (2011)	1	1	0	1**	0	1	1	1	1	1	8
Jeong, J. et al. (2013)	0	0	0	1**	0	1	1	1	0	1	5
Lim, J. et al. (2013, oral)	1	1	0	1**	1	1	1	1	1	1	9
Lim, J. et al. (2013, PLoS ONE)	1	1	0	1**	1	1	1	1	1	1	9
Xiong, X. et al. (2014)	1	1	0	1**	0	1	1	1	1	1	8
Chen, Y. and Niu, Z. et al. (2014)	1	1	0	1**	0	1	1	0	1	1	7
An, H. et al. (2015)	1	1	0	1**	1	1	1	1	1	1	8
Li, Z. et al. (2015)	1	1	0	1**	0	1	1	1	1	1	8
Wang, Z. et al. (2016)	1	1	0	1**	1	1	1	1	1	1	9
Wang, Z. et al. (2017)	1	1	0	1**	1	1	1	1	1	1	9
Choi, J. et al. (2018)	1	1	0	1**	1	1	1	1	1	1	9
Shin, H. et al. (2018)	1	1	0	1**	0	1	1	1	1	1	8
Shin, H. et al. (2018)	1	1	0	1**	0	1	1	1	1	1	8
Elsaadany, B. et al. (2019)	1	1	0	1**	1	1	1	1	1	1	9
Mulyani, S. et al. (2019)	1	1	0	1**	0	1	1	1	0	1	7

Title

Long-term effectiveness and safety of mesenchymal stromal cell therapy for radiation-induced hyposalivation in head and neck cancer survivors: A randomised, phase-2, trial

Authors

Amanda-Louise Fenger Carlander^{1,2*}, Kathrine Kronberg Jakobsen^{1*}, Tobias Todsen¹, Natasja Paaske¹, Anne Kathrine Østergaard Madsen¹, Simone Kloch Bendtsen¹, Jens Kastrup³, Jeppe Friberg⁴, Charlotte Duch Lynggaard¹, Anne Werner Hauge⁵, Robin Christensen^{2,6}, Christian Grønhøj¹, Christian von Buchwald¹

Affiliation

*Shared first authorship

¹Department of Otorhinolaryngology, Head and Neck Surgery & Audiology, Copenhagen University Hospital - Rigshospitalet, Denmark.

²Section for Biostatistics and Evidence-Based Research, the Parker Institute, Copenhagen University Hospital - Bispebjerg and Frederiksberg, Denmark

³Cardiology Stem Cell Centre, The Heart Centre, Copenhagen University Hospital - Rigshospitalet, Denmark

⁴Department of Oncology, Copenhagen University Hospital - Rigshospitalet, Denmark

⁵Department of Clinical Immunology, Copenhagen University Hospital - Rigshospitalet, Denmark

⁶Research Unit of Rheumatology, Department of Clinical Research, University of Southern Denmark, Odense University Hospital, Denmark.

Address for Correspondence

Amanda-Louise Fenger Carlander, MD, Ph.D.-fellow

Department of Otorhinolaryngology, Head and Neck Surgery and Audiology, Copenhagen University Hospital – Rigshospitalet

Tel: 00 45 30 22 31 02, E-mail: amanda-louise.fenger.carlander@regionh.dk

Conflict of Interest

Charlotte Lynggaard, Christian Grønhøj, and Christian von Buchwald are co-inventors on a patent application for “Stem cell therapy for patients with salivary gland dysfunction”, PCT/EP2020/053878. The patent is owned by Rigshospitalet, Copenhagen University Hospital and University of Copenhagen. Jens Kastrup, Annette Ekblond, and Mandana Haack-Sørensen hold a patent application for the investigated stem cell product “Stem cell therapy based on adipose-derived stem cells”, WO2017068140. None of the other co-authors declare conflicts of interest.

Funding

Funding for this study was received from the non-profit Candys Foundation (grant number: 2020-352) and Copenhagen University Hospital (unnumbered grant). The Section for Biostatistics and Evidence-Based Research at the Parker Institute is supported by the Oak Foundation (Grant: OCAY-18-774-OFIL).

Keywords

Mesenchymal Stem/stromal Cells; Mesenchymal Stromal Cells; MSC; Xerostomia; hyposalivation, Radiotherapy; Head and Neck cancer

Translational Relevance

The most common side effect of radiotherapy for head and neck cancer is radiation-induced hyposalivation, which impacts swallowing, the oral health and the quality of life in cancer survivors. Currently, only options for symptomatic relief exists, but mesenchymal stromal cells (MSCs) have been suggested as a regenerative treatment. We have previously shown that intraglandular treatment with MSCs hold significant potential in restoring the salivary gland function, and this study is the first long-term results of a randomised, placebo-controlled phase II trial investigating allogeneic MSCs for radiation-induced hyposalivation. The study revealed that MSC treatment improved the subjective feeling of dry mouth. However, treatment with MSCs was not superior to placebo in restoring salivary gland function evaluated by objective salivary flow rate measurements. This study underscores the clinical potential of MSC treatment in cancer rehabilitation, however, identification of patient subgroups along with implementation of repeated MSC treatment may contribute to refining therapeutic strategies.

Abstract

Background: The long-term effect of adipose-derived mesenchymal stromal cells (ASCs) to restore radiation-induced salivary gland hypofunction in previous head and neck cancer patients have not been validated in larger, randomised settings.

Methods: The study was the 12-months follow-up of a randomised, phase-2 trial, including patients with hyposalivation. Patients were randomised to receive allogeneic ASCs or placebo in the submandibular glands. Primary endpoint was unstimulated whole saliva (UWS) followed by stimulated whole saliva, patient-reported outcomes (European Organization for Research and Treatment of Cancer Quality of Life Questionnaire, Head and Neck Module and the Xerostomia Questionnaire) and safety (serious adverse events and immune response).

Results: Of the 120 enrolled patients, 117 (97.5%) were assessed at 12 months. Treatment with ASCs did not increase UWS compared to placebo: the increase in UWS was 0.02 mL/min (95% CI 0.01 to 0.04) in the ASC group and 0.02 mL/min (95% CI 0 to 0.03) in the placebo group, $p=0.56$. ASCs significantly reduced the symptom burden for dry mouth with -10.07 units (95% CI -13.39 to -6.75) compared to -4.15 units (95% CI -7.46 to -0.84) in the placebo group, $p=0.01$. Compared to placebo, ASCs did not improve sticky saliva (-9.27 vs. -4.55 units, $p=0.13$), swallowing (-4.50 vs. 3.49 units, $p=0.5$) or xerostomia -3.12 vs. -2.74 units, $p=0.82$). Treatment was safe and associated with a transient immune response.

Conclusion: Intraglandular ACS therapy in the submandibular glands significantly relieved subjective dry mouth symptoms. Both ASCs and placebo increased UWS, but ASCs did not prove superior to placebo in restoring salivary gland function, based on salivary flow rate.

Introduction

Head and neck cancer (HNC) affects globally more than 900,000 individuals each year, and with an increasing incidence¹. Most patients are treated with radiation therapy (RT) either as a single modality or in combination with other treatment modalities^{2,3}. Despite a reduction in salivary gland radiation dose following introduction of intensity-modulated RT (IMRT), the salivary glands are still often affected^{4,5}. The salivary glands are especially susceptible to radiation which induces both acute and chronic salivary gland damage, including inflammation, fibrosis, and loss of acinar and progenitor cells, ultimately leading to salivary gland hypofunction^{6,7}. The most common side effect of RT is hyposalivation and xerostomia, or dry mouth syndrome, and affects more than 80% of head and neck cancer patients treated with RT⁸. Xerostomia impacts not only the quality of life but also the health of head and neck cancer survivors: hyposalivation is associated with an increased risk of oral infections, discomfort, pain, swallowing difficulties, impaired speech, and sleep disturbances⁹⁻¹². Current treatment modalities for xerostomia only provide symptomatic relief¹³, emphasizing the urgency for novel therapeutic opportunities.

Mesenchymal stromal cells (MSCs) have been investigated as a possible therapeutic and disease-modifying treatment to restore radiation-induced salivary gland damage and hypofunction¹⁴. This is attributed to their regenerative potential which includes mesodermal differentiation, efferocytosis, and microenvironmental reorganization as well as trophic properties encompassing immunosuppression, antifibrosis, and angiogenic effects^{15,16}. The mode of action is not fully understood but they are assumed to function through a “hit and run” mechanism¹⁷ while also escaping the innate immune system¹⁸ enabling an easily accessible use of “off the shelf” donor MSCs. Intraglandular MSC therapy has shown to improve quality of life and enhance the salivary flow rate (SFR) in both preclinical, animal studies¹⁹ and in humans²⁰⁻²³. We recently conducted a randomised, clinical trial, demonstrating that intraglandular allogeneic ASC therapy was safe and was associated with an increase of 38% in saliva production, although not superior to placebo 4 months following intervention²². Also, long-term outcomes indicate a persistent effect, however, this has not been validated in a larger, randomised trial setting^{24,25}.

Therefore, the aim of this phase 2 trial was to evaluate the long-term efficacy and safety of intraglandular treatment with allogeneic ASCs for radiation-induced hyposalivation in previous patients with HNC from our randomised clinical trial²².

Materials and methods

Trial design

The MESRIX-III trial was a single-centre, investigator-initiated, randomised, placebo-controlled, double-blinded trial conducted at the Department of Otorhinolaryngology, Head and Neck Surgery & Audiology, University Hospital of Copenhagen. Patients were followed for one year and the present study constitutes the long-term follow-up of the trial²². The trial was approved by the National Ethics Committee (1802872), The Danish Data Protection Agency (P-2020-1164), and the Danish Medical Agency (EudraCT: 2018-000348-24). The study protocol was previously published²⁶ and registered at ClinicalTrials.gov (NCT04776538). The Good Clinical Practice unit (GCP) at University of Copenhagen monitored the trial. In accordance with the Helsinki Declaration included patients provided both verbal and written consent to participate.

Participants

Patients eligible for inclusion encompassed: 1) age between 18-75 years 2) previously RT treated for head and neck cancer 3) clinically hyposalivation with unstimulated whole saliva flow rate (UWS) 0.05-0.25 mL/min 3), and a minimum of two years since diagnosis or recurrence^{22,26}. Patients were excluded if they received xerogenic medicine, had other salivary gland disease, other cancer within the last four years, recent alcohol abuse or smoking history, penicillin or streptomycin allergy, pregnancy, or breastfeeding.

Interventions

Patients were randomised 1:1 to receive intraglandular injection with ASC therapy or injection with a placebo solution in both submandibular glands. Into each gland, an amount of 0.5 mL was injected ultrasound guided without using local anaesthesia, corresponding to a dose of 25×10^6 cells for patients receiving ASCs per gland. The injections were, as far as possible, distributed in a fan-shaped pattern within each gland²². The Cardiology Stem Cell Centre (CSCC) provided the ASCs according to good manufacturing practice (GMP) from three healthy donors^{27,28}, with each vial containing cells from one individual donor. Placebo consisted of the freezing media for ASCs CryoStor10 (BiolifeSolutions) with 10% dimethyl sulfoxide (DMSO). The interventions of the study are previously well described^{22,26}.

UWS and stimulated whole saliva flow rate (SWS) was evaluated by sialometry. Patient-reported outcomes on quality of life were evaluated by two questionnaires, both the xerostomia questionnaire (XQ) and the European Organization for Research and Treatment of Cancer Quality of Life Questionnaire, Head and Neck Module (EORTC QLQ-H&N35) domains for dry mouth (HNDR), sticky saliva (HNSS), and swallowing (HNSW). Immune response was evaluated by the development of de novo human leukocyte antigen (HLA) donor specific antibodies. Safety was evaluated by serious adverse events (SAEs). The assessments of the study are previously well described^{22,26}.

Objectives and Outcomes

The primary objective of this study was to assess the effectiveness of ASCs compared to placebo on UWS, measured as change in mL/min from baseline to 12 months follow-up.

Secondary objectives were to assess ASCs compared to placebo from baseline to 12 months on the following: 1) the effectiveness measured as change in SWS in mL/min 2) the impact on patient-reported outcomes (XQ, EORCT QLQ-H&N35 domains: HNDR, HNSS, HNSW) measured as change in sum score 3) the safety measured as development of SAEs and 4) immune response measured as development of donor specific antibodies. An increase or decrease in donor specific antibodies was defined as a change in mean fluorescence intensity sum of donor specific antibodies ≥ 3000 .

Sample size, randomisation, and blinding

A total of 120 patients were included yielding a power of 80% and an alpha of 0.05. Patients were randomised 1:1 according to their treatment order and a predetermined block randomisation code ensured no treatment clustering. The sample size and randomisation are previously well described^{22,26}. The trial was kept blinded to sponsor, investigators, and research staff until the 4 months follow-up as previously described^{22,26}. The 12 months follow-up was conducted unblinded to treatment allocation to both study personnel and participants.

Statistical methods

Prior to conducting the analysis, a statistical analysis plan was completed, **Supplementary Statistical Analysis Plan**. The primary analyses were based on the intention to treat (ITT) principle; participants allocated to a treatment group (ASC and Placebo, respectively) was followed up, assessed, and analysed as members of that group, irrespective of their adherence to the planned course of treatment (i.e., independent of withdrawals and cross-over phenomena). Descriptive statistics: Categorical data was reported as number and percentages and continuous data as means and standard deviations (SD). Multilevel repeated measurements mixed effects model with participants as a random effects factor and the outcome variable as a dependent variable was used. The time (3 levels; month 0, 4, 12) was set as a fixed effect factor based on a restricted maximum likelihood model. While adjusting for baseline levels, the model facilitated comparisons between the time points and the trajectory over the entire study period. Inferential statistics is reported as least square means with standard errors and the differences between them with 95% confidence intervals (95%CI). All 95%CIs and P values are two sided. Safety was reported in a descriptive manner. A non-responder sensitivity analysis was performed with baseline observations carried forward to replace missing data. The statistical analyses were performed in SAS and R Studio.

Data availability

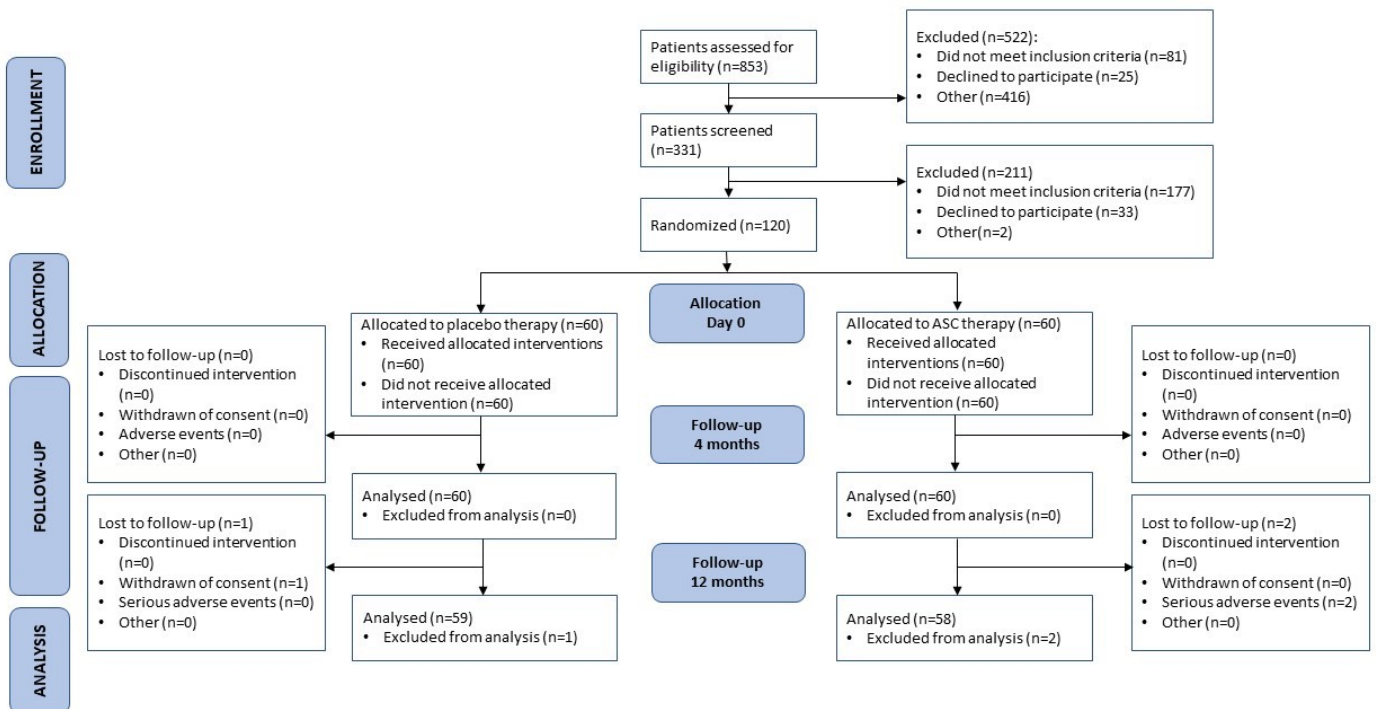
Due to sensitivity reasons, the data from this study is not publicly accessible. However, upon reasonable request to the corresponding author data are available.

Results

Study flow and baseline characteristics

A total of 120 patients with previous head and neck cancer were included at the Department of Otorhinolaryngology, Head and Neck Surgery & Audiology, Copenhagen University Hospital and received the intended intervention, while 117 were assessed at the 12-month follow-up visit. **Figure 1** illustrates the progress through the trial from enrolment to the 12-month follow-up (January 19th, 2021, to February 12th, 2024). Three patients were lost to follow-up: one patient (placebo) withdrew consent, and two patients (ASCs) were excluded due to new primary cancer in accordance with the protocol withdrawal criteria²⁶.

Figure 1. CONSORT Flow diagram.



The two groups were comparable at baseline²². Most patients had oropharyngeal cancer, 107 (89%), the mean age was 61.4 years (SD: 7.1 years), 88 (73%) were males, 89 (74%) had stage I-II, and were 53 (44%) never smokers. UWS at baseline was 0.13 mL/min (SD: 0.01 mL/min). See **Table 1**.

Table 1. Baseline characteristics in the intention-to-treat population.

Characteristics	ASC (n=60)	Placebo (n=60)
Age (years)	61.0 (7.4)	61.8 (6.8)
Male, sex, no. (%)	41 (68)	47 (78)
Performance score (0-1), no. (%)	60 (100)	60 (100)
Previous smoking history		
0 pack years	29 (48)	24 (40)
1-10 pack years	7 (12)	6 (10)
>10 pack years	24 (40)	30 (50)
Cancer location, no. (%)		
Oropharynx	52 (87)	55 (92)
Other	7 (12)	5 (8)
UICC8 stage, no. (%)		
I-II	45 (75)	44 (73)
III-IV	15 (25)	16 (27)
Mean radiation dose to the four large salivary gland, Gy	39.9 (7.1)	40.5 (8.9)
Salivary flow rate, mL/min		
Unstimulated whole saliva flow rate	0.13 (0.05)	0.13 (0.06)
Stimulated whole saliva flow rate	1.14 (0.59)	0.99 (0.60)
EORCT QLQ-H&N35 (0-100)		
HNDR	81.9 (23.4)	76.3 (23.2)
HNSS	55.6 (38.2)	57.1 (37.7)
HNSW	33.9 (22.8)	31.8 (19.9)
XQ (0-100)	51.0 (20.5)	47.9 (19.9)

Estimates are reported as means and SDs unless otherwise indicated.

Abbreviations: ASC, adipose-derived mesenchymal stem cells; EORCT QLQ-H&N35, The European Organization for Research and Treatment of Cancer Quality of Life Questionnaire; Gy, gray; HNDR, domains for dry mouth; HNSS, domains for sticky saliva; HNSW, domains for swallowing; no., number; UICC, union for international cancer control; XQ, xerostomia questionnaire.

Salivary gland function

From baseline to 12 months, an increase in UWS of was observed in the ASC group (0.02 mL/min; 95% CI [0.01 to 0.04]) and in the placebo group (0.02 mL/min; 95% CI [0 to 0.03]), with no difference in change between the groups (0.01 mL/min; 95% CI [-0.02 to 0.03]), see **Table 2**. The UWS increased from 0.13 mL/min (95% CI 0.11 to 0.15) to 0.16 mL/min (95% CI 0.14 to 0.18) in the ASC group, and from 0.13 mL/min (95% CI 0.11 to 0.15) to 0.17 mL/min (95% CI 0.1 to 0.19) in the placebo group, see **Figure 2**. This

corresponded to an increase at 12 months of 25% (95% CI 18 to 34) in the ASC group and 27% (95% CI 20 to 33) in the placebo group compared to baseline. At 12 months, 23 ASC patients and 18 placebo patients had an increase in UWS >30% with 6 in the ASC group and 4 in the placebo group acquiring a normal saliva flow rate of ≥ 0.3 mL/min.

Table 2. Comparison of change from in primary outcomes from baseline to 12 months follow-up (trial secondary end point).

	ASC (n = 60)	Placebo (n = 60)	Difference (95% CI)	p-value*
Change From Baseline to 12 months				
Unstimulated saliva flow rate, mL/min	0.02 (0.01)	0.02 (0.01)	0.01 (-0.02 to 0.03)	0.56
Stimulated saliva flow rate, mL/min	0.04 (0.03)	0.08 (0.03)	-0.04 (-0.17 to 0.04)	0.35
XQ-summary score (0-100)	-3.12 (1.18)	-2.74 (1.17)	-0.38 (-3.68 to 2.91)	0.82
<i>EORCT QLQ-H&N35 (score 0-100):</i>				
HNDR	-10.07 (1.68)	-4.15 (1.67)	-5.93 (-10.62 to -1.22)	0.01*
HNSS	-9.27 (2.17)	-4.55 (2.16)	-4.72 (-10.78 to 1.35)	0.13
HNSW	-4.50 (1.05)	-3.49 (1.05)	-1.00 (-3.94 to 1.94)	0.5

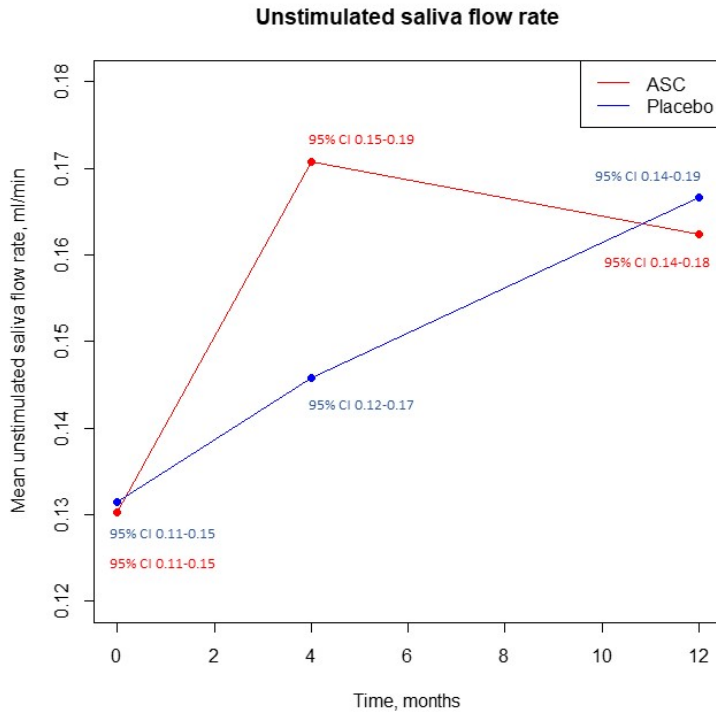
Values are least squares means with standard errors for each group; the difference is reported with 95% confidence intervals (95% CIs). Estimates are derived from the repeated measures, mixed effects models.

*Significant p-value.

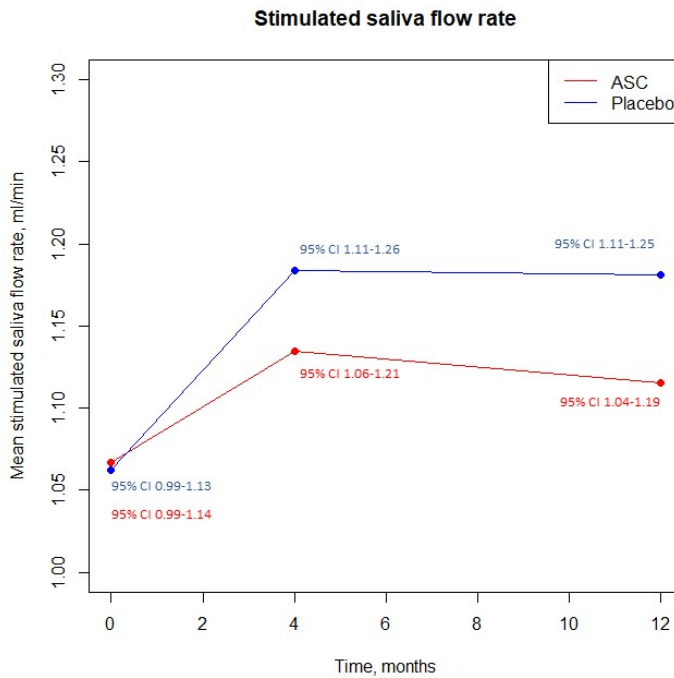
Abbreviations: ASC, adipose-derived mesenchymal stem cells; CI, confidence interval; EORCT QLQ-H&N35, The European Organization for Research and Treatment of Cancer Quality of Life Questionnaire; HNDR, domains for dry mouth; HNSS, domains for sticky saliva; HNSW, domains for swallowing.

Figure 2. Change in salivary gland function measured over the 12 months study period. **A.** Unstimulated whole salivary flow rate (UWS) **B.** Stimulated whole saliva flow rate (SWS)

A.



B.



Abbreviations: ASC, adipose-derived mesenchymal stem cells; CI, confidence interval, min, minute; mL, milliliters.

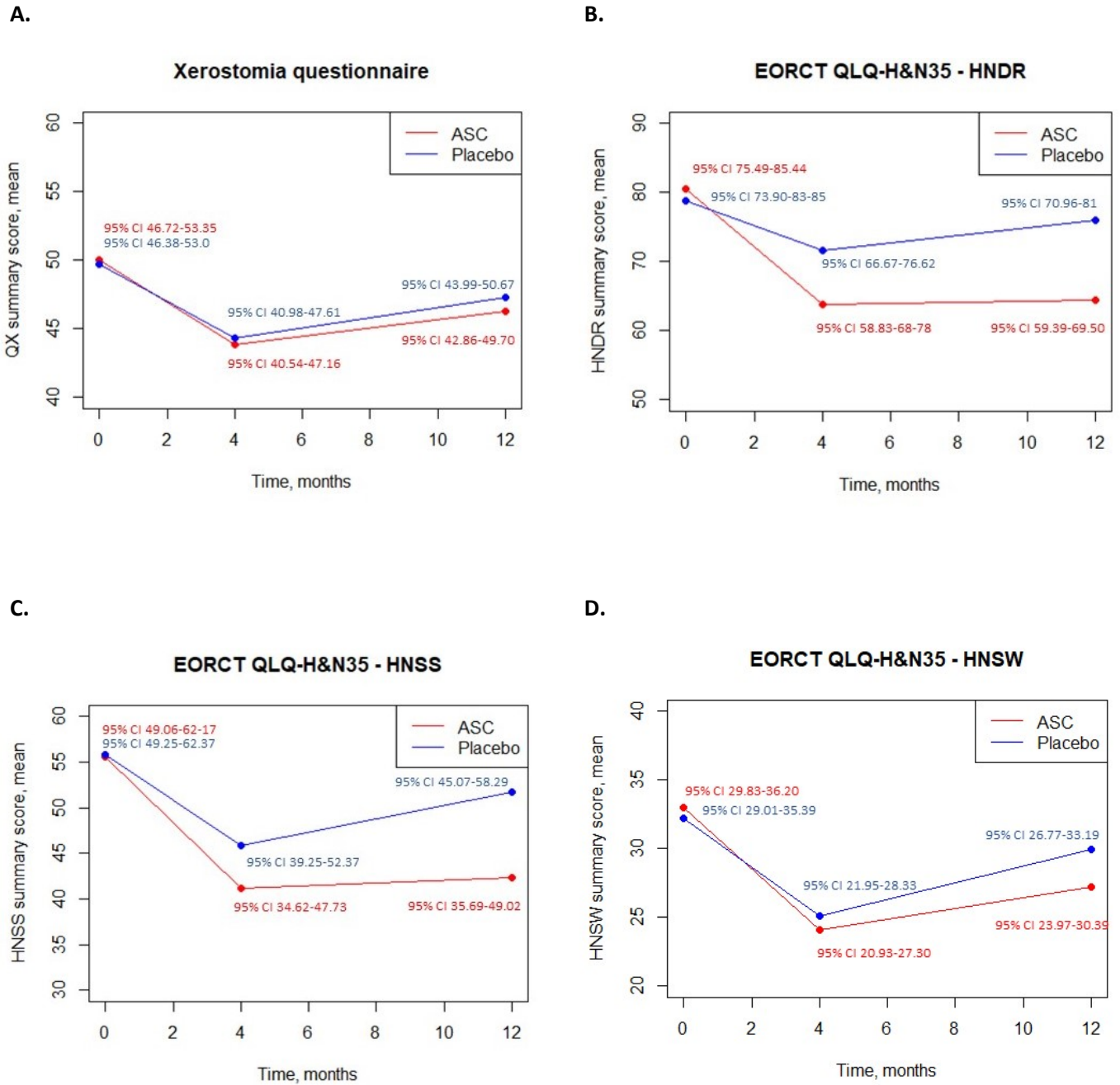
From baseline to 12 months, an increase in SWS was observed in the ASC group (0.04 mL/min; 95% CI [-0.01 to 0.1]) and in the placebo group (0.08 mL/min; 95% CI [0.02 to 0.13]), with no difference in change between the groups (-0.04 mL/min; 95% CI [-0.12 to 0.04]), see **Table 2**. The SWS increased from 1.07 mL/min (95% CI 0.99 to 1.14) to 1.12 mL/min (95% CI 1.04 to 1.19) in the ASC group, and from 1.06 mL/min (95% CI 0.99 to 1.13) to 1.18 mL/min (95% CI 1.11 to 1.25) in the placebo group, see **Figure 2**. Overall, no difference between the ASC and placebo group was observed in salivary gland function.

Three patients were lost to follow-up at the 12-month follow-up visit (one in the placebo group and two in the ASC group). No differences in mean salivary flow rates were observed in the non-responder analysis, see **Supplementary 2**.

Patient-reported outcome measurements

From baseline to 12 months, a decrease in EORCT-H&N35 HNDR sum score was observed in the ASC group (-10.07 units; 95% CI [-13.39 to -6.75]) and in the placebo group (-4.15 units; 95% CI [-7.46 to -0.84]), with a significant difference in change between the groups of -5.93 units (95% CI -10.62 to -1.22), see **Table 2**. The EORCT-H&N35 HNDR sum score decreased from 80.47 units (95% CI 75.49 to 85.44) to 64.45 units (95% CI 59.39 to 69.5) in the ASC group, and from 78.87 units (95% CI 73.90 to 83.85) to 75.97 units (95% CI 70.96 to 81) in the placebo group, see **Figure 3**. No difference in change between the ASC group and the placebo group was observed for EORCT-H&N35 domains HNSS (-4.72 units, [95% CI -10.78 to 1.35]) and HNSW (-1 unit [95% CI -3.94 to 1.94]), or XQ (-0.38 units [-3.68 to 2.91]).

Figure 3. Change in salivary gland function measured over the 12 months study period in the patient reported outcomes. **A.** Xerostomia questionnaire (XQ). **B.** EORCT-QLQ-H&N35 HNDR (domains for dry mouth). **C.** EORCT-QLQ-H&N35 HNSS (domains for sticky saliva) **D.** EORCT-QLQ-H&N35 HNSW (domains for swallowing).



Sum scores for both the XQ and the EORCT QLQ-H&N35 questionnaires range from 0-100. A Higher sum score is associated with increased symptom burden, while a lower sum score is associated with lower symptom burden.

Abbreviations: ASC, adipose-derived stem cells; CI, confidence interval; EORCT QLQ-H&N35, The European Organization for Research and Treatment of Cancer Quality of Life Questionnaire; Gy, gray; HNDR, domains for dry mouth; HNSS, domains for sticky saliva; HNSW, domains for swallowing; SMG, submandibular gland. XQ, xerostomia questionnaire.

Two patients (one in the ASC group and one in the placebo group) did not respond to all HNSW baseline questions, two patients did not respond to the XQ at 12 months (two in the ASC group) while three patients were lost to follow-up at 12 months (one the placebo group and two in the ASC group) and were consequently not included in the patient-reported outcomes analyses. No differences in mean scores were observed in the non-responder analysis, see **Supplementary 2**.

Safety and immune response

Through the 12 months study period no deaths were observed. In total, 16 SAEs were observed from baseline to the 12-month follow-up, and which were all deemed unrelated to the treatment. Three patients experienced a SAE from intervention to the 4-months follow-up (two in the placebo group and one in the ASC group), which are previously described²². From the 4-month to the 12-month follow-up additionally 13 SAEs occurred (six in the ASC group and seven in the placebo group). Four patients developed a new malignancy in the study period: two ASCs patients developed oesophageal cancer (one adenocarcinoma and one squamous cell carcinoma), one ASC patient developed both skin cancer and malignant melanoma, and one placebo patient developed chromophobe renal carcinoma. None of the SAEs were deemed related to the treatment and no difference was observed between ASCs and placebo, see **Table 3**. For further details see **Supplementary 3**.

Table 3. Safety profile up to 12 months from 4 months.

Adverse events	ASC group (n=60)	Placebo group (n=60)	Risk Difference (95% CI)
Deaths, no (%)	0 (0)	0 (0)	NA
Serious adverse events, no (%)	6 (10)	7 (12)	0.03 (-0.08-0.15)
Second primary cancer, no. (%)	4 (7)	1 (2)	
Infectious disease requiring hospitalization, no. (%)	0 (0)	3 (5)	
Other, no. (%)	2 (3)	3 (5)	
De novo donor specific HLA antibodies, no. (%)	23 (38)	NA	NA
Patients with persistent response from 4 to 12 m, no. (%)*	5 (8)		
Patients with increased response from 4 to 12 m, no. (%)*	1 (2)		
Patients with reduced response from 4 to 12 m, no. (%)*	8 (13)		
Patients with resolved response from 4 to 12 m, no. (%)**	8 (13)		
Unknown status at 12 months, no. (%)	1 (2)		

*An increase or decrease is defined as a change in mean fluorescence intensity (MFI) sum of donor specific antibodies \geq 3000. **Resolved response is defined as no detection of donor specific antibodies in a patient with a former de novo response.

In all, 23 (38%) patients receiving ASCs experienced an immune response to the treatment and developed de novo HLA class I donor specific antibodies. Of these, 8 (35%) patients experienced a resolved immune response at 12 months follow-up with no detectable donor specific antibodies remaining. Eight (35%) patients had a reduction in the immune response, 5 (22%) patients had a persistent immune response, and one patient (4%) had an increased immune response. One patient (4%) was lost to follow-up. See **Table 3**. For further details see **Supplementary 4**.

Subgroup analysis

Higher mean radiation dose to the four major salivary glands was associated with less increase in UWS with an average -0.004 mL/min (95% CI -0.01 to 0) per increase in Gy. Also, patients who developed donor specific antibodies (-0.01 mL/min [95% CI -0.07 to 0.05]), age over 60 (-0.04 mL/min, [95% CI -0.09 to 0.01]) and previous smokers (-0.03 mL/min [95% CI -0.07 to 0.02]) tended to experience a lower increase in UWS.

Discussion

This study represents the long-term results on effectiveness and safety of the MESRIX-III clinical trial: the largest human randomised trial investigating ASCs for radiation-induced salivary gland hypofunction and xerostomia in head and neck cancer survivors. At 12 months following intervention, we found an increase in UWS in both arms, with a 25% increase in the ASC group and a 27% increase in the placebo group, with no statistically significant difference between the two groups (difference in change -0.01 mL/min [95% CI -0.02 to 0.03]).

We found a significant decrease in the symptom burden related to dry mouth (EORTH-H&N35 HNDR) for ASC treated patients from 80.47 units (95% CI 75.49 to 85.44) to 64.45 units (95% CI 59.39 to 69.5) compared to a placebo (from 78.87 units; 95% CI [73.90 to 83.85] to 75.97 units; 95% CI [70.96 to 81]) favouring ASC therapy at 12 months. Significant long-term effect of ASCs compared to placebo on patients-reported outcomes was also observed in the MESRIX-I study²⁴. In addition, we found a decrease in the symptom burden for all other patient-reported outcomes including sticky saliva, swallowing, and xerostomia, with no significant differences in change between the groups (HNSS -4.72 units; 95% CI [-10.78 to 1.35], HNSW -1 unit; 95% CI [-3.94 to 1.94], XQ -0.38 units; 95% CI [-3.68 to 2.91]). However, the reduction in sum scores for all patient-reported outcomes was most pronounced in the ASC group and approached a potentially clinically significant change of 5-10 units, as indicated by previous studies^{29,30}.

The increase in both salivary gland function and patient-reported outcomes observed in the placebo group is noteworthy. Improvements in salivary gland function and xerostomia related symptoms is not expected more than two years after radiotherapy^{31,32}, although a continuous recovery have been observed up to four years post-RT³². These results indicate a continuous recovery following radiation-induced salivary gland damage. However, it could also indicate that the placebo Cryostor10 (BiolifeSolutions) containing 10% DMSO may act as a therapeutic agent in salivary gland damage, and DMSO may be anti-inflammatory as shown in other diseases³³⁻³⁵.

We did not observe any differences between the groups in long-term effect on SWS, with a difference in change of -0.04 mL/min (95% CI -0.17 to 0.04), corresponding to a 10% increase in the ASC group and a 11% increase in the placebo group. We did not treat the parotid glands, which are responsible for 50% of the stimulated saliva secretion³⁶, thus we did not expect changes in the SWS, and the effect was also lower than the effect on UWS. This supports the indication of a continuing natural salivary gland repair following RT.

No treatment-related SAEs following ASC therapy were observed, and we did not observe a risk difference between ASCs and placebo. This is in line with our previous MESRIX-studies^{20,21,24,25} and others²³. Four patients developed a new malignancy in the study period (three in the ASC group and one in the placebo group). There is no indication that MSC therapy leads to malignancy or undergo malignant transformation *in vivo*^{17,37,38}. Instead, previous cancer patients, including head and neck cancer patients, are at risk of developing a new secondary primary cancer^{39,40}. Also, patients were older (mean age 61.4 years) and with a history of smoking (56%) which also increases the risk of cancer.

In total, 38% of ASC patients developed HLA class I donor specific antibodies. Most of these either had a reduced or a resolved response at 12 months compared to 4 months, indicating a transient immune response, which is also described for patients receiving platelet transfusions⁴¹. Subgroup analysis revealed a tendency to a less increase in UWS in those who developed donor specific antibodies. Immunization with

HLA antibodies is known to cause challenges in solid organ transplantation and may eventually be important for a very small subset of patients receiving treatment with ASCs⁴².

In line with the results from the MESRIX-III at 4 months follow-up²², exploratory subgroup analyses also revealed that high mean radiation dose to the four major salivary glands was associated with a less increase in UWS (-0.004 mL/min, 95% CI [-0.01-0.00] per increase in Gy). Also, age over 60 years, and ever smokers trended to experience a lower effect of ASCs on UWS. Moving forward, further investigation is warranted to comprehensively elucidate the nuanced interplay between ASC therapy and patient-specific factors influencing treatment outcomes. Identifying potential subgroups might contribute to refining therapeutic strategies and optimizing patient care. This could also involve ASC treatment immediately following RT, when the salivary glands have a higher potential for regeneration, unlike the two-year post-treatment inclusion criteria which was used in this study.

While the observed increase in salivary flow rates suggests a degree of functional improvement, the absence of a significant difference between ASCs and placebo underscores the intricacies involved in restoring salivary gland function post-RT and prompt a deeper investigation into the mechanisms underlying salivary gland regeneration and the effect of ASC therapy. In addition, multiple administrations with MSCs might promote sustained beneficial effects compared to a single administration in both a variety of neurological diseases and in osteoarthritis⁴³⁻⁴⁷. Similarly, repeated treatment with ASCs may hold a potential to further improve saliva production in radiation-induced xerostomia, but has currently not been investigated.

This study was limited by the unblinded design at 12 months with both participants and study personnel being unblinded to the intervention, in contrast to the follow-up at 4 months which was blinded. This might overestimate the effect of ASCs while also underestimating the effect of placebo. Although this study is the largest human clinical trial investigating intraglandular ASCs for radiation-induced xerostomia, it remains insufficient for detailed exploration of potential subgroups, that may experience a more pronounced benefit from ASC therapy. Larger, multicentre studies are needed to further investigate if patient-specific variables impact treatment outcomes.

In conclusion, this study showed the first long-term results of a randomised, placebo-controlled phase II clinical trial investigating intraglandular ACS therapy for radiation-induced xerostomia and hyposalivation in head and neck cancer survivors. ASC therapy demonstrated a significant improvement in alleviating the subjective feeling of dry mouth compared to placebo at 12 months. While significant increases in unstimulated flow rates were observed for both ASCs and placebo, our findings did not establish the superiority of ACSs over placebo to restore salivary gland function as assessed by objective salivary flow rate measurements.

Author's contributions

ALFC: Conceptualization, resources, data curation, funding acquisition, formal analysis, validation, investigation, methodology, validation, writing–original draft, project administration. **KKJ:** Conceptualization, resources, data curation, funding acquisition, validation, investigation, methodology, writing–review and editing, project administration. **TT:** Supervision, validation, methodology, writing – review and editing. **NP:** Data curation, methodology, project administration, writing – review and editing. **AKØM:** Data curation, methodology, writing – review and editing. **SKB:** Data curation, writing – review and editing. **JK:** Conceptualization, supervision, validation, methodology, writing – review and editing. **AE:** Conceptualization, writing – review and editing. **MHS:** Conceptualization, writing – review and editing. **MF:** Data curation, writing – review and editing. **CM:** Data curation, writing – review and editing. **JF:** Data curation, supervision, writing – review and editing. **CDL:** Conceptualization, methodology, writing – review and editing. **AWH:** Data curation, writing – review and editing. **RC:** Formal analysis, supervision, writing – review and editing. **CG:** Conceptualization, resources, supervision, funding acquisition, validation, investigation, methodology, writing – review and editing. **CvB:** Conceptualization, resources, supervision, funding acquisition, validation, investigation, methodology, writing – review and editing.

Acknowledgements

We thank the patients who participated in this study. We thank the colleagues who provided assistance or instruction Mandana Haack-Sørensen, Annette Ekblond and Jacob Melchior.

References

1. Sung H, Ferlay J, Siegel RL, et al. Global Cancer Statistics 2020: GLOBOCAN Estimates of Incidence and Mortality Worldwide for 36 Cancers in 185 Countries. *CA Cancer J Clin.* 2021;71(3):209-249. doi:10.3322/caac.21660
2. Marur S, Forastiere AA. Head and Neck Squamous Cell Carcinoma: Update on Epidemiology, Diagnosis, and Treatment. *Mayo Clin Proc.* 2016;91(3):386-396. doi:10.1016/j.mayocp.2015.12.017
3. Borrás JM, Barton M, Grau C, et al. The impact of cancer incidence and stage on optimal utilization of radiotherapy: Methodology of a population based analysis by the ESTRO-HERO project. *Radiotherapy and Oncology.* 2015;116(1):45-50. doi:10.1016/j.radonc.2015.04.021
4. Vissink A, van Luijk P, Langendijk JA, Coppes RP. Current ideas to reduce or salvage radiation damage to salivary glands. *Oral Dis.* 2015;21(1):e1-10. doi:10.1111/odi.12222
5. Nutting CM, Morden JP, Harrington KJ, et al. Parotid-sparing intensity modulated versus conventional radiotherapy in head and neck cancer (PARSPORT): a phase 3 multicentre randomised controlled trial. *Lancet Oncol.* 2011;12(2):127-136. doi:10.1016/S1470-2045(10)70290-4
6. Jasmer KJ, Gilman KE, Muñoz Forti K, Weisman GA, Limesand KH. Radiation-Induced Salivary Gland Dysfunction: Mechanisms, Therapeutics and Future Directions. *J Clin Med.* 2020;9(12). doi:10.3390/jcm9124095
7. Cooper JS, Fu K, Marks J, Silverman S. Late effects of radiation therapy in the head and neck region. *Int J Radiat Oncol Biol Phys.* 1995;31(5):1141-1164. doi:10.1016/0360-3016(94)00421-G
8. Jensen SB, Vissink A, Limesand KH, Reyland ME. Salivary Gland Hypofunction and Xerostomia in Head and Neck Radiation Patients. *J Natl Cancer Inst Monogr.* 2019;2019(53). doi:10.1093/jncimonographs/lgz016
9. Schulz RE, Bonzanini LIL, Ortigara GB, et al. Prevalence of hyposalivation and associated factors in survivors of head and neck cancer treated with radiotherapy. *J Appl Oral Sci.* 2021;29:e20200854. doi:10.1590/1678-7757-2020-0854

10. Pinna R, Campus G, Cumbo E, Mura I, Milia E. Xerostomia induced by radiotherapy: an overview of the physiopathology, clinical evidence, and management of the oral damage. *Ther Clin Risk Manag.* 2015;11:171-188. doi:10.2147/TCRM.S70652
11. Vissink A, Jansma J, Spijkervet FKL, Burlage FR, Coppes RP. Oral sequelae of head and neck radiotherapy. *Crit Rev Oral Biol Med.* 2003;14(3):199-212.
12. Høxbroe Michaelsen S, Grønhøj C, Høxbroe Michaelsen J, Friborg J, von Buchwald C. Quality of life in survivors of oropharyngeal cancer: A systematic review and meta-analysis of 1366 patients. *Eur J Cancer.* 2017;78:91-102. doi:10.1016/j.ejca.2017.03.006
13. Riley P, Glenny AM, Hua F, Worthington H V. Pharmacological interventions for preventing dry mouth and salivary gland dysfunction following radiotherapy. *Cochrane Database Syst Rev.* 2017;7(7):CD012744. doi:10.1002/14651858.CD012744
14. Jaguar GC, Prado JD, Campanhã D, Alves FA. Clinical features and preventive therapies of radiation-induced xerostomia in head and neck cancer patient: a literature review. *Applied Cancer Research.* 2017;37(1):1-8. doi:10.1186/s41241-017-0037-5
15. Singer NG, Caplan AI. Mesenchymal Stem Cells: Mechanisms of Inflammation. *Annu Rev Pathol Mech Dis.* 2011;6:457-478. doi:10.1146/annurev-pathol-011110-130230
16. Krampera M, Le Blanc K. Mesenchymal stromal cells: Putative microenvironmental modulators become cell therapy. *Cell Stem Cell.* 2021;28(10):1708-1725. doi:10.1016/j.stem.2021.09.006
17. Von Bahr L, Batsis I, Moll G, et al. Analysis of tissues following mesenchymal stromal cell therapy in humans indicates limited long-term engraftment and no ectopic tissue formation. *Stem Cells.* 2012;30(7):1575-1578. doi:10.1002/stem.1118
18. Ankrum JA, Ong JF, Karp JM. Mesenchymal stem cells: Immune evasive, not immune privileged. *Nat Biotechnol.* 2014;32(3):252-260. doi:10.1038/nbt.2816
19. Carlander ALF, Gundestrup AK, Jansson PM, et al. Mesenchymal Stromal/Stem Cell Therapy Improves Salivary Flow Rate in Radiation-Induced Salivary Gland Hypofunction in Preclinical in vivo Models: A Systematic Review and Meta-Analysis. *Stem Cell Rev Rep.* 2024;20(4):1078-1092. doi:10.1007/s12015-024-10700-y

20. Grønhøj C, Jensen DH, Glovinski P V, et al. First-in-man mesenchymal stem cells for radiation-induced xerostomia (MESRIX): study protocol for a randomized controlled trial. *Trials*. 2017;18(1):108. doi:10.1186/s13063-017-1856-0
21. Lynggaard CD, Grønhøj C, Christensen R, et al. Intraglandular Off-the-Shelf Allogeneic Mesenchymal Stem Cell Treatment in Patients with Radiation-Induced Xerostomia: A Safety Study (MESRIX-II). *Stem Cells Transl Med*. 2022;11(5):478-489. doi:10.1093/stcltm/szac011
22. Jakobsen K, Carlander ALF, Todsén T, et al. Mesenchymal Stem/Stromal Cell Therapy for Radiation-Induced Xerostomia in Previous Head and Neck Cancer Patients: A Phase 2 Randomised, Placebo-Controlled Trial. *Clin Cancer Res*. Published online March 2024. doi:10.1158/1078-0432.CCR-23-3675
23. Blitzer GC, Glazer T, Burr A, et al. Marrow-Derived Autologous Stromal Cells for the Restoration of Salivary Hypofunction (MARSH): A pilot, first-in-human study of interferon gamma-stimulated marrow mesenchymal stromal cells for treatment of radiation-induced xerostomia. *Cytotherapy*. 2023;25(11):1139-1144. doi:10.1016/j.jcyt.2023.07.009
24. Lynggaard CD, Grønhøj C, Jensen SB, et al. Long-term Safety of Treatment with Autologous Mesenchymal Stem Cells in Patients with Radiation-Induced Xerostomia: Primary Results of the MESRIX Phase I/II Randomized Trial. *Clin Cancer Res*. 2022;28(13):2890-2897. doi:10.1158/1078-0432.CCR-21-4520
25. Kronberg Jakobsen K, Duch Lynggaard C, Paaske N, et al. Long-Term Outcome Following Treatment with Allogeneic Mesenchymal stem/Stromal Cells for Radiation-Induced Hyposalivation and Xerostomia. *Stem Cells Transl Med*. Published online 2023.
26. Jakobsen KK, Carlander ALF, Grønhøj C, et al. Effectiveness and safety of mesenchymal stem/stromal cell for radiation-induced hyposalivation and xerostomia in previous head and neck cancer patients (MESRIX-III): a study protocol for a single-centre, double-blinded, randomised, placebo-controlled, pha. *Trials*. 2023;24(1):1-9. doi:10.1186/s13063-023-07594-5
27. Haack-Sørensen M, Johansen EM, Højgaard LD, Kastrup J, Ekblond A. GMP Compliant Production of a Cryopreserved Adipose-Derived Stromal Cell Product for Feasible and Allogeneic Clinical Use. *Stem Cells Int*. 2022;2022:4664917. doi:10.1155/2022/4664917

28. Kastrup J, Haack-Sørensen M, Juhl M, et al. Cryopreserved Off-the-Shelf Allogeneic Adipose-Derived Stromal Cells for Therapy in Patients with Ischemic Heart Disease and Heart Failure—A Safety Study. *Stem Cells Transl Med.* 2017;6(11):1963-1971. doi:10.1002/sctm.17-0040
29. Musoro JZ, Coens C, Sprangers MAG, et al. Minimally important differences for interpreting EORTC QLQ-C30 change scores over time: A synthesis across 21 clinical trials involving nine different cancer types. *Eur J Cancer.* 2023;188:171-182. doi:10.1016/j.ejca.2023.04.027
30. Osoba D, Rodrigues G, Myles J, Zee B, Pater J. Interpreting the significance of changes in health-related quality-of-life scores. *J Clin Oncol.* 1998;16(1):139-144. doi:10.1200/JCO.1998.16.1.139
31. Eisbruch A, Kim HM, Terrell JE, Marsh LH, Dawson LA, Ship JA. Xerostomia and its predictors following parotid-sparing irradiation of head-and-neck cancer. *Int J Radiat Oncol Biol Phys.* 2001;50(3):695-704. doi:10.1016/s0360-3016(01)01512-7
32. Hiraoka S, Yoshimura M, Nakajima A, Nakashima R, Mizowaki T. Long-term outcomes of stimulated salivary flow and xerostomia after definitive intensity-modulated radiation therapy for patients with head and neck cancer†. *J Radiat Res.* 2024;65(1):71-77. doi:10.1093/jrr/rrad087
33. Karim M, Boikess RS, Schwartz RA, Cohen PJ. Dimethyl sulfoxide (DMSO): a solvent that may solve selected cutaneous clinical challenges. *Arch Dermatol Res.* 2023;315(6):1465-1472. doi:10.1007/s00403-022-02494-1
34. Huang SH, Wu CH, Chen SJ, Sytwu HK, Lin GJ. Immunomodulatory effects and potential clinical applications of dimethyl sulfoxide. *Immunobiology.* 2020;225(3):151906. doi:10.1016/j.imbio.2020.151906
35. Hoang C, Nguyen AK, Nguyen TQ, et al. Application of Dimethyl Sulfoxide as a Therapeutic Agent and Drug Vehicle for Eye Diseases. *J Ocul Pharmacol Ther.* 2021;37(8):441-451. doi:10.1089/jop.2021.0043
36. Mercadante V, Jensen SB, Smith DK, et al. Salivary Gland Hypofunction and/or Xerostomia Induced by Nonsurgical Cancer Therapies: ISOO/MASCC/ASCO Guideline. *J Clin Oncol.* 2021;39(25):2825-2843. doi:10.1200/JCO.21.01208

37. Thompson M, Mei SHJ, Wolfe D, et al. Cell therapy with intravascular administration of mesenchymal stromal cells continues to appear safe: An updated systematic review and meta-analysis. *EClinicalMedicine*. 2020;19:100249. doi:10.1016/j.eclinm.2019.100249
38. Lalu MM, McIntyre L, Pugliese C, et al. Safety of cell therapy with mesenchymal stromal cells (SafeCell): a systematic review and meta-analysis of clinical trials. *PLoS One*. 2012;7(10):e47559. doi:10.1371/journal.pone.0047559
39. Andersen L, Jakobsen KK, Carlander ALF, et al. The Incidence, Survival, and HPV Impact of Second Primary Cancer following Primary Oropharyngeal Squamous Cell Carcinoma: A 20-Year Retrospective and Population-Based Study. *Viruses*. 2022;15(1). doi:10.3390/v15010034
40. Baxi SS, Pinheiro LC, Patil SM, Pfister DG, Oeffinger KC, Elkin EB. Causes of death in long-term survivors of head and neck cancer. *Cancer*. 2014;120(10):1507-1513. doi:10.1002/cncr.28588
41. McGrath K, Wolf M, Bishop J, et al. Transient platelet and HLA antibody formation in multitransfused patients with malignancy. *Br J Haematol*. 1988;68(3):345-350. doi:10.1111/j.1365-2141.1988.tb04212.x
42. Ziemann M, Altermann W, Angert K, et al. Preformed Donor-Specific HLA Antibodies in Living and Deceased Donor Transplantation: A Multicenter Study. *Clin J Am Soc Nephrol*. 2019;14(7):1056-1066. doi:10.2215/CJN.13401118
43. Matas J, Orrego M, Amenabar D, et al. Umbilical Cord-Derived Mesenchymal Stromal Cells (MSCs) for Knee Osteoarthritis: Repeated MSC Dosing Is Superior to a Single MSC Dose and to Hyaluronic Acid in a Controlled Randomized Phase I/II Trial. *Stem Cells Transl Med*. 2019;8(3):215-224. doi:10.1002/sctm.18-0053
44. Petrou P, Kassis I, Ginzberg A, et al. Long-Term Clinical and Immunological Effects of Repeated Mesenchymal Stem Cell Injections in Patients With Progressive Forms of Multiple Sclerosis. *Front Neurol*. 2021;12:639315. doi:10.3389/fneur.2021.639315
45. Song Y, Du H, Dai C, et al. Human adipose-derived mesenchymal stem cells for osteoarthritis: a pilot study with long-term follow-up and repeated injections. *Regenerative Med*. 2018;13(3):295-307. doi:10.2217/rme-2017-0152

46. Pan K, Deng L, Chen P, et al. Safety and Feasibility of Repeated Intrathecal Allogeneic Bone Marrow-Derived Mesenchymal Stromal Cells in Patients with Neurological Diseases. *Stem Cells Int.* 2019;2019:8421281. doi:10.1155/2019/8421281

47. Hlebokazov F, Dakukina T, Potapnev M, et al. Clinical benefits of single vs repeated courses of mesenchymal stem cell therapy in epilepsy patients. *Clin Neurol Neurosurg.* 2021;207:106736. doi:10.1016/j.clineuro.2021.106736

Supplementary 1. Deviations to protocol from month 4 to month 12.

Deviation	Number of patients	Reason	Consequence
Follow-up time was too late (more than 12 months +/- 4 weeks)	1	Due to illness, work commitments, and others, one patient rescheduled their appointment.	Registered in the trial master file.
Received a smaller amount of paraffin wax during sialometry at 12 months measuring stimulated salivary flow rate	8 (4 ASC and 4 placebo)	Measuring error at baseline.	Registered in the trial master file. An equivalent amount was used for the subsequent follow-up visits.
Sialometry was not conducted simultaneously at the same time of day at 12 months as the previous assessments.	1	Due to work commitments and other, one patient rescheduled their appointment	Registered in the trial master file.

Supplementary 2. Sensitivity analysis. Non-responder imputation with baseline values carried forward.

	ASC (n = 60)	Placebo (n = 60)	Difference (95% CI)	p-value*
Change From Baseline to 12 months				
Unstimulated saliva flow rate, mL/min	0.02 (0.01)	0.02 (0.01)	0.01 (-0.02 to 0.03)	0.57
Stimulated saliva flow rate, mL/min	0.04 (0.03)	0.08 (0.03)	-0.04 (-0.11 to 0.04)	0.36
XQ-summary score (0-100)	-3.25 (1.15)	-2.63 (1.15)	-0.62 (-3.84 to 2.60)	0.70
<i>EORTC QLQ-H&N35 (score 0-100):</i>				
HNDR	-9.77 (1.67)	-4.12 (1.67)	-5.65 (-10.35 to -0.95)	0.02*
HNSS	-8.99 (2.18)	-3.98 (2.18)	-5.01 (-11.11 to 1.09)	0.11
HNSW	-4.82 (1.05)	-2.86 (1.05)	-1.96 (-4.91 to 0.99)	0.19

Abbreviation: EORTC QLQ-H&N35: the European Organization for Research and Treatment of Cancer Quality of Life Questionnaire, Head and Neck Module; HNDR: domains for dry mouth (HNDR); HNSS: domains for sticky saliva; HNSW: domains for swallowing; XQ: xerostomia Questionnaire; ASC: adipose-derived mesenchymal stem/stromal cells

Supplementary 3. Serious adverse events from month 4 to month 12.

In total, 13 patients had a serious adverse event (SAE) from the 4-month follow-up to the 12-month follow-up. Of these, seven patients received placebo and six patients received adipose-derived mesenchymal stromal cells.

Patients who received placebo experienced: One patient were diagnosed with chromofobe renal cell carcinoma (stage pT1a) and underwent partial nefrectomy. The patient was deemed curative treated; one patient had pneumonia and was hospitalized to receive i.v antibiotics for 4 days; one patient had pyelonephritis and were hospitalized to receive i.v antibiotics for one day; one patient had gallstones complicated with cholecystitis and was cholecyctomised; one patient was admitted for insertion of a canalicular monostent due to persistent tearing; one patient underwent gastroscopy to due epigastric pain and was diagnosed with inflammation; one patient were diagnosed with a laryngeal papilloma without suspicion of cancer which was surgically removed.

Patients who received ASCs experienced: One patient were diagnosed with esophageal cancer (stage T3N1M0) five months following the intervention and received palliative treatment; one patient had skin squamous cell carcinoma which was surgically removed and the patient was deemed curatively treated. The patient had a history of both skin squamous cell carcinoma and malignant melanoma. The same patients were also diagnosed with malignant melanoma at the 4-month follow-up as reported previsouly¹. One patient had complicated kidney stones and was hospitalized to endoscopic removal; one patient was hospitalized due to amaurosis fugax and no underlying diagnosis was established; one patient was diagnosed with esophageal cancer (stage cT4bN2M0) seven months following the intervention and received palliative treatment. None of the SAEs were deemed related to the treatment. The two patients who experienced esophageal cancer were lost to follow-up, since they received palliative treatment.

Supplementary 4. Immune response and development of HLA antibodies from baseline to 12 months in the ASC group.

Donor 1: HLA-A3,-; B18,37; Cw6,7; DR4,14; DR52, DR53; DQ5(1),8(3); DPB1*02:01,16:01

Donor 2: HLA-A2,11; B57,B62(15); Cw9(3),6; DR7,13(6); DR52,DR53N; DQ6(1),9(3); DPB1*04:01,-

Donor 3: HLA-A2,3;B7,-;Cw7,-;DR15(2),-;DR51;DQ6(1),-; DPB1*02:01,04:01.

Trial ID	Donor	Donor ID	Donor-specific HLA DSA (MFI)				Antibody types detected
			HLA Antigen	Baseline MFI	4 months MFI	12 months MFI	
002	1	IH23	A3	(-)	3000	(-)	De novo DSA and preformed DSA
			B18	4000	10000	6000	
			B37	2000	5000	3000	
			DQ5	4000	4000	5000	
003	1	IH23	-	(-)	(-)	(-)	
007	1	IH23	-	(-)	(-)	(-)	
008	1	IH23	B37	2000	1000	2000	Preformed DSA
010	1	IH23	-	(-)	(-)	(-)	
015	2	AA25	B57	(-)	2000	1000	De novo DSA
			B62	(-)	2000	(-)	
016	2	AA25	-	(-)	(-)	(-)	
018	2	AA25	A11	(-)	(-)	4000	
020	1	IH23	-	(-)	(-)	(-)	
021	2	AA25	A2	(-)	2000	1000	De novo DSA
			B57	(-)	6000	5000	
			B62(15)	(-)	1000	2000	
022	2	AA25	-	ND	(-)	(-)	
024	2	AA25	A2	(-)	7000	5000	De novo DSA
			B57	(-)	14.000	6000	
			B62(15)	(-)	2000	1000	
			Cw9(3)	(-)	1000	(-)	
026	2	AA25	B57	(-)	2000	(-)	De novo DSA
			B62(15)	(-)	1000	(-)	
028	1	IH23	-	(-)	(-)	(-)	
029	1	IH23	-	(-)	(-)	(-)	
033	1	IH23	-	(-)	(-)	(-)	
034	1	IH23	-	(-)	(-)	(-)	
035	1	IH23	-	(-)	(-)	(-)	
037	3	ML28	A2	(-)	16000	ND	De novo DSA
			A3	(-)	7000	ND	
			B7	(-)	15000	ND	
038	3	ML28	-	(-)	(-)	(-)	
040	3	ML28	B7	(-)	1000	2000	De novo DSA
041	3	ML28	-	(-)	(-)	(-)	
043	3	ML28	A3	(-)	2000	(-)	De novo DSA
			DQ6	3000	3000	3000	
047	3	ML28	(-)	(-)	(-)	(-)	
049	3	ML28	A3	(-)	10000	9000	De novo DSA and preformed DSA
			B7	3000	22000	23000	
			DR15	10000	10000	9000	
			DR51	11000	11000	12000	
			DQ6(1)	6000	6000	6000	
052	3	ML28	-	(-)	(-)	(-)	
053	3	ML28	-	(-)	(-)	(-)	
057	3	ML28	A2	(-)	<2000	(-)	De novo DSA
059	3	ML28	A3	(-)	12000	9000	De novo DSA
			B7	(-)	8000	5000	
			DQ6(1)	7000	7000	7000	

060	2	AA25	-	(-)	(-)	(-)	
061	3	ML28	-	(-)	(-)	(-)	
064	3	ML28	-	(-)	(-)	(-)	
066	2	AA25	B62(15) Cw9	2000 2000	12000 9000	9000 8000	Preformed DSA
069	3	ML28	-	(-)	(-)	(-)	
071	3	ML28	-	(-)	(-)	(-)	
072	3	ML28	-	(-)	(-)	(-)	
073	2	AA25	A2 B57 B62(15) DQ6(1)	(-) (-) (-) 1000	2000 5000 1000 1000	(-) (-) (-) (-)	De novo DSA
074	2	AA25	-	(-)	(-)	(-)	
078	3	ML28	-	(-)	(-)	(-)	
080	2	AA25	A2 A11 B57 B62(15)	(-) (-) (-) (-)	7000 2000 14000 5000	2000 (-) 3000 3000	De novo DSA
081	3	ML28	-	(-)	(-)	(-)	
084	3	ML28	A3 B7	(-) (-)	1000 2000	(-) (-)	De novo DSA
086	2	AA25	-	(-)	(-)	ND	
087	3	ML28	A2	(-)	2000	(-)	De novo DSA
089	1	IH23	-	(-)	(-)	(-)	
090	3	ML28	A2 A3	(-) (-)	8000 6000	1000 (-)	De novo DSA
093	3	ML28	-	(-)	(-)	(-)	
094	1	IH23	-	(-)	(-)	(-)	
098	1	IH23	-	(-)	(-)	(-)	
099	3	ML28	-	(-)	(-)	(-)	
103	3	ML28	-	(-)	(-)	(-)	
104	3	ML28	-	(-)	(-)	(-)	
105	3	ML28	A2 A3 B7	(-) (-) (-)	6000 2000 4000	(-) (-) 2000	De novo DSA
108	1	IH23	-	(-)	(-)	(-)	
109	1	IH23	-	(-)	(-)	(-)	
110	2	AA25	A2 B57	(-) (-)	2000 2000	(-) (-)	De novo DSA
114	3	ML28	-	(-)	(-)	(-)	
117	1	IH23	Cw7	1000	(-)	(-)	Preformed DSA
119	3	ML28	A2 A3 B7	(-) (-) (-)	5000 7000 5000	(-) 1000 (-)	De novo DSA
120	1	IH23	-	(-)	(-)	(-)	

Abbreviations: DSA: Donor specific antibodies; HLA: human leucocyte antigen; ID: identification; MFI: normalized mean fluorescence intensity; ND = not determined; (-): negative. MFI is approximate values (rounded to the nearest thousand). Negative is defined as MFI < 1000 for Labscreen Single Antigen, One Lambda, NBG<3.0 for HLA class I and NBG<4.0 for HLA class II for Labscreen Mixed, One Lambda

Supplementary 5. Statistical Analysis Plan

Statistical Analysis Plan

Title

Long-term Effectiveness and Safety of Mesenchymal Stromal Cell Therapy for Alleviating Radiation-Induced Hyposalivation and Xerostomia in Head and Neck Cancer Survivors: Statistical Analysis Plan for the secondary analyses of the Single-center, Double-blind, Randomized, Placebo-Controlled MESRIX-III Trial

Trial Registration

Danish Data Protection Agency (protocol number P-2020-1164)

The National Ethics Committee protocol number: (Protocol number: 1802872)

The Danish Medical Agency (2018-000348-24)

ClinicalTrials.gov database (NCT04776538)

SAP version:

Version 1, 3st of March 2024

SAP associated with protocol version 2.10

Primary authors of the SAP: Amanda-Louise Fenger Carlander, Kathrine Kronberg Jakobsen and Robin Christensen

Roles and responsibilities

Senior biostatistician responsible: Robin Christensen, BSc, MSc, PhD, Professor

Principal investigator: Kathrine Kronberg Jakobsen, MD, PhD-fellow

Investigator: Amanda-Louise Fenger Carlander, MD, PhD-fellow

Sponsor: Christian von Buchwald, MD, DMSc, Professor

This SAP is reported following the recommendations from “Guidelines for the Content of Statistical Analysis Plans in Clinical Trials.” by Gamble C, Krishan A, Stocken D, Lewis S, Juszcak E, Doré C, et al. published in JAMA 2017;318:2337-43.

Indholdsfortegnelse

Introduction.....	32
Objectives.....	32
Safety objectives.....	32
Study methods.....	33
Randomization.....	33
Blinding.....	33
Sample size and power considerations	33
Framework.....	34
Statistical interim analyses and stopping guidance.....	34
Timing of final analysis	34
Timing of outcome assessments	35
Table 1.	35
Statistical principles	36
Adherence and protocol deviations	36
Analysis populations.....	36
Trial population.....	36
Baseline patient characteristics.....	36
Analysis.....	37
Analysis methods.....	37
Sensitivity.....	38
Harms.....	38
Statistical software	39
MANUSCRIPT OUTLINE.....	40
Figure 1.	40
Figure 2.	41
Table 2.	42
Table 3.	44
Supplementary Figure 1.	Fejl! Bogmærke er ikke defineret.

Introduction

Background and rationale

A prevalent adverse outcome of radiotherapy in head and neck cancer cases is the diminished function of salivary glands leading to xerostomia. Intraglandular therapy involving mesenchymal stem cells has exhibited promising outcomes in addressing xerostomia¹⁻⁴. This trial aims to assess the long-term effectiveness and safety of administering adipose-derived mesenchymal stem cells (ASC) through submandibular gland injection as a potential novel, disease-modifying intervention for post-radiation xerostomia. Anticipated results could establish a foundation for a clinically feasible approach to alleviate xerostomia in head and neck cancer survivors previously treated with radiotherapy.

Objectives

Primary objective

To compare the effect of ASC injection, relative to placebo, on changes in unstimulated salivary gland function from baseline to month 12:

i.e., effectiveness: change in salivary gland function is measured by 12 months change in unstimulated whole saliva flow rate between ASC and placebo.

Secondary objectives

To compare the effect of ASC injection, relative to placebo, from baseline to month 12 on the following outcomes:

1. Effectiveness: Changes in stimulated salivary gland function measured by 12 months change in stimulated whole saliva flow rate.
2. Patient-reported outcome: The European organization for research and treatment of cancer quality of life questionnaire, head and neck-35 (EORTC QLQ-H& N35):
 - Change in domains for dry mouth (HNDR)
 - Change in domains for sticky saliva (HNSS)
 - Change in domains for swallowing (HNSW)
3. Patient-reported outcome: Xerostomia Questionnaire (XQ).

Safety objectives

1. Safety up to 12 months duration from baseline will be evaluated by the incidence of:

- a) Serious adverse events
- b) Deaths

2. Immunological response to treatment: up to 12 months duration from baseline, evaluated by the

- a) Development of de novo HLA antibodies as a response to ASC treatment in the first 12 months of the study period.
- Patients with persistent antibodies from 4 months to 12 months
 - Patients with increased antibodies from 4 months to 12 months
 - Patients with reduced antibodies from 4 months to 12 months
 - Patients with resolved antibodies from 4 months to 12 months

Study methods

Trial design

The study was a randomized, single-center, double-blinded, placebo-controlled trial to compare the safety, tolerability, and effectiveness of intraglandular allogeneic ASCs as a treatment for radiation-induced hyposalivation and xerostomia in previous head and neck cancer patients. We intended to treat 120 patients with xerostomia, randomized in a 1:1 ratio, to receive ultrasound-guided injections of either ASCs or placebo in the submandibular glands. Placebo consisted of CryoStor10 (BiolifeSolutions), the freeze media for ASCs containing 10% Dimethyl sulfoxide (DMSO). Patients were followed up after four month (+/- 14 days) and after 12 months (+/- 4 weeks). The trial design is previously described⁵.

Randomization

A predefined randomization code was established for all 120 patients according to patient treatment order (1-120) from the start of the trial. The allocation sequence was generated using www.randomization.com. The table with randomization numbers was available to one specified person at the Cardiology Stem Cell Centre (CSCC), The Heart Centre, Rigshospitalet, who was not involved in analyzing data related to the study endpoints. Randomization was performed in blocks of six, three patients receiving ASCs and three patients receiving placebo. Within blocks of six a balanced use of donors was applied. For the purpose of safety and potential unblinding, two sets of sealed envelopes containing the randomization code for each patient were made. Envelopes were locked up at CSCC production facility until allocation of treatment. After treatment one envelope remained at the CSCC and one envelope was kept at the clinical site at sponsor and thereby available for the investigator. The randomization is previously described⁵.

Blinding

The sponsor, investigators, study staff (except for staff involved in stem cell preparation and staff involved in bioanalytical analyses), and patients were blinded to treatment assignment. A project nurse thawed the frozen suspension of ASCs or placebo before injection, and the syringes were covered in sterile green tape to ensure that neither the patients nor the study staff could see the suspension injected. After a follow-up period of 4 months, both patients and study staff were informed about the treatment, resulting in an unblinded 12-month follow-up phase. The blinding is previously described⁵.

Sample size and power considerations

From our previous study MESRIX-I we assessed the increase in saliva production for whole unstimulated saliva flow rate (in ml/min) by about 33% or in absolute numbers from 0.125 to 0.155 after 4 months. The

power calculation was derived from a power of 0.8 and an alpha of 0.05. Consequently, the required total number of patients for inclusion would be 100, with 50 allocated to each group. We expected a dropout rate of up to 20% and therefore aimed to include 120 participants in total (i.e., representing the intention-to-treat population). The sample size and power consideration are previously described⁵.

Framework

The trial was designed as a superiority trial, with the primary objective of showing that the response to the investigational product was superior to the comparative agent (experimental intervention and placebo comparator, respectively) as previously described⁵.

Statistical interim analyses and stopping guidance

Four months data has been published⁴.

Timing of final analysis

The primary endpoint, as well as the key secondary outcomes will all be evaluated based on the 12 months assessment.

Timing of outcome assessments

Table 1. Schedule of enrolment, interventions, and assessments.

	Study period			
	Enrolment	Allocation	Post-allocation	Close-out
Timepoint	Up to 90 days prior to intervention	0	4 months after intervention	12 months after intervention
ENROLMENT:				
<i>Eligibility screen</i>	X			
<i>Informed consent</i>	X			
<i>Allocation</i>		X		
INTERVENTION:				
<i>ASC</i>		X		
<i>Placebo</i>		X		
ASSESSMENTS:				
<i>Saliva flow rate</i>	X		X*	X
<i>Saliva quality</i>	X		X**	
<i>QoL questionnaires</i>	X		X*	X
<i>HLA-response</i>	X		X*	X
<i>Safety</i>			X*	X
<i>Ultrasound of the submandibular glands</i>	X		X*	X

*Reported in primary paper⁴.

**Reported in separate paper.

Statistical principles

Confidence intervals and P values

All 95% confidence intervals and P values will be two sided. We will not apply explicit adjustments for multiplicity, rather we will analyze the key secondary outcomes in a prioritized order (e.g., “gatekeeping procedure”): The analyses of the key secondary outcomes will be performed and interpreted in sequence until one of the analyses potentially fails to show the statistically significant difference, or until all analyses have been completed at a statistical significance level of 0.05. The key secondary statistical tests will be reported with P values for hypothesis tests and claims of statistical significance.

Adherence and protocol deviations

Adherence is defined as participants who has a full registration and has completed every assessment for all the time points in the study. The adherence to the assessments will be summarized with number and percent compliance. All deviations from the protocol will be described.

Analysis populations

The primary analyses will be based on the Intention to Treat (ITT) population, i.e., based on the Full Analysis Set. The ITT principle asserts the effect of a treatment policy (that is, the planned treatment regimen), rather than the actual treatment given (i.e., it is independent of treatment adherence). Accordingly, participants allocated to a treatment group (X_{ASC} and $X_{Placebo}$, respectively) will be followed up, assessed, and analyzed as members of that group, irrespective of their adherence to the planned course of treatment (i.e., independent of withdrawals and cross-over phenomenon).

We will use a (multilevel) repeated measures mixed effects model with participants as a random effects factor and the particular outcome variable (Y_i) as a dependent variable. The time (months; 3 levels: 0, 4, 12) is set as a fixed effect factor based on a restricted maximum likelihood model. While also adjusting for the level at baseline, the statistical model holds all between-time comparisons for assessment points up to 12 months from baseline (day 0), and allows for evaluation of the changes, as well as the trajectory over time from baseline to 12 months follow-up.

Trial population

Screening data, eligibility, and recruitment

Information on screening, eligibility, recruitment and withdrawal was published in the primary MESRIX-III paper⁴.

Baseline patient characteristics

Descriptive statistics for categorical data will be evaluated as numbers and percentages while continuous data will be summarized by mean and standard deviation. We will not perform tests of statistical

significance for baseline characteristics. Baseline patient characteristics was published in the primary MESRIX-III paper⁴.

Analysis

Outcome definitions

1. Effectiveness: Saliva gland function measured as the change in unstimulated whole saliva flow rate in the group receiving ASCs compared with the group of participants receiving placebo. Timeframe: 12 months. The saliva flow rate will be measured as ml/min.
2. Effectiveness: Saliva gland function measured as change in stimulated saliva flow rate in the group receiving ASCs compared with the group of participants receiving placebo. Timeframe: 12 months. The saliva flow rate will be measured as ml/min.
3. Effectiveness: Impact on quality of life measured as a change in patient-reported outcome of quality of life and xerostomia. Timeframe: 12 months. Patients will fill out the EORTC QLQ-H&N35.
 - a. HNDR
 - b. HNSS
 - c. HNSW
4. Effectiveness: Impact on quality of life measured as a change in patient-reported outcome of quality of life and xerostomia. Patients will fill out the XQ. The results will be reported as a collected score. Timeframe: 12 months
5. Safety: evaluated by the number of patients with serious adverse events. Timeframe: 12 months.
 - a. Development of serious adverse events
 - b. Deaths
6. Immune response to treatment with ASC: Development of HLA antibodies as a response to ASC from 4 months to 12 months (de novo donor specific HLA antibodies) measured as sum mean fluorescence intensity (MFI) sum.
 - a. Patients with persistent MFI sum from 4 months to 12 months
 - b. Patients with increased MFI sum ≥ 3000 from 4 months to 12 months
 - c. Patients with reduced MFI sum ≥ 3000 from 4 months to 12 months
 - d. Patients with resolved MFI sum from 4 months to 12 months

Analysis methods

The repeated measures designs aim to draw conclusions about the mean values of the populations from which participants are selected by considering treatment and time effects in the model. The objective is usually achieved by considering both treatment and time effects (as well as the interaction between them) in the model. Data will be analyzed using SAS or Rstudio, with the particular outcome variable (Y_i) as the dependent variable, using a multilevel repeated measures random effects model with participants as the random effect factor, and time (with 3 levels, incl. baseline) as fixed effect factors based on a restricted maximum likelihood (REML) model. This statistical model will hold all between-time comparisons for all assessment points up 12 months from baseline (incl. baseline) and thus allows for evaluation of the average effect, as well as the trajectory over time from baseline to 12 months follow-up.

Sensitivity

Exploring the robustness of the main analyses is a concept that refers to the sensitivity of the overall conclusions to various limitations of the data, assumptions, and analytic approaches to data analysis. Robustness implies that the treatment effect and primary conclusions of the trial are not substantially affected when analyses are carried out based on alternative assumptions or analytic approaches.

Loss to follow-up and missing data for various reasons is difficult to avoid in randomized trials and in pragmatic trials. We will apply the analysis framework suggested by White et al (2011) in which missing data related to the ITT approach depend on making plausible assumptions about the missingness of the data and including all participants in subsequent sensitivity analyses:

1. Attempt to follow up all randomized participants, even if they withdraw from allocated treatment (i.e., contact all individuals unless they explicitly stated that they had withdrawn their consent)
2. Perform a main analysis of all observed data that are valid under a plausible assumption about the missingness of the data (i.e., using Multiple imputation, assuming that data are 'Missing at Random' [MAR])
3. Perform sensitivity analyses to explore the effect of departures from the assumption made in the main (#2) analysis (i.e., a non-responder-imputation: using the value at baseline to replace missing data will correspond to a non-responder imputation; these models will potentially be informative even if data are 'Missing Not At Random' [MNAR])
4. Account for all randomized participants, at least in the sensitivity analyses (covered by #2 and #3 above, plus the corresponding analyses based on the per protocol population).

The interpretation of the corresponding statistical measures of uncertainty of the treatment effect and treatment comparisons will involve consideration of the potential contribution of bias to the P-value, 95% confidence interval, and of the inference in general.

Ad#1+2: Our primary analysis population will be all participants with available data at baseline, statistically modelled using repeated-measures linear mixed models (see above). These models will be valid if data are 'MAR'.

Ad#3+4 Sensitivity: We will analyze all variables, with missing data being handled by a simplistic non-responder imputation techniques (i.e., baseline observation carried forward).

When the different sensitivity analyses agree, and the analyses on the sensitivity analyses and the main analysis leads to essentially the same conclusions, confidence in the trial results is increased.

Harms

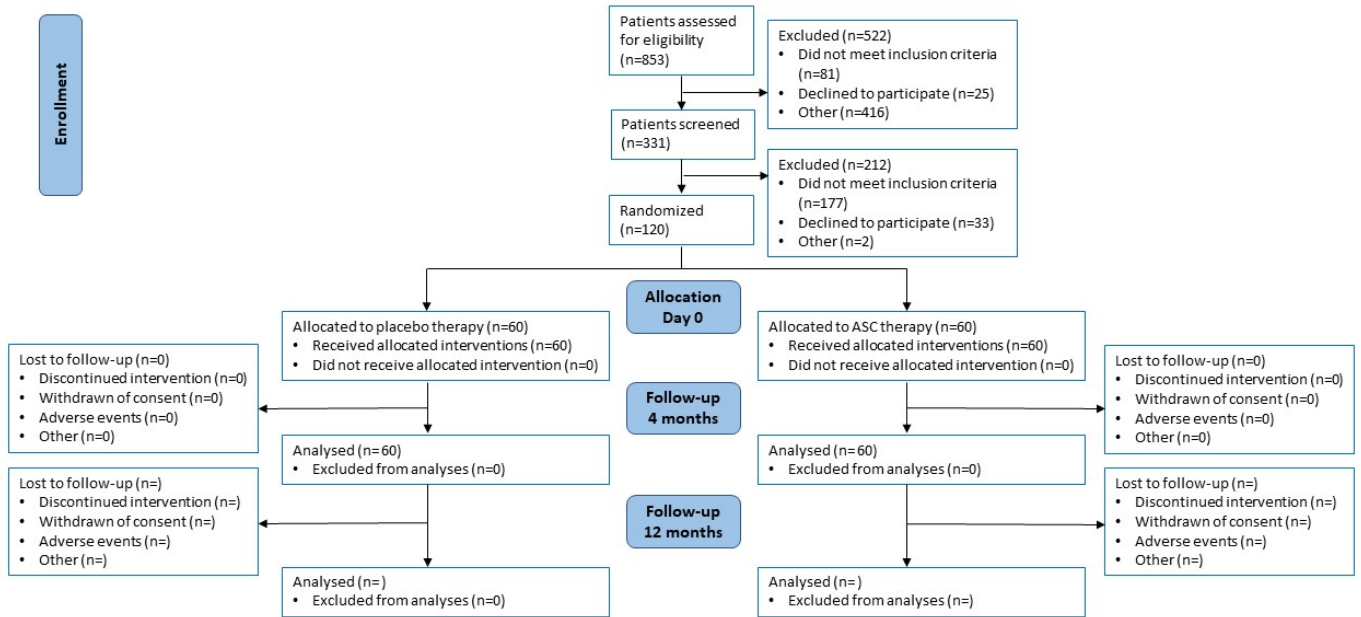
Any serious adverse events during the trial will be noted and reported in Table 3.

Statistical software

All statistical analyses will be performed in SAS and R-studio.

MANUSCRIPT OUTLINE *(Mock Ups)*

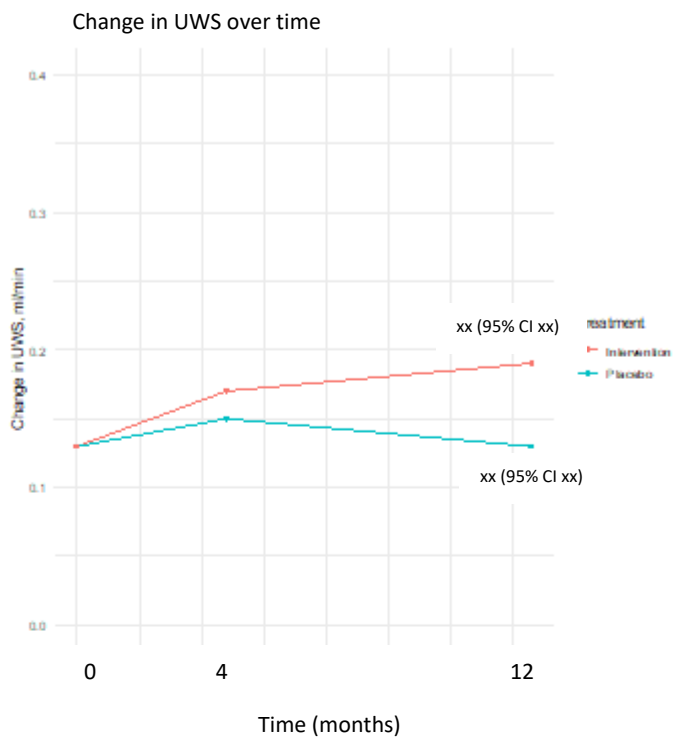
Figure 1. CONSORT Flow diagram illustrating the trial flow Patient Flow Throughout the 12 months follow-up in the MESRIX-III Randomized Clinical Trial



Abbreviation: ENT: Department of otorhinolaryngology, head and neck surgery

Figure 2. Simulated data: Change in salivary gland function measured over the 12 months period in **A.** Unstimulated whole saliva flow rate (UWS) **B.** Stimulated whole saliva flow rate (SWS).

A.



B.

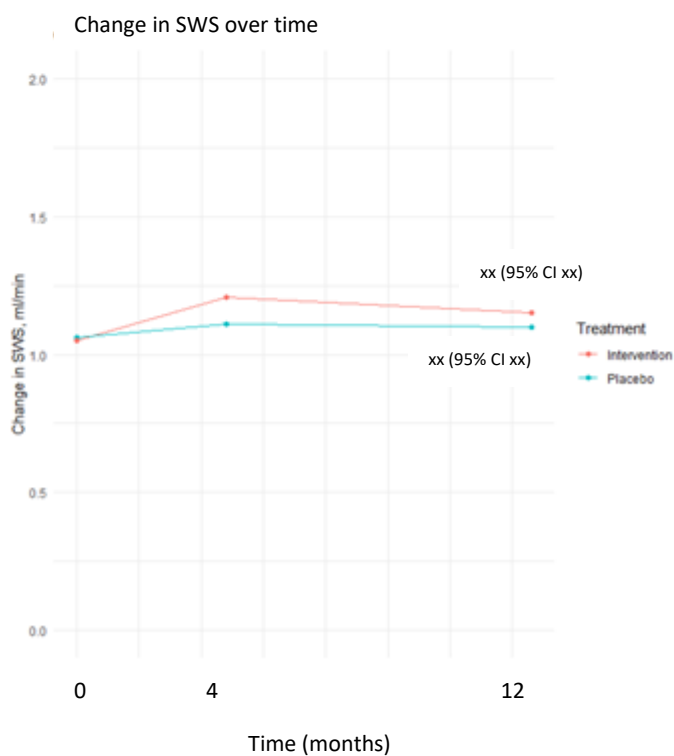


Figure 3. Simulated data: Change in salivary gland function measured over the 12 months period in the patient reported outcomes. **A.** XQ questionnaire. **B.** HNDR: domains for dry mouth evaluated by EORTC QLQ-H&N35; **C.** HNSS: domains for sticky saliva evaluated by EORTC QLQ-H&N35. HNSW: domains for swallowing evaluated by EORTC QLQ-H&N35

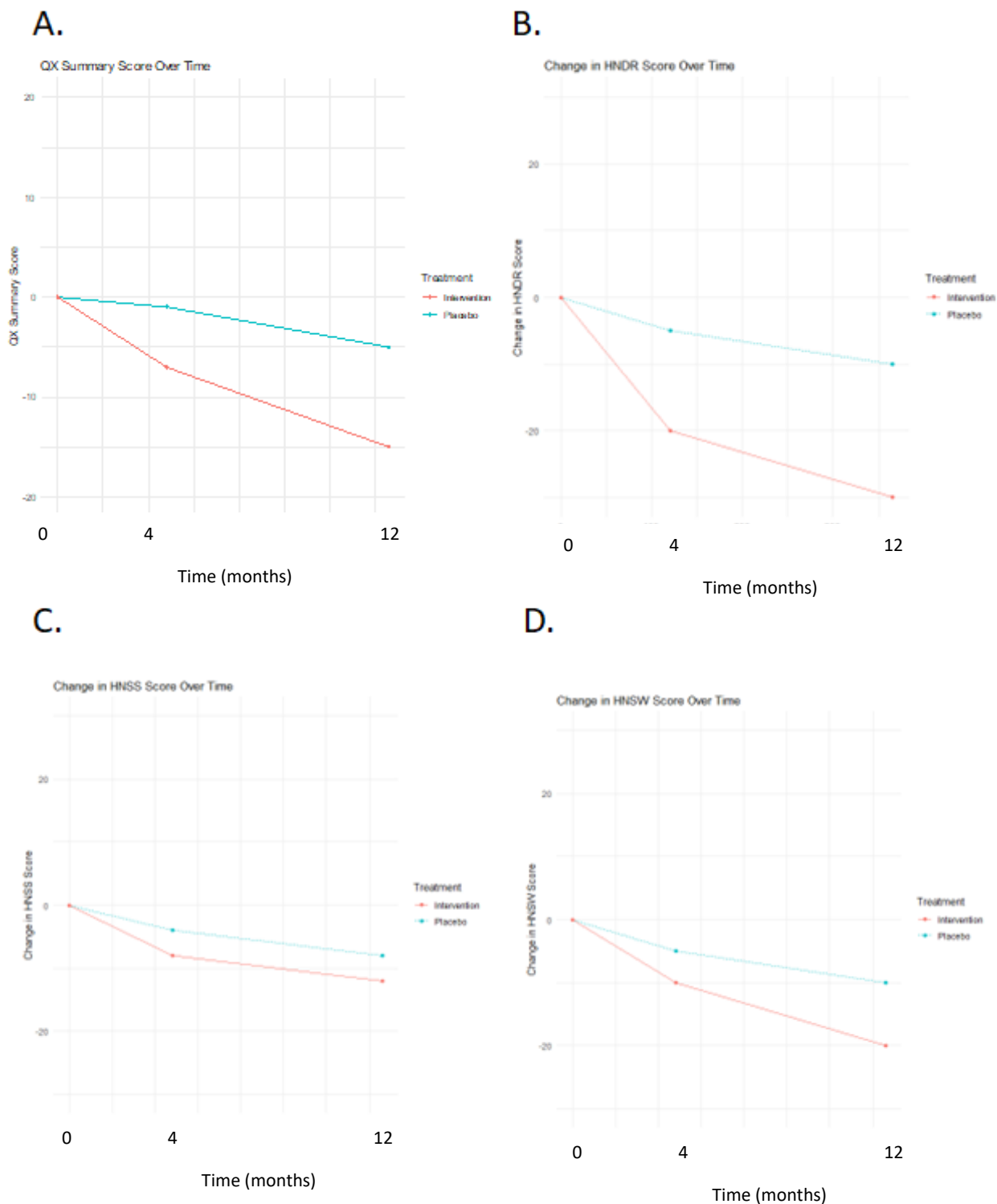


Table 2. Prespecified systematic harms; Safety profile up to 12 months from baseline.

Adverse events	ASC group (n=60)	Placebo group (n=60)	Risk Difference (95%CI)
Deaths, no (%)			
Serious adverse events, no (%)			
De novo donor specific antibodies, no. (%) Patients with persistent response from 4 to 12 m, no. (%)* Patients with increased response from 4 to 12 m, no. (%)* Patients with reduced response from 4 to 12 m, no. (%)* Patients with resolved response from 4 to 12 m, no. (%)*		NA	NA

*An increase or decrease is defined as a change in MFI sum of DSA \geq 3000.

Table 3. Comparison of change from in secondary outcomes from baseline to 12 months follow-up (Trial secondary end point)

	ASC (n = 60)	Placebo (n = 60)	Difference (95% CI)	P-value
Change From Baseline to 12 months				
Unstimulated saliva flow rate, ml/min				
Stimulated saliva flow rate, ml/min				
XQ-summary score (0-100)				
<i>EORCT QLQ-H&N35 (score 0-100):</i>				
HNDR				
HNSS				
HNSW				

Values will be least squares means with standard errors for each group; the difference between these will be reported with 95% confidence intervals (95% CIs). Estimates will be derived from the repeated measures, mixed effects models.

Supplementary Table 1. Immune response and development of HLA antibodies from baseline to 12 months in the ASC group.

Donor 1: HLA-A3,-; B18,37; Cw6,7; DR4,14; DR52, DR53; DQ5(1),8(3); DPB1*02:01,16:01

Donor 2: HLA-A2,11; B57,B62(15); Cw9(3),6; DR7,13(6); DR52,DR53N; DQ6(1),9(3); DPB1*04:01,-

Donor 3: HLA-A2,3;B7,-;Cw7,-;DR15(2),-;DR51;DQ6(1),-; DPB1*02:01,04:01

Pt ID no	Donor / Placebo	Donor-specific HLA antibodies (MFI)				Notes 4 months follow-up	Notes – 12 months follow-up
		HLA Antigen	Baseline MFI	4 months MFI	12 months MFI		
002	1	A3	(-)	3000		De novo antibodies and preformed antibodies	
		B18	4000	10000			
		B37	2000	5000			
		DQ5	4000	4000			
003	1	-	(-)	(-)			
007	1	-	(-)	(-)			
008	1	B37	2000	1000		Preformed antibodies	
010	1	-	(-)	(-)			
015	2	B57	(-)	2000		De novo antibodies	
		B62	(-)	2000			
016	2	-	(-)	(-)			
018	2	A11	(-)	(-)			
020	1	-	(-)	(-)			
021	2	A2	(-)	2000		De novo antibodies	
		B57	(-)	6000			
		B62(15)	(-)	1000			
022	2	-	ND	(-)			
024	2	A2	(-)	7000		De novo antibodies	
		B57	(-)	14.000			
		B62(15)	(-)	2000			
		Cw9(3)	(-)	1000			
026	2	B57	(-)	2000		De novo antibodies	
		B62(15)	(-)	1000			
028	1	-	(-)	(-)			
029	1	-	(-)	(-)			
033	1	-	(-)	(-)			
034	1	-	(-)	(-)			
035	1	-	(-)	(-)			
037	3	A2	(-)	16000		De novo antibodies	
		A3	(-)	7000			
		B7	(-)	15000			
038	3	-	(-)	(-)			
040	3	B7	(-)	1000		De novo antibodies	
041	3	-	(-)	(-)			
043	3	A3	(-)	2000		De novo antibodies	
		DQ6	3000	3000			
047	3	(-)	(-)	(-)			
049	3	A3	(-)	10000		De novo antibodies and preformed antibodies	
		B7	3000	22000			
		DR15	10000	10000			
		DR51	11000	11000			
		DQ6(1)	6000	6000			
053	3	-	(-)	(-)			
054	3	-	(-)	(-)			
059	3	A2	(-)	<2000		De novo antibodies	
061	3	A3	(-)	12000		De novo antibodies	
		B7	(-)	8000			
		DQ6(1)	7000	7000			
062	2	-	(-)	(-)			

063	3	-	(-)	(-)		
066	3	-	(-)	(-)		
068	2	B62(15) Cw9	2000 2000	12000 9000		Preformed antibodies
071	3	-	(-)	(-)		
073	3	-	(-)	(-)		
074	3	-	(-)	(-)		
075	2	A2 B57 B62(15) DQ6(1)	(-) (-) (-) 1000	2000 5000 1000 1000		De novo antibodies
076	2		(-)	(-)		
080	3	-	(-)	(-)		
082	2	A2 A11 B57 B62(15)	(-) (-) (-) (-)	7000 2000 14000 5000		De novo antibodies
083	3	-	(-)	(-)		
086	3	A3 B7	(-) (-)	1000 2000		De novo antibodies
088	2	-	(-)	(-)		
089	3	A2	(-)	2000		De novo antibodies
091	1	-	(-)	(-)		
092	3	A2 A3	(-) (-)	8000 6000		De novo antibodies
095	3	-	(-)	(-)		
096	1	-	(-)	(-)		
100	1	-	(-)	(-)		
101	3	-	(-)	(-)		
105	3	-	(-)	(-)		
106	3	-	(-)	(-)		
107	3	A2 A3 B7	(-) (-) (-)	6000 2000 4000		De novo antibodies
111	1	-	(-)	(-)		
112	1	-	(-)	(-)		
113	2	A2 B57	(-) (-)	2000 2000		De novo antibodies
117	3	-	(-)	(-)		
120	1	Cw7	1000	(-)		Preformed antibodies
122	3	A2 A3 B7	(-) (-) (-)	5000 7000 5000		De novo antibodies
123	1	-	(-)	(-)		

*Missing data.

Abbreviations: MFI: normalized mean fluorescence intensity; MFI is approximate values (rounded to the nearest thousand); (-): negative (MFI < 1000 for Labscreen Single Antigen, One Lambda, NBG<3.0 for HLA class I and NBG<4.0 for HLA class II for Labscreen Mixed, One Lambda)

Supplementary Table 2. Sensitivity analysis, missing data replaced using a simplistic non-responder imputation. Comparison of change from in secondary outcomes from baseline to 12 months follow-up (Trial secondary end point)

	ASC (n = 60)	Placebo (n = 60)	Difference (95% CI)	P-value
Change From Baseline to 12 months				
Unstimulated saliva flow rate, ml/min				
Stimulated saliva flow rate, ml/min				
XQ-summary score (0-100)				
<i>EORCT QLQ-H&N35 (score 0-100):</i>				
HNDR				
HNSS				
HNSW				

Values will be least squares means with standard errors for each group; the difference between these will be reported with 95% confidence intervals (95% CIs). Estimates will be derived from the repeated measures, mixed effects models.

References

1. Lynggaard CD, Grønhøj C, Jensen SB, et al. Long-term Safety of Treatment with Autologous Mesenchymal Stem Cells in Patients with Radiation-Induced Xerostomia: Primary Results of the MESRIX Phase I/II Randomized Trial. *Clin Cancer Res.* 2022;28(13):2890-2897. doi:10.1158/1078-0432.CCR-21-4520
2. Grønhøj C, Jensen DH, Vester-Glowinski P, et al. Safety and Efficacy of Mesenchymal Stem Cells for Radiation-Induced Xerostomia: A Randomized, Placebo-Controlled Phase 1/2 Trial (MESRIX). *Int J Radiat Oncol Biol Phys.* 2018;101(3):581-592. doi:10.1016/j.ijrobp.2018.02.034
3. Lynggaard CD, Grønhøj C, Christensen R, et al. Intraglandular Off-the-Shelf Allogeneic Mesenchymal Stem Cell Treatment in Patients with Radiation-Induced Xerostomia: A Safety Study (MESRIX-II). *Stem Cells Transl Med.* 2022;11(5):478-489. doi:10.1093/stcltm/szac011
4. Jakobsen K, Carlander ALF, Todsén T, et al. Mesenchymal Stem/Stromal Cell Therapy for Radiation-Induced Xerostomia in Previous Head and Neck Cancer Patients: A Phase 2 Randomised, Placebo-Controlled Trial. *Clin Cancer Res.* Published online March 2024. doi:10.1158/1078-0432.CCR-23-3675
5. Jakobsen KK, Carlander ALF, Grønhøj C, et al. Effectiveness and safety of mesenchymal stem/stromal cell for radiation-induced hyposalivation and xerostomia in previous head and neck cancer patients (MESRIX-III): a study protocol for a single-centre, double-blinded, randomised, placebo-controlled, pha. *Trials.* 2023;24(1):1-9. doi:10.1186/s13063-023-07594-5

Supplementary 6. Study Representativeness Table.

Cancer type(s)/subtype(s)/stage(s)/condition	Head and Neck Cancer
Considerations related to:	
Sex	In Demark, overall, 65% of patients with head and neck cancer are men, and up to 75% in specific subtypes such as oropharyngeal cancer (OPSCC), but the incidence in women is rising. Men have a poorer survival than women with a relative 5-year overall survival of 67 % (95% CI 64-70%) compared to 62 % (95% CI 59-65%) in female in the period from 2017-2021.
Age	Most patients (60%) with head and neck cancer are over 60 years old at diagnosis and a mean age of 62.6 years (95% CI 62.3 to 62.8) Patients with head and neck cancer driven by human papillomavirus (HPV) are usually younger, however an increasing incidence among elderly have been observed.
Race/ethnicity	The incidence of head and neck cancer differ by ethnicity, which is also associated with overall survival. It has been shown that black race is associated with a worse survival, but the mechanisms are poorly understood. Differences in socioeconomic status, clinical and treatment-related factors might contribute to the observed differences.
Geography	Globally, more than 900,000 get head and cancer each year and with an increasing incidence. The rise in head and neck cancer is primarily driven by HPV-associated oropharyngeal cancer. The HPV prevalence vary greatly among diverse geographical areas, and the highest prevalences are observed in Western countries. In Denmark, approximately 65% of oropharyngeal cancers are HPV-positive.
Other considerations	Head and neck cancer is a heterogenous group of cancers, but is most often squamous cell carcinomas arising from the mucosal

	<p>epithelium. Historically, head and neck cancers are associated with alcohol and tobacco consumption, but for the last decades infection with HPV has been associated with oropharyngeal cancer, especially in the palatine tonsils and base of the tongue.</p> <p>Most head and neck cancers are treated with radiotherapy with or without concomitant chemotherapy and/or surgery, most often multimodal. The overall survival is best for HPV-positive oropharyngeal cancer, with a 5-year overall survival of 81 % (95% CI 80-83%) compared to 62% (95% CI 59-65%) for all head and neck cancers.</p>
<p>Overall representativeness of this study</p>	<p>The median age in this study population was 61 years, which corresponds to the mean age in the general head and neck cancer population. Also, most patients in this study population were males (73%) which align with the sex distribution in general ranging from 65-75% depending on anatomical subsite and HPV-status.</p> <p>Most patients had oropharyngeal cancer (87%), with the majority being HPV-positive. With higher survival rates, these patients have more time to experience treatment-related side effects, potentially leading to a prolonged burden of these adverse effects throughout their lives.</p>

References

1. Jakobsen K, Carlander ALF, Todsén T, et al. Mesenchymal Stem/Stromal Cell Therapy for Radiation-Induced Xerostomia in Previous Head and Neck Cancer Patients: A Phase 2 Randomised, Placebo-Controlled Trial. *Clin Cancer Res*. Published online March 2024. doi:10.1158/1078-0432.CCR-23-3675
2. Kronberg Jakobsen K, Fenger Carlander AL, Todsén T, et al. Mesenchymal Stem/Stromal Cell Therapy for Radiation-Induced Xerostomia in Previous Head and Neck Cancer Patients: A Phase 2 Randomised, Placebo-Controlled Trial. *Clinical Cancer Research*. 2024;accepted.
3. Lynggaard CD, Grønhøj C, Jensen SB, et al. Long-term Safety of Treatment with Autologous Mesenchymal Stem Cells in Patients with Radiation-Induced Xerostomia: Primary Results of the MESRIX Phase I/II Randomized Trial. *Clin Cancer Res*. 2022;28(13):2890-2897. doi:10.1158/1078-0432.CCR-21-4520
4. Kronberg Jakobsen K, Duch Lynggaard C, Paaske N, et al. Long-Term Outcome Following Treatment with Allogeneic Mesenchymal stem/Stromal Cells for Radiation-Induced Hyposalivation and Xerostomia. *Stem Cells Transl Med*. Published online 2023.
5. Grønhøj C, Jensen DH, Vester-Glowinski P, et al. Safety and Efficacy of Mesenchymal Stem Cells for Radiation-Induced Xerostomia: A Randomized, Placebo-Controlled Phase 1/2 Trial (MESRIX). *Int J Radiat Oncol Biol Phys*. 2018;101(3):581-592. doi:10.1016/j.ijrobp.2018.02.034
6. Lynggaard CD, Grønhøj C, Christensen R, et al. Intraglandular Off-the-Shelf Allogeneic Mesenchymal Stem Cell Treatment in Patients with Radiation-Induced Xerostomia: A Safety Study (MESRIX-II). *Stem Cells Transl Med*. 2022;11(5):478-489. doi:10.1093/stcltm/szac011

Title

No Changes in the Salivary Proteome Composition Detected After Mesenchymal Stem Cell Therapy for Radiation-Induced Hyposalivation in Head and Neck Cancer Patients: A Randomized, Phase 2 Trial

Authors

Amanda-Louise Fenger Carlander¹, Kathrine Kronberg Jakobsen¹, Rosa Jersie-Christensen², Marcus Kildegaard Nielsen², Jens Kastrup³, Daniel Belstrøm⁴, Charlotte Duch Lynggaard, Christian Grønhøj¹ and Christian von Buchwald¹

Affiliation

¹Department of Otorhinolaryngology, Head and Neck Surgery & Audiology, Rigshospitalet – Copenhagen University Hospital, Copenhagen, Denmark

²Department of Science and Environment, Roskilde University, Roskilde, Denmark

³Cardiology Stem Cell Centre, The Heart Centre, Rigshospitalet, Copenhagen, Denmark

⁴Department of Odontology, Section for Clinical Oral Microbiology, Faculty of Health and Medical Sciences, University of Copenhagen, Copenhagen, Denmark

Address for correspondence

Amanda-Louise Fenger Carlander, MD, Ph.D.-fellow

Department of Otorhinolaryngology, Head and Neck Surgery and Audiology, Rigshospitalet

Tel: 00 45 30 22 31 02, e-mail: amanda-louise.fenger.carlander@regionh.dk

Conflict of interest

Lynggaard, Grønhøj, and von Buchwald are co-inventors on patent application for “Stem cell therapy for patients with salivary gland dysfunction”, PCT/EP2020/053878, owned by Rigshospitalet, Copenhagen University Hospital and University of Copenhagen.

Kastrup, Ekblond, and Haack-Sørensen hold a patent application for the investigated stem cell product “Stem cell therapy based on adipose-derived stem cells”, WO2017068140. None of the other co-authors declare conflicts of interest.

Keywords

Mesenchymal stem/stromal cells, mesenchymal stromal cells, MSC, xerostomia, hyposalivation, radiotherapy, proteomics, proteome.

Author contributions

ALFC: Conceptualization, data curation, funding acquisition, formal analysis, investigation, methodology, writing–original draft, project administration. **KKJ:** Conceptualization, data curation, funding acquisition, investigation, methodology, writing–review and editing, project administration **RJC:** Conceptualization, formal analysis, methodology, writing–original draft. **JK:** conceptualization, writing–review and editing. **DB:** Conceptualization, writing–review and editing. **CDL:** Conceptualization, methodology, writing–review and editing. **CG:** Conceptualization, methodology, supervision, investigation, writing–review and editing. **CVB:** Conceptualization, methodology, supervision, investigation, writing–review and editing.

Abstract

Background

Observations indicate that the salivary proteome may change following intraglandular adipose-derived mesenchymal stem cell (ASC) therapy. The aim of this study was to compare the salivary proteome four months following ASC therapy to placebo in previously irradiated head and neck cancer patients from a randomized trial.

Methods

120 patients were randomized 1:1 to receive either ASC therapy or placebo (Crystor10). Unstimulated whole saliva samples were collected at baseline and four months and analyzed with mass spectrometry-based proteomics. The primary endpoint was change in the salivary proteome following ASC therapy compared to placebo at four months. Results were visualized with principal component analysis and volcano plots, and significance was evaluated with a double-sided t-test.

Results

In total, 178 salivary samples from 89 previously irradiated head and neck cancer patients were collected with a total of 1510 proteins identified. In the ASC group, 1438 different proteins (mean 890 per sample; range: 532- 1099) were identified compared to 1432 different proteins in the placebo group (mean of 900 per sample; range: 653-1067). No significantly differentially expressed proteins were identified between the groups at four months. Several proteins upregulated in healthy saliva were insignificantly enriched in the ASC group compared to placebo.

Conclusions

We detected no significant differences in the salivary proteome four months following intraglandular ASC therapy compared to placebo in irradiated patients. While several proteins associated with healthy saliva were non-significantly enriched in the ASC therapy group, their levels remained below those observed in healthy individuals, suggesting a partial effect in maintaining salivary health.

Plain language summary

Dry mouth syndrome and reduced saliva production are the most common side effects following radiotherapy in the head and neck area, and no successful treatment options are available. Injections with stem cells in the salivary glands have shown a potential to restore saliva production and induce changes in the salivary protein composition related to regeneration. We compared the salivary protein composition following stem cell treatment to placebo in irradiated head and neck cancer patients. The results showed no significant change in the protein composition after stem cell treatment, though there was a slight increase in healthy salivary proteins, suggesting partial restoration.

Introduction

Worldwide 900,000 patients are diagnosed with head and neck cancer every year, and most patients undergo radiation-based therapy^{1,2}. Despite, that intensity-modulated radiation therapy aims to reduce toxicity to healthy tissue, the salivary glands are often damaged following radiation therapy (RT)³⁻⁵. RT induces complex damages to the salivary glands encompassing inflammation, loss of salivary-producing acinar and progenitor cells, and fibrosis^{6,7}. This may ultimately lead to salivary gland hypofunction and reduced salivary production, or dry mouth syndrome, xerostomia^{8,9}. Changes in the saliva composition are further observed with alterations in the salivary proteins and protein abundance⁹⁻¹². Consequently, RT-induced salivary gland hypofunction reduces overall oral health with increased risk of oral infections, impairs speech and swallowing, and impacts the overall quality of life in previous head and neck cancer patients^{13,14}. There exist no available disease-modifying treatment strategies for hyposalivation, emphasizing the need for more treatment possibilities¹⁵⁻¹⁷.

Mesenchymal stem cells (MSCs) possess various supportive and regenerative functions rendering them a potential therapeutic agent for the repair of salivary gland damage caused by RT^{18,19}. Since MSCs can easily be isolated from various tissues including adipose tissue they offer a simply available, off-the-shelf, cell-based therapy for clinical use^{20,21}.

We have demonstrated that intraglandular adipose-derived mesenchymal stromal cells (ASCs) have the potential to enhance the salivary flow rate and improve the quality-of-life (QoL) in patients with previous head and neck cancer suffering from radiation-induced hyposalivation²²⁻²⁴, although a superior effect of ACSs compared to placebo was not established in our latest study²².

The mode of action of intraglandular ASC therapy remains unclear, but in our previous MESRIX-II study, we found that intraglandular ASC therapy modified the stimulated salivary proteome, leading to an increase in proteins linked to tissue regeneration, although not restored to normal after four months²⁵. However, this has not been validated in a larger clinical trial.

Therefore, the aim of this study was to evaluate the change in the salivary proteome in unstimulated whole saliva (UWS) four months after intraglandular allogeneic ASC therapy compared to placebo in previous head and neck cancer patients with radiation-induced salivary gland hypofunction in a randomized, double-blinded, placebo-controlled trial. Secondary aims were to 1) evaluate the changes in the salivary proteome in the group receiving ASCs from baseline to four months 2) evaluate the change in the salivary proteome composition in ASC-treated subgroups associated with improved clinical effect of ASC therapy and 3) characterize the salivary proteome following RT at baseline in both groups compared to healthy controls. Our study did not demonstrate a change in the composition of salivary proteome following intraglandular ASC therapy compared to placebo in irradiated head and neck cancer patients. Although insignificant, we did observe a slight enrichment in important salivary proteins in the ASC group which are also upregulated in the healthy salivary proteome, indicating a partial saliva restoration.

Method and materials

Details on the study design, setting, population, sample size, randomization, blinding, and intervention were previously described in detail^{22,26}. Additionally, stimulated saliva samples from ten healthy controls were obtained from our previous study²². Analysis of the salivary proteome was recorded as a pre-specified secondary endpoint, and a study protocol was published²⁷.

Study design

The study was a sub-study to the randomized, placebo-controlled, double-blinded trial investigating the effect on salivary gland function intraglandular ASCs treatment for radiation-induced hyposalivation in previous head and neck cancer patients^{22,26}. The study was approved by the Danish Data Protection Agency (protocol number P-2020-1164), the National Ethics Committee protocol number: (Protocol number: 1802872), and the Danish Medical Agency (EudraCT: 2018-000348-24) and registered at the ClinicalTrials.gov database (NCT04776538)^{22,26}.

The study was conducted in coherence with the protocol and complied with the Declaration of Helsinki and monitored by the Good Clinical Practice (GCP) unit at the University of Copenhagen. Written informed consent was obtained from all study participants prior to enrolment. The study followed the Consolidated Standards of Reporting Trials (CONSORT) reporting guidelines for randomized clinical trials²⁸.

Study setting and participants

The study was conducted at the Department of Otorhinolaryngology, Head and Neck Surgery and Audiology, Rigshospitalet, Denmark, as previously described in detail^{22,26}.

Eligible patients were previously irradiated head and neck cancer patients with clinically observed hyposalivation of UWS 0.05-0.25 ml/min and with two years without recurrence, between 18-75 years, without xerogenic medicine, and informed consent. Patients were excluded if they had other cancers in the last four years, other salivary gland disease, were pregnant or breastfeeding, were smokers, or had an alcohol abuse^{22,26}.

Sample size, randomization, and blinding

Based on our previous study, 120 patients were included^{22,26}. The sample size was developed for the primary endpoint in the main study, which was unstimulated salivary flow rate. Allocation of intervention and blinding were previously described in detail^{22,26}. The follow-up at 4 months was kept blinded to participants and all study personnel involved in both data collection and data analysis.

Intervention

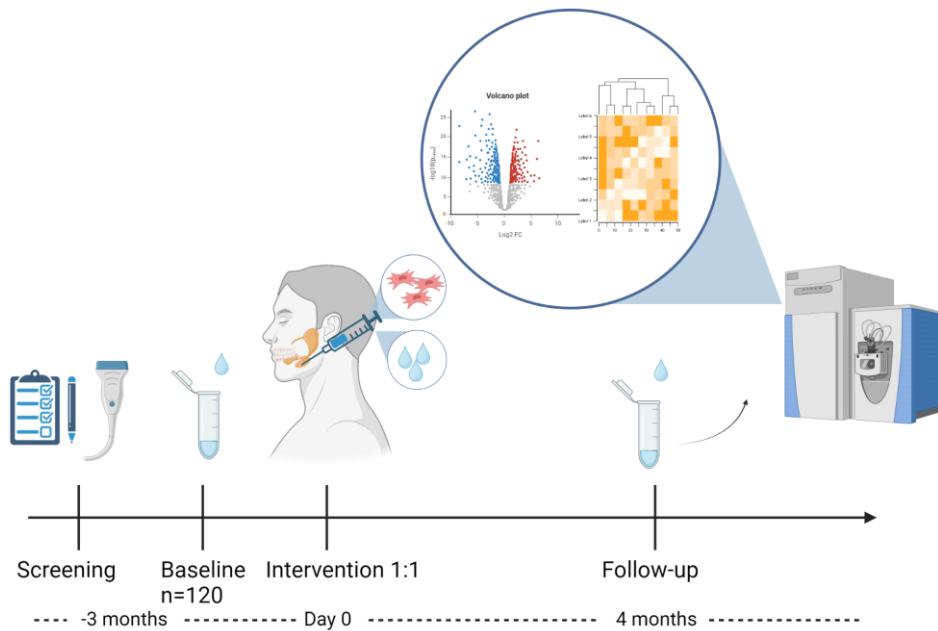
Patients received intraglandular ASC therapy consisting of 0.5 mL of ASCs (25×10^6 ASCs per gland) or placebo, which was CryoStor10 (BiolifeSolutions) containing 10% dimethyl sulfoxide (DMSO). The injections were performed ultrasound-guided by free hand into the submandibular glands. Details on the interventions are previously described in detail^{22,26}.

Saliva collection

UWS was collected by sialometry using the spitting method over 10 minutes at baseline and at four months after intervention. The sialometry was performed at the same time of the day at subsequent collections throughout the study. UWS was collected for 10 min. The saliva samples were immediately stored on dry

ice and afterward stored in Cryotubes at -80 °C until analysis. Further details on the sialometry are previously described in detail^{22,26}. See study flow in **Figure 1**.

Figure 1. Study flow. A total of 120 patients were included and randomized 1:1 to receive either intraglandular ASC therapy or placebo. UWS samples were collected at baseline and four months after intervention and subsequently underwent proteomic analysis using LC-MS/MS.



*Created with Biorender.

Abbreviations: ASC, adipose-derived mesenchymal stem cell; LC-MS/MS, nanoscale liquid chromatography-tandem mass spectrometry; UWS, unstimulated whole saliva.

Sample preparation

Sample preparation was modified from https://doi.org/10.1007/978-1-4939-3049-4_17 and is previously described²⁷. Briefly, 500 μ L UWS was lysed with 500 μ L 95 °C warm lysis buffer (8 M Guanidinium hydrochloride (GuHCl), 5 mM tris(2-carboxyethyl) phosphine (TCEP), 10 mM chloro-acetamide (CAA), 100 mM Tris-HCl pH 8.5) and heated for 10 min. at 95 °C followed by sonication. Subsequently, 200 μ g protein from each sample was diluted with 25mM Tris-HCL (pH 8.5, 5x) and digested with Trypsin (MS grade; Thermo) overnight at 37 °C in a 1:50 w/w ratio. To stop digestion, 10% trifluoroacetic acid (TFA) was added to a final concentration of 1%. Sep-Pak C18 (Waters) with activation by 100% acetonitrile (ACN) and equilibration with 0.1% TFA was used to desalt and concentrate peptides. Samples were then washed with 0.1% TFA and eluted with 40% ACN followed by 60% ACN. Lastly, peptides were dried in a SpeedVac for 1 h at 60 °C and resuspended in 50 μ L 5% ACN in 0.1% TFA.

Mass spectrometry analysis

The mass spectrometry analysis is previously described²⁷. In short, approximately 1 µg peptide solution was analyzed by online nanoscale liquid chromatography-tandem mass spectrometry (LC-MS/MS). Peptides were separated on a 50 cm C18-column (Thermo EasySpray ES804A) using an Ultimate 3000 system (Thermo Scientific) at 40 °C. A gradient flow rate of 250 nl/min was used, transitioning from 1% to 40% MS buffer B over 80 minutes, followed by a 10-minute step to 95%, a 5-minute hold, and then re-equilibration at 5% for 10 minutes. MS buffer A consisting of 0.1% FA and MS buffer B of 90% ACN, 0.1% FA. The Q Exactive Plus instrument (Thermo Scientific) operated in data-dependent acquisition mode with a top 20 Higher-energy Collisional Dissociation (HCD)-MS/MS method (scan range 375–1500 m/z, full-scan resolution 70,000 m/z, AGC target of 3e6, maximum injection time (IT) of 15 ms). Peptides were fragmented with a normalized collision energy of 30, dynamic exclusion of 10 s, excluding unassigned ions, and those with a charge state of 1, 6–8. MS/MS resolution was set at 17,500 m/z, with an AGC target of 1e5 and a maximum IT of 45 ms.

Data analysis

Data analysis is previously described²⁷. The raw LC-MS/MS data files were processed with MaxQuant version 2.1.4.0 using default settings and label-free quantification. In brief, variable modifications included oxidation (M), protein N-termini acetylation, and met-loss, with cysteine carbamidomethyl as a static modification. A 1% false discovery rate and match between runs were applied. MaxQuant output was analyzed with Perseus version 1.6.5.0, excluding proteins with fewer than two peptides for quality assurance. Data was searched against reviewed human database from Uniprot downloaded May 2019, containing 73911 entries, uniprot.org, <https://doi.org/10.1093/nar/gkac1052>. PCA plots were done with proteins identified in 50% of all the relevant samples, and with imputation from normal distribution. Volcano plots were created with proteins identified in 50% of all the relevant samples, and double-sided t-test. Salivary proteins that we previously identified to be upregulated in healthy saliva (Cystatin-D, -D, -SA, -SN, Glutaredoxin-1, Histatin-1, Llpocalin-1, Statherin) was identified as an indicator of repairment²⁵. Enriched biological terms were analyzed in the Database of Annotation, Visualization and Integrated Discovery (DAVID) 2021 version v2023q4. Functional annotation included Gene Ontology Biological Process (GO:BP) using cluster analysis. Enriched clusters were analyzed in a descriptive manner and compared to salivary proteomic profiles from healthy controls obtained in our previous study²⁵. A non-responder analysis was performed to compare included and excluded patients in key baseline characteristics (intervention, age, gender, smoking status, head and neck cancer anatomical location, early stage I-II, mean radiation dose to the four large salivary glands, and duration since radiotherapy). Categorical variables were evaluated for significance with Person's Chi Square test, while continuous data were evaluated using *t-test*. The statistics were performed in R statistics version 4.1.3.

Results

Study characteristics and clinical outcome

The results of the clinical characteristics and clinical outcome are previously described²². Briefly, no differences were observed between the group receiving ASC therapy and placebo in baseline characteristics²². Median age was 61 years (range: 43-75 years) with the majority being male (73%) and most patients had an HPV-positive/p16-positive oropharyngeal cancer (78%). Most patients were diagnosed at early stages (I/II: 74%) and received multimodality treatment (chemoradiotherapy: 85%) with a mean duration since RT of 5.7 years.

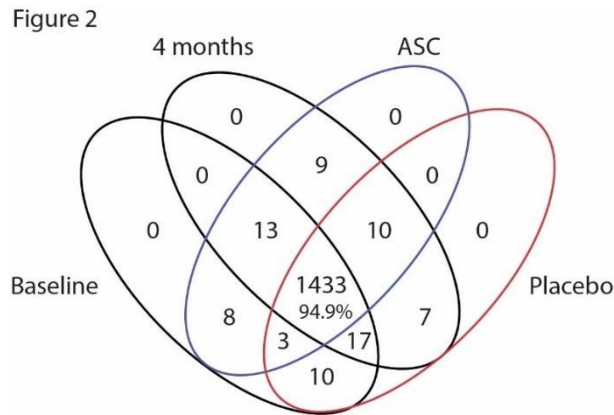
As previously published, the study revealed no significant clinical effect on the UWS in the ASC group compared to placebo, but an increase within the ASC group was observed²². Sub-analyses revealed that smoking, higher mean radiation dose to the four major salivary glands, and development of donor-specific antibodies (DSAs) were associated with less improvement in UWS.

Protein expression

In total, 238 samples of UWS from 119 previously RT head and neck cancer patients (one per patient at baseline and four-month follow-up) were collected. One patient had an UWS of 0 mL/min at the four-month follow-up visit and was therefore excluded from the analysis. Two outliers were removed from the dataset. Due to inconsistent sample preparation, additionally 28 patients were excluded from the dataset. This resulted in a dataset consisting of 89 patients. A non-responder analysis revealed that excluded patients were significantly more smokers than included patients but were comparable on all other baseline characteristics, data not shown.

Across all samples (n=178), 1510 proteins were identified. From the baseline samples, 1484 different proteins were identified with a mean number of 889 per sample (range: 523-1165). At four months after the intervention, 1438 different proteins were identified with a mean number of 890 per sample (range: 532- 1099) in the ASC group, compared to 1432 different proteins were identified with a mean number of 900 per sample (range: 653-1067) in the placebo group. See **Figure 2**.

Figure 2. Venn diagram of identified proteins in both the ASC group and in the placebo group at four months. In total, 1510 proteins were identified across all samples. Most proteins were shared between the groups (1433, 95%).



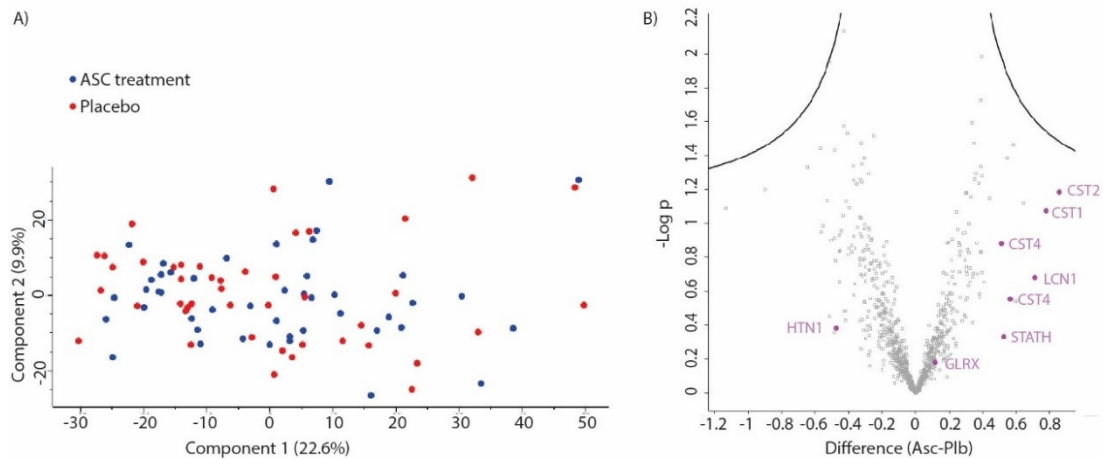
Abbreviations: ASC, adipose-derived mesenchymal stem cell.

The salivary proteome following ASC therapy compared to placebo

The PCA shows no clear separation of samples from patients receiving ASC therapy compared to patients receiving placebo at four months following the intervention. See **Figure 3A**. The Volcano plot in **Figure 3B** illustrates the distribution of the proteins in the ASC group compared to the placebo group and shows no significantly differentially expressed proteins in the two groups. However, proteins that we previously identified to be upregulated in saliva samples from healthy controls²⁵, trended to be enriched in the ASC group compared to the placebo group, see **Figure 3B**. These proteins included Cystatin-S, Cystatin-D, Cystatin-SA, Cystatin-SN, Glutaredoxin-1, and Lipocalin-1, while Histatin-1 was downregulated in the ASC group. See **Table 2**.

Figure 3. Comparison of patients receiving ASC therapy compared to placebo at four months. A. PCA demonstrating no clear separation. **B.** Volcano plot illustrating no differentially expressed proteins. Proteins that trended to be significantly differentially expressed between the ASC group and the placebo group are highlighted.

Figure 3



Purple circle = marked proteins associated with healthy saliva.

Abbreviations: ASC, adipose-derived mesenchymal stem cell; PCA, principal component analysis.

Table 2. Proteins upregulated in healthy saliva and at four months in patients receiving ASC therapy trending to be differentially expressed compared to placebo.

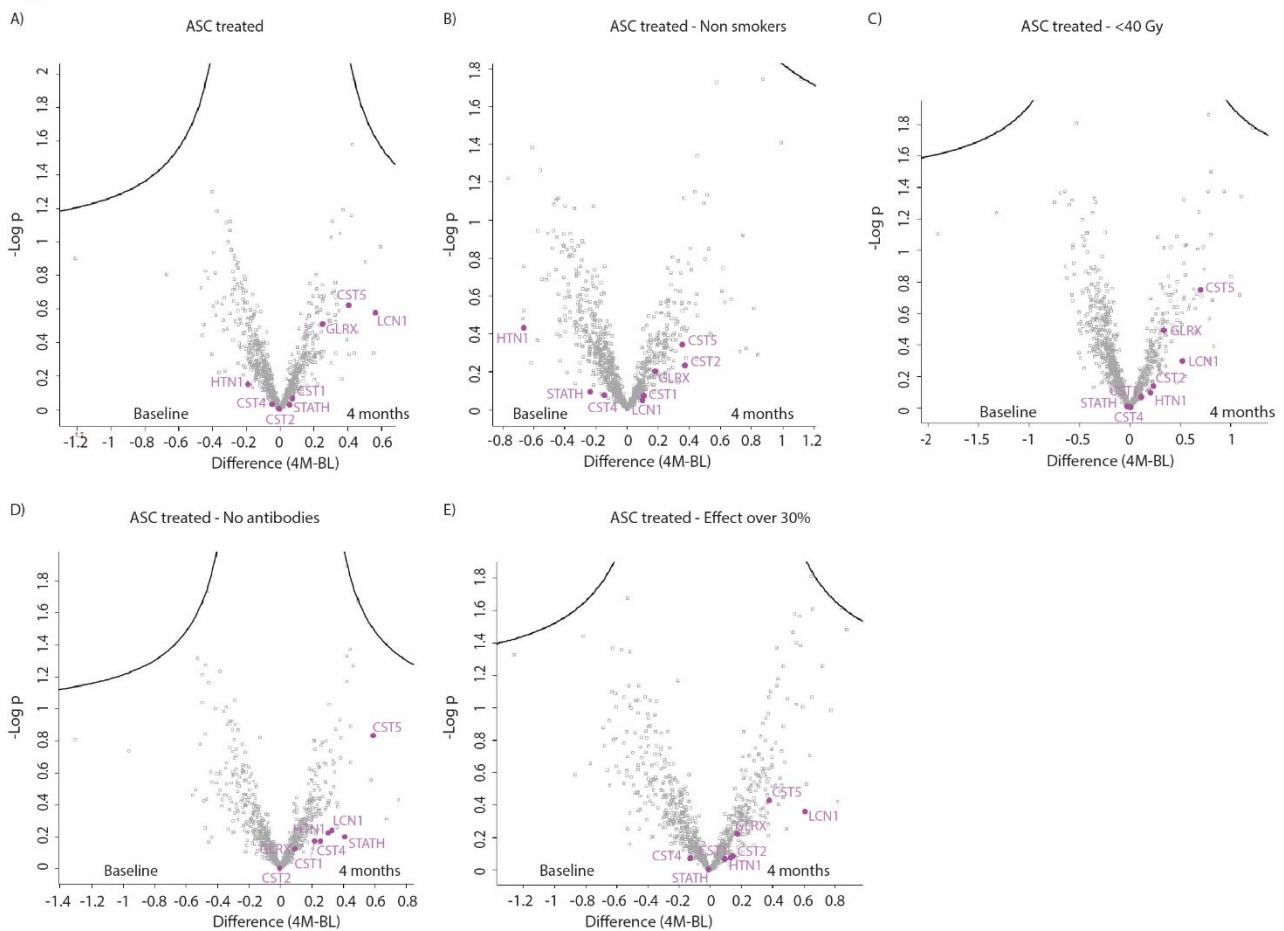
Accession	Gene name	Proteins with higher or lower intensity in the ASC group vs placebo	-LOG (P value)	Difference
P28325	CST5	Cystatin-D	0.883	0.512
P01036	CST4	Cystatin-S	0.555	0.566
P09228	CST2	Cystatin-SA	1.187	0.857
P01037	CST1	Cystatin-SN	1.077	0.778
P35754	GLRX	Glutaredoxin-1	0.181	0.117
P15515	HTN1	Histatin-1	0.382	-0.475
P31025	LCN1	Lipocalin-1	0.680	0.710
P02808	STATH	Statherin	0.332	0.526

Sub-analysis of the salivary proteome following ASC therapy

Within the ASC group, no differentially expressed proteins were observed following ASC treatment at four months compared to baseline. No differences in the salivary proteins upregulated in healthy saliva were observed either. See **Figure 4A**. While a less increase in the clinical outcome UWS was associated with smoking, higher mean radiation dose, and development of DSAs in the main study²², no differences in the salivary proteome were observed following ASC treatment at four months compared to baseline for never smokers, patients receiving a mean radiation dose the large salivary glands < 40 Gy, or patients who did not develop DSAs, see **Figure 4B-D**. However, proteins that we previously identified to be upregulated in healthy controls²⁵, tended to be enriched in the group who did not develop DSAs at four months compared to the baseline, See **Figure 4D**. These proteins included Cystatin-D, Cystatin-SN, Cystatin-SA, Glutaredoxin-1, Histatin-1 and Lipocalin-1. A difference in the salivary proteome was not observed within the group who experienced a clinical effect on UWS of 30% or more either, see **Figure 4E**.

Figure 4. Comparison of patients receiving ASC therapy at baseline and four months after intervention. A. Volcano plot illustrating no differentially expressed proteins in patients receiving ASC therapy. **B.** Volcano plot illustrating no differentially expressed proteins in patients with no history of smoking and who received ASC therapy. **C.** Volcano plot illustrating no differentially expressed proteins in patients with a mean radiation dose to the four large salivary glands <40 Gy and who received ASC therapy. **D.** Volcano plot illustrating no differentially expressed proteins in patients who did not develop DSAs and who received ASC therapy. **E.** Volcano plot illustrating no differentially expressed proteins in patients with an effect on clinical evaluation of the unstimulated flow rate of 30% or more and who received ASC therapy.

Figure 4



Purples circle = marked proteins associated with healthy saliva.

Abbreviations: ASC, adipose-derived mesenchymal stem cell; BL, baseline; DSAs, donor-specific antibodies; Gy, Gray; M, months.

The salivary proteome in irradiated head and neck cancer patients compared to healthy controls

Descriptive functional annotation analysis of GO:BP revealed that the most enriched clusters in irradiated patients compared to healthy controls were metabolic processes (score 18.9 versus 9.2), immune response

(score 19.7 versus 22.7), oxidative stress and detoxification processes (score 12.5 versus 9.4), injury response (score 8.2 versus 7.3), and antimicrobial response (score 5.5 versus 12.1). See **Table 3**.

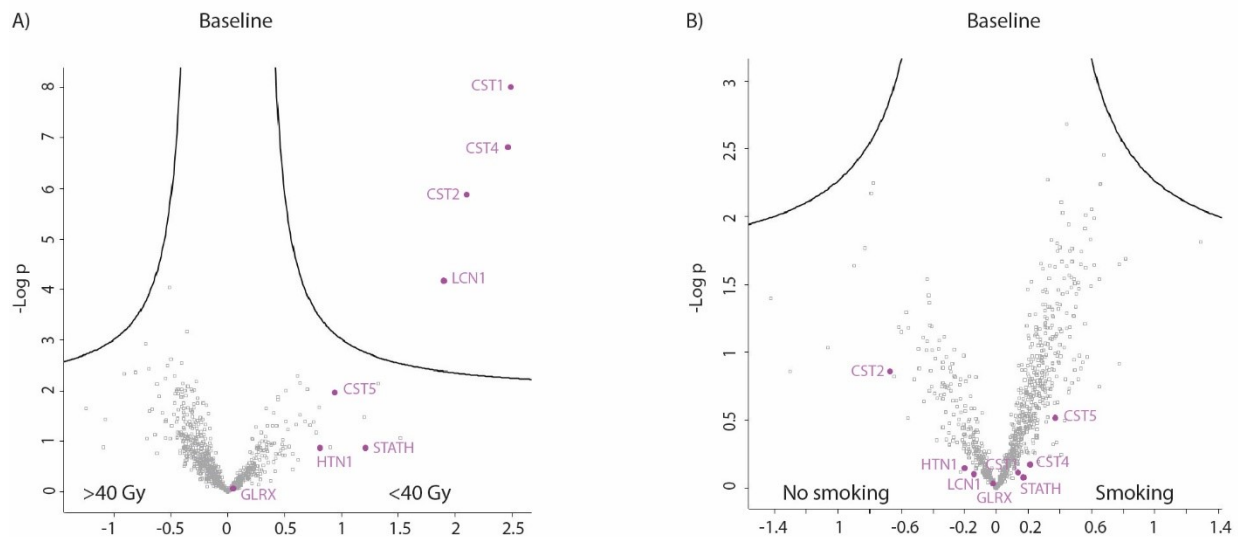
Table 3. List of enriched proteins identified in saliva from irradiated patients and healthy controls.

Gene ontology	Baseline, (n=89)		Healthy controls, (n=10)	
Biological function (level 3)	Enrichment score, count	P value	Enrichment score, count	P value
Metabolic processes	18.9		9.2	
Phosphorus metabolic process	206	5.7E-9	143	3.2E-2
nucleobase-containing small molecule metabolic process	139	1.2E-31	95	4.2E-14
purine-containing compound metabolic process	123	4.7E-31	84	3.0E-14
carbohydrate derivate biosynthetic process	72	3.9E-4	46	3.1E-1
Immune response	19.7		22.7	
Defense response	226	5.4E-23	212	2.5E-25
Innate immune response	129	3.1E-15	123	1.7E-17
response to bacterium	113	2.5E-12	115	3.9E-23
humoral immune response	-		72	2.9E-17
Oxidative stress and detoxification processes	12.5		9.4	
cellular oxidant detoxification	36	1.1E-16	31	7.9E-14
reactive oxygen species metabolic process	36	4.8E-13	28	6.5E-9
Injury response	8.2		7.3	
response to wounding	79	6.0E-12	71	3.0E-11
blood coagulation	39	61.3E-8	34	1.8E-7
regulation of body fluids	60	3.6E-8	49	6.2E-6
Antimicrobial response	5.5		12.1	
response to fungus	19	1.2E-6	21	4.7E-9
killing of cells of another organism	21	5.5E-6	31	1.4E-14

For irradiated patients at baseline, significantly differentially expressed proteins were observed in patients who received a lower mean radiation dose to the four salivary glands (<40 Gy) as compared to those who received a higher dose. Cystatin-SN (difference 2.45, $-\log p$ -value 6.85), Cystatin-S (difference 2.37, $-\log p$ -value 5.54), Cystatin-SA (difference 1.91, $-\log p$ -value 4.36), and Lipocalin-1 (difference 1.77, $-\log p$ -value 3.56) were significantly upregulated in patients with a lower mean radiation dose. See **Figure 5A**. No differences were observed in patients with no history of smoking as compared to smokers. See **Figure 5B**.

Figure 5. Comparison of irradiated patients at baseline. A. Volcano plot illustrating significantly differentially expressed proteins in irradiated patients who received a mean radiation dose to the four large salivary glands < 40 Gy as compared to > 40 Gy. **B.** Volcano plot illustrating no differentially expressed proteins in irradiated patients with no history of smoking as compared to smokers.

Figure 5



Purple circle = significantly upregulated proteins.

Abbreviations: Gy, Gray.

Discussion

This is the largest randomized, placebo-controlled trial investigating the salivary proteome following intraglandular ASC therapy compared to placebo in previous head and neck cancer patients with radiation-induced hyposalivation. We did not find any significant differences in the salivary proteome between patients receiving ASC therapy compared to placebo four months following intervention.

Our results align with the primary endpoint of our study, which was clinical change in UWS, as the increase in UWS in the ASC group was 0.04 mL/min compared to 0.01 mL/min in the placebo group (difference of 0.03 mL/min, $p=0.11$)²². Yet, we previously found significant changes in the salivary proteome following ASC therapy in our pilot study, MESRIX-II, with upregulation of major salivary proteins as well as proteins involved in cell growth, immune system, and regeneration²⁵. However the study was small ($n=10$) and not randomized, while this study included 120 irradiated head and neck cancers and was compared to placebo. We found that patients who received ASC therapy tended to have a salivary proteome more similar to that of healthy saliva compared to placebo, with insignificant upregulation of Cystatin-S, Cystatin-D, Cystatin-SA, Cystatin-SN, Glutaredoxin-1, and Lipocalin-1 which have shown to be significantly upregulated in healthy individuals²⁵. Still, the levels did not reach those observed in healthy saliva as compared to irradiated head and neck cancer patients^{10,25}. Both cystatins, statherins, and histatins are components of the first-line immune defense in the oral cavity, e.g., is cystatin-S among others associated with oral gingivitis and caries while histatins are important antifungal proteins²⁹⁻³¹. Cystatin-D also impacts the subjective feeling of oral dryness, while statherins are important for calculus formation, and Lipocalin-1 is involved in taste reception³¹⁻³³.

Some patients receiving ASC therapy might have a more favorable clinical response to ASC therapy, as less improvement in the clinical outcome UWS was seen in patients with a history of smoking, who received a higher mean radiation dose, and who developed DSAs as shown in our previous study²². Therefore, we hypothesized that changes in the salivary proteome following ASC therapy would be more evident in patients with no history of smoking, who received a lower mean radiation dose (<40 Gy), who did not develop DSAs and with a substantial clinical effect (>30% increase in UWS). However, no differences in the salivary proteome within these subgroups were observed, but in ASC-treated patients who did not develop DSAs, a non-significant upregulation of Cystatin-D, Cystatin-SN, Cystatin-SA, Glutaredoxin-1, Histatin-1 and Lipocalin-1 was observed. This indicates that immunological tolerance without development of DSAs might be important in restoring the saliva composition.

In line with results from our MESRIX-II study²⁵, we found notable differences in the functional annotation of proteins identified in the UWS in irradiated patients compared to healthy individuals. An increased metabolic activity was identified in both groups, emphasizing the metabolic and biosynthetic processes in saliva necessary for e.g., enzyme synthesis. Yet, the metabolic processes were more increased in irradiated patients, which could reflect increased metabolic activity as cells attempt to repair and recover from radiation-induced damage. Also, enrichment in oxidative stress was found possibly due to radiation exposure, which is known to generate reactive oxygen species that lead to oxidative stress³⁴. Both groups had an enriched immune and injury response, which is expected as saliva plays a crucial role in defending against pathogens³⁵. Hynne et al., also identified differentially expressed salivary proteins involved in inflammation, host cell injury, oxidative stress response activation, and tissue restoration in irradiated patients compared to healthy controls¹⁰. A recent prospective study also identified increased levels of inflammatory protein markers following RT as compared to before RT¹². Interestingly we found that patients who received a mean radiation dose to the four large salivary glands below 40 Gy had significantly increased levels of Cystatin-SN (more than twofold) Cystatin-S (more than twofold), Cystatin-SA and Liocalin-1. All four proteins are associated with a healthy salivary proteome^{11,25}, and are involved in maintenance of the overall oral health and taste reception^{31,33}. This emphasizes how crucial it is to administer RT while minimizing dose and/or protecting the salivary glands when feasible to protect not only salivary flow rates but also salivary quality^{5,36}.

Several limitations of this study need to be addressed. Not all samples were included in the proteomic profiling due to inconsistent sample preparation, and significantly more of the excluded patients were smokers. Smoking was significantly associated with a decrease in UWS²², and could potentially impact the salivary proteome as well. Second, we found substantial interpersonal differences already present at baseline, making it difficult to identify small changes induced by ASC therapy. Additionally, we compared the unstimulated salivary proteome in irradiated patients with the stimulated salivary proteome in healthy controls. Unstimulated and stimulated saliva differ in essential aspects^{37,38}; which might contribute to the differences observed in this study. We did not evaluate or account for the oral status at baseline which also influences the salivary proteome^{39,40}. Lastly, the study sample size was powered to detect a clinical change in salivary flow rate and not changes in the salivary proteome composition²⁶.

As the salivary proteins levels displayed great interpersonal heterogeneity already present at baseline, the future of intraglandular ASC therapy might be reserved for a smaller group of head and neck cancer patients, and future studies should be powered to investigate patient-specific factors. Early intervention

following RT might also enhance the effect of ASC therapy, since the salivary gland damage may be more reversible at this stage. Also, repeated ASC treatment could be beneficial as demonstrated in e.g., neurological disease^{41,42}.

In conclusion, we found no significant differences in the salivary proteome four months following intraglandular ASC therapy in previous head and neck cancer patients with radiation-induced hyposalivation compared to placebo. However, while several proteins upregulated in healthy saliva were non-significantly enriched in the ASC therapy group, their levels remained below those observed in healthy individuals. This suggests that ASC may have a partial effect on maintaining salivary health.

References

1. Sung H, Ferlay J, Siegel RL, et al. Global Cancer Statistics 2020: GLOBOCAN Estimates of Incidence and Mortality Worldwide for 36 Cancers in 185 Countries. *CA Cancer J Clin.* 2021;71(3):209-249. doi:10.3322/caac.21660
2. Borrás JM, Barton M, Grau C, et al. The impact of cancer incidence and stage on optimal utilization of radiotherapy: Methodology of a population based analysis by the ESTRO-HERO project. *Radiotherapy and Oncology.* 2015;116(1):45-50. doi:10.1016/j.radonc.2015.04.021
3. Langendijk JA, Doornaert P, Verdonck-de Leeuw IM, Leemans CR, Aaronson NK, Slotman BJ. Impact of late treatment-related toxicity on quality of life among patients with head and neck cancer treated with radiotherapy. *J Clin Oncol.* 2008;26(22):3770-3776. doi:10.1200/JCO.2007.14.6647
4. Jensen SB, Pedersen AML, Vissink A, et al. A systematic review of salivary gland hypofunction and xerostomia induced by cancer therapies: prevalence, severity and impact on quality of life. *Support Care Cancer.* 2010;18(8):1039-1060. doi:10.1007/s00520-010-0827-8
5. Nutting CM, Morden JP, Harrington KJ, et al. Parotid-sparing intensity modulated versus conventional radiotherapy in head and neck cancer (PARSPORT): a phase 3 multicentre randomised controlled trial. *Lancet Oncol.* 2011;12(2):127-136. doi:10.1016/S1470-2045(10)70290-4
6. Jasmer KJ, Gilman KE, Muñoz Forti K, Weisman GA, Limesand KH. Radiation-Induced Salivary Gland Dysfunction: Mechanisms, Therapeutics and Future Directions. *J Clin Med.* 2020;9(12). doi:10.3390/jcm9124095
7. Eisbruch A, Ten Haken RK, Kim HM, Marsh LH, Ship JA. Dose, volume, and function relationships in parotid salivary glands following conformal and intensity-modulated irradiation of head and neck cancer. *Int J Radiat Oncol Biol Phys.* 1999;45(3):577-587. doi:10.1016/s0360-3016(99)00247-3
8. Vissink A, Jansma J, Spijkervet FKL, Burlage FR, Coppes RP. Oral sequelae of head and neck radiotherapy. *Crit Rev Oral Biol Med.* 2003;14(3):199-212.
9. Pinna R, Campus G, Cumbo E, Mura I, Milia E. Xerostomia induced by radiotherapy: an overview of the physiopathology, clinical evidence, and management of the oral damage. *Ther Clin Risk Manag.* 2015;11:171-188. doi:10.2147/TCRM.S70652
10. Hynne H, Aqrabi LA, Jensen JL, et al. Proteomic Profiling of Saliva and Tears in Radiated Head and Neck Cancer Patients as Compared to Primary Sjögren's Syndrome Patients. *Int J Mol Sci.* 2022;23(7). doi:10.3390/ijms23073714
11. Laheij AMGA, Rasch CN, Brandt BW, et al. Proteins and peptides in parotid saliva of irradiated patients compared to that of healthy controls using SELDI-TOF-MS Oral Health. *BMC Res Notes.* 2015;8(1):1-7. doi:10.1186/s13104-015-1641-7
12. Brandt E, Keskin M, Räisänen IT, et al. Induction of Collagenolytic MMP-8 and -9 Tissue Destruction Cascade in Mouth by Head and Neck Cancer Radiotherapy: A Cohort Study. *Biomedicines.* 2023;12(1). doi:10.3390/biomedicines12010027
13. Jellema AP, Slotman BJ, Doornaert P, Leemans CR, Langendijk JA. Impact of radiation-induced xerostomia on quality of life after primary radiotherapy among patients with head and neck cancer. *Int J Radiat Oncol Biol Phys.* 2007;69(3):751-760. doi:10.1016/j.ijrobp.2007.04.021

14. Høxbroe Michaelsen S, Grønhøj C, Høxbroe Michaelsen J, Friberg J, von Buchwald C. Quality of life in survivors of oropharyngeal cancer: A systematic review and meta-analysis of 1366 patients. *Eur J Cancer*. 2017;78:91-102. doi:10.1016/j.ejca.2017.03.006
15. Jensen DH, Oliveri RS, Trojahn Kølle SF, et al. Mesenchymal stem cell therapy for salivary gland dysfunction and xerostomia: a systematic review of preclinical studies. *Oral Surg Oral Med Oral Pathol Oral Radiol*. 2014;117(3):335-342.e1. doi:10.1016/j.oooo.2013.11.496
16. Riley P, Glenny AM, Hua F, Worthington H V. Pharmacological interventions for preventing dry mouth and salivary gland dysfunction following radiotherapy. *Cochrane Database Syst Rev*. 2017;7(7):CD012744. doi:10.1002/14651858.CD012744
17. Jensen SB, Vissink A, Limesand KH, Reyland ME. Salivary Gland Hypofunction and Xerostomia in Head and Neck Radiation Patients. *J Natl Cancer Inst Monogr*. 2019;2019(53). doi:10.1093/jncimonographs/lgz016
18. Singer NG, Caplan AI. Mesenchymal Stem Cells: Mechanisms of Inflammation. *Annu Rev Pathol Mech Dis*. 2011;6:457-478. doi:10.1146/annurev-pathol-011110-130230
19. Jaguar GC, Prado JD, Campanhã D, Alves FA. Clinical features and preventive therapies of radiation-induced xerostomia in head and neck cancer patient: a literature review. *Applied Cancer Research*. 2017;37(1):1-8. doi:10.1186/s41241-017-0037-5
20. Zuk PA, Zhu M, Mizuno H, et al. Multilineage cells from human adipose tissue: implications for cell-based therapies. *Tissue Eng*. 2001;7(2):211-228. doi:10.1089/107632701300062859
21. Krampera M, Le Blanc K. Mesenchymal stromal cells: Putative microenvironmental modulators become cell therapy. *Cell Stem Cell*. 2021;28(10):1708-1725. doi:10.1016/j.stem.2021.09.006
22. Jakobsen K, Carlander ALF, Todsen T, et al. Mesenchymal Stem/Stromal Cell Therapy for Radiation-Induced Xerostomia in Previous Head and Neck Cancer Patients: A Phase 2 Randomised, Placebo-Controlled Trial. *Clin Cancer Res*. Published online March 2024. doi:10.1158/1078-0432.CCR-23-3675
23. Lynggaard CD, Grønhøj C, Christensen R, et al. Intraglandular Off-the-Shelf Allogeneic Mesenchymal Stem Cell Treatment in Patients with Radiation-Induced Xerostomia: A Safety Study (MESRIX-II). *Stem Cells Transl Med*. 2022;11(5):478-489. doi:10.1093/stcltm/szac011
24. Grønhøj C, Jensen DH, Vester-Glowinski P, et al. Safety and Efficacy of Mesenchymal Stem Cells for Radiation-Induced Xerostomia: A Randomized, Placebo-Controlled Phase 1/2 Trial (MESRIX). *Int J Radiat Oncol Biol Phys*. 2018;101(3):581-592. doi:10.1016/j.ijrobp.2018.02.034
25. Lynggaard CD, Jersie-Christensen R, Juhl M, et al. Intraglandular mesenchymal stem cell treatment induces changes in the salivary proteome of irradiated patients. *Communications medicine*. 2022;2(1):160. doi:10.1038/s43856-022-00223-3
26. Jakobsen KK, Carlander ALF, Grønhøj C, et al. Effectiveness and safety of mesenchymal stem/stromal cell for radiation-induced hyposalivation and xerostomia in previous head and neck cancer patients (MESRIX-III): a study protocol for a single-centre, double-blinded, randomised, placebo-controlled, pha. *Trials*. 2023;24(1):1-9. doi:10.1186/s13063-023-07594-5
27. Carlander ALF, Jakobsen KK, Jersie-Christensen R, et al. Exploring the salivary proteome following intraglandular mesenchymal stromal cell therapy for radiation-induced hyposalivation in previous head and neck cancer patients: a secondary study protocol for the MESRIX-III, randomised, controlled trial. *Trials*.:In review.

28. Moher D, Hopewell S, Schulz KF, et al. CONSORT 2010 explanation and elaboration: updated guidelines for reporting parallel group randomised trials. *BMJ*. 2010;340:c869. doi:10.1136/bmj.c869
29. da Silva CVF, Bacila Sade Y, Naressi Scapin SM, da Silva-Boghossian CM, de Oliveira Santos E. Comparative proteomics of saliva of healthy and gingivitis individuals from Rio de Janeiro. *Proteomics Clin Appl*. 2023;17(5):e2200098. doi:10.1002/prca.202200098
30. Koopaie M, Salamati M, Montazeri R, Davoudi M, Kolahdooz S. Salivary cystatin S levels in children with early childhood caries in comparison with caries-free children; statistical analysis and machine learning. *BMC Oral Health*. 2021;21(1):650. doi:10.1186/s12903-021-02016-x
31. Lynge Pedersen AM, Belstrøm D. The role of natural salivary defences in maintaining a healthy oral microbiota. *J Dent*. 2019;80 Suppl 1:S3-S12. doi:10.1016/j.jdent.2018.08.010
32. Yamamoto K, Hiraishi M, Haneoka M, Fujinaka H, Yano Y. Protease inhibitor concentrations in the saliva of individuals experiencing oral dryness. *BMC Oral Health*. 2021;21(1):661. doi:10.1186/s12903-021-02024-x
33. Stolle T, Grondinger F, Dunkel A, Hofmann T. Quantitative proteomics and SWATH-MS to elucidate peri-receptor mechanisms in human salt taste sensitivity. *Food Chem*. 2018;254:95-102. doi:10.1016/j.foodchem.2018.01.160
34. Azzam EI, Jay-Gerin JP, Pain D. Ionizing radiation-induced metabolic oxidative stress and prolonged cell injury. *Cancer Lett*. 2012;327(1-2):48-60. doi:10.1016/j.canlet.2011.12.012
35. Fábíán TK, Hermann P, Beck A, Fejérdy P, Fábíán G. Salivary defense proteins: their network and role in innate and acquired oral immunity. *Int J Mol Sci*. 2012;13(4):4295-4320. doi:10.3390/ijms13044295
36. Steenbakkers RJHM, van Rijn-Dekker MI, Stokman MA, et al. Parotid Gland Stem Cell Sparing Radiation Therapy for Patients With Head and Neck Cancer: A Double-Blind Randomized Controlled Trial. *Int J Radiat Oncol Biol Phys*. 2022;112(2):306-316. doi:10.1016/j.ijrobp.2021.09.023
37. Gomar-Vercher S, Simón-Soro A, Montiel-Company JM, Almerich-Silla JM, Mira A. Stimulated and unstimulated saliva samples have significantly different bacterial profiles. *PLoS One*. 2018;13(6):e0198021. doi:10.1371/journal.pone.0198021
38. Foratori-Junior GA, Le Guennec A, Fidalgo TK da S, et al. Comparison of the Metabolic Profile between Unstimulated and Stimulated Saliva Samples from Pregnant Women with/without Obesity and Periodontitis. *J Pers Med*. 2023;13(7):1-21. doi:10.3390/jpm13071123
39. Hartenbach FARR, Velasquez É, Nogueira FCS, Domont GB, Ferreira E, Colombo APV. Proteomic analysis of whole saliva in chronic periodontitis. *J Proteomics*. 2020;213:103602. doi:10.1016/j.jprot.2019.103602
40. Belstrøm D, Jersie-Christensen RR, Lyon D, et al. Metaproteomics of saliva identifies human protein markers specific for individuals with periodontitis and dental caries compared to orally healthy controls. *PeerJ*. 2016;4:e2433. doi:10.7717/peerj.2433
41. Petrou P, Kassis I, Ginzberg A, et al. Long-Term Clinical and Immunological Effects of Repeated Mesenchymal Stem Cell Injections in Patients With Progressive Forms of Multiple Sclerosis. *Front Neurol*. 2021;12:639315. doi:10.3389/fneur.2021.639315
42. Pan K, Deng L, Chen P, et al. Safety and Feasibility of Repeated Intrathecal Allogeneic Bone Marrow-Derived Mesenchymal Stromal Cells in Patients with Neurological Diseases. *Stem Cells Int*. 2019;2019:8421281. doi:10.1155/2019/8421281